

TWO STEROIDS FROM *CALVATIA CYATHIFORMIS*

NOBUO KAWAHARA,* SETSUKO SEKITA and MOTOYOSHI SATAKE

National Institute of Health Sciences (NIHS), Kamiyoga 1-18-1, Setagaya-ku, Tokyo 158, Japan

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Abstract—Calvasterols A (14 α -hydroxyergosta-4,7,9,22-tetraen-3,6-dione) and B (9 α ,14 α -dihydroxyergosta-4,7,22-trien-3,6-dione), two new steroids, have been isolated from *Calvatia cyathiformis*. Their molecular structures have been defined by spectroscopic means and chemical correlations.

INTRODUCTION

In a previous paper [1], we reported the isolation of two new steroids designated as cyathisterone (3) and cyathisterol (4) from the dichloromethane extract of the fruiting body of *Calvatia cyathiformis* (Bosc.) Morg., a Chinese crude drug ('Ma Bo' in Chinese; 'Mabotsu' in Japanese), used in China as a haemostatic and pharyngodynia [2]. Further investigation of this extract led us to isolate two new steroids designated as calvasterols A (1) and B (2). The structural elucidation of the above compounds (1 and 2) is reported in this paper.

RESULTS AND DISCUSSION

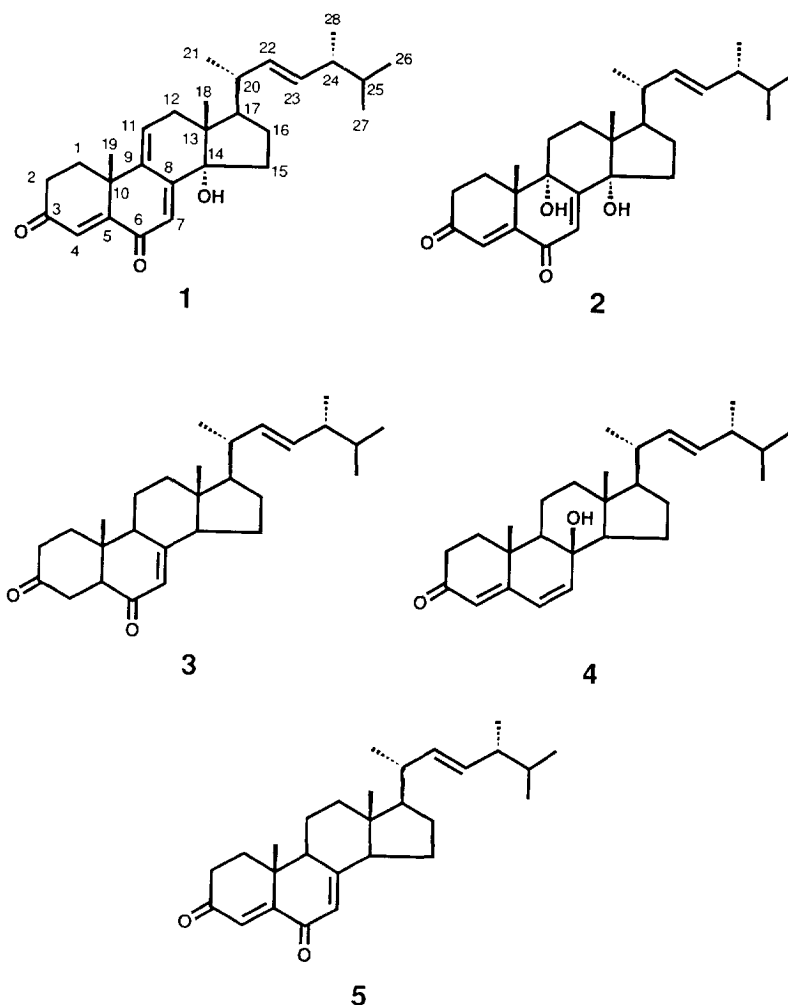
Calvasterol A (1), had the molecular formula $C_{28}H_{38}O_3$ as shown by the HR mass spectrum, which had a molecular ion at m/z 422 $[M]^+$. The IR absorption at 3450 and 1660 cm^{-1} suggested the presence of a hydroxyl and an α,β -unsaturated carbonyl group. The 1H NMR spectrum of 1 clearly showed two tertiary methyl signals at δ 0.76 and 1.46, four secondary methyl signals at δ 0.84, 0.85, 0.93 and 1.04, which suggested an ergostane skeleton [3]. This was supported by the fact that the chemical shift of the signals for the secondary methyl groups together with resonances for a *trans*-disubstituted double bond at δ 5.22 and 5.30 were consistent with those of ergosterol. On the other hand, two olefinic proton signals at δ 6.17 (1H, s) and 6.45 (1H, s) in the 1H NMR spectrum of 1, together with two tertiary carbon signals at δ 121.8 and 127.0, two quaternary carbon signals at δ 156.0 and 156.3 and two carbonyl carbon signals at δ 188.7 and 199.7 observed in the ^{13}C NMR spectrum of 1 (Table 1) were similar to those of ergosta-4,7,22-triene-3,6-dione (5) [1, 4]. Also a negative Cotton effect at 275 nm and a positive Cotton effect at

234 nm, respectively, observed in the CD spectrum of 1 were closely similar to those of 5 (270 nm, negative; 234 nm, positive). All these data indicated that calvasterol A (1) was an ergosta-4,7,22-trien-3,6-dione (5) derivative including its stereochemistry.

An olefinic proton signal at δ 6.22 (1H, dd, $J = 1.8, 6.6$ Hz), which was only coupled with the methylene protons at δ 2.38 (1H, dd, $J = 6.6, 18.3$ Hz) and 2.72 (1H, dd, $J = 1.8, 18.3$ Hz), showed three-bond correlations with three quaternary carbon signals at δ 38.8 (C-10), 46.3 (C-13) and 156.0 (C-8) in the HMBC spectrum of 1 (Table 2). Thus the olefinic proton and methylene protons were assigned at C-11 and C-12, respectively. Also the assignment of a quaternary carbon signal (δ 84.7), which was conjoining a hydroxyl group, was confirmed as C-14. This was supported by the fact that the olefinic proton, which was assigned at C-7, was not coupled with any other protons. Finally, a proton signal at δ 2.72, which was assigned at H-12 α using a NOE correlation between δ 0.76 (H-18) and 2.38 (H-12 β) in the NOESY spectrum of 1, was shifted down-field to δ 3.04 (pyridine- d_5) by the pyridine induced deshielding effect [5] indicating that a hydroxyl group and a proton (OH-14 α and H-12 α) occupied 1,3-*syn*-periplanar positions. Therefore, the configuration at C-14 has been assigned as *S*. These data led to the conclusion that calvasterol A (1) was elucidated as 14 α -hydroxyergosta-4,7,9,22-tetraen-3,6-dione.

Calvasterol B (2), had the molecular formula $C_{28}H_{40}O_4$ since the high resolution mass spectrum gave a molecular ion at m/z 440 $[M]^+$. The IR absorption at 3450 and 1660 cm^{-1} of 2 also suggested the presence of a hydroxyl and α,β -unsaturated carbonyl group. The 1H NMR spectrum of 2 showed two tertiary methyl signals at δ 0.77 and 1.37, four secondary methyl signals at δ 0.84, 0.85, 0.93 and 1.05 and a *trans*-disubstituted double bond signal at δ 5.20 and 5.29. This information revealed the presence of an ergostane skeleton. The 1H and ^{13}C NMR spectra of 2 were similar to that of calvasterol

*Author to whom correspondence should be addressed.



A (**1**) except for the disappearance of an olefinic proton signal at $\delta 6.22$ (1H, *dd*, $J = 1.8, 6.6$ Hz) in the ^1H NMR spectrum of **1** and the appearance of a secondary carbon signal $\delta 27.6$ and a quaternary carbon signal at $\delta 74.4$ in the ^{13}C NMR spectrum of **2** instead of a tertiary carbon signal at $\delta 132.9$ and a quaternary carbon signal at $\delta 138.5$ seen in the spectrum of **1** (Table 1). Thus the structure of **2** was considered to be the dihydroxy derivative of ergosta-4,7,22-trien-3,6-dione (**5**).

Hydroxylation of ergosta-4,7,22-trien-3,6-dione (**5**) was examined to give dihydroxysteroid, which was identical with calvasterol B (**2**), including the optical rotation. The assignment of two quaternary carbon signals ($\delta 74.4$ and 87.0), which were joining a hydroxyl group, were confirmed as C-9 and C-14, respectively, using a HMBSC correlation of **2** (Table 2). This was supported by the fact that the olefinic proton, which was assigned at C-7, was not coupled with any other protons. Furthermore, two proton signals at $\delta 2.79$ (H-1 α) and 2.18 (H-12 α) were also shifted downfield to $\delta 3.04$ and 2.48 (pyridine- d_5) by the pyridine induced deshielding effect [5] indicating that two hydroxyl groups and two protons (OH-9 α and H-1 α , OH-14 α and H-12 α) occupied 1,3-*syn*-periplanar posi-

tions. Therefore, the configurations at C-9 and C-14 have been assigned as *S*, respectively. From the above results, the structure of calvasterol B (**2**) was established as 9 α ,14 α -dihydroxyergosta-4,7,22-trien-3,6-dione.

A large number of steroids, which have the ergostane skeleton, have been isolated from fungi. However, calvastrols A (**1**) and B (**2**) are the first examples of a naturally occurring hydroxylated ergosta-4,7,22-trien-3,6-dione type of steroid.

EXPERIMENTAL

General. Mps: uncorr. IR spectra were recorded in KBr discs. EI-MS were taken at 70 eV. ^1H NMR (600 MHz) and ^{13}C NMR (150 MHz) were recorded in CDCl_3 with TMS as int. standard. Low pressure LC (LP-LC) was performed on a Nihon Seimitsu NP-FX-20 in a glass column (300 \times 10 mm) packed with silica gel CQ-3 (30–50 μ ; Wako).

Isolation of metabolites 1 and 2. The fruiting body of *Calvatia cyathiformis* (4 kg) was extracted with CH_2Cl_2 , and the organic layer was dried (Na_2SO_4) and evapd in

Table 1. ^{13}C NMR data of calvasterols A (1) and B (2), and related compound 5 (in CDCl_3)

C	1	2	5
1	34.5	27.7	35.5
2	34.3	34.3	34.4
3	199.7	200.1	200.1
4	127.0	125.5	124.4
5	156.3	155.4	168.4
6	188.7	188.1	187.7
7	121.8	129.1	126.4
8	156.0	163.4	158.7
9	138.5	74.4	47.3
10	38.8	44.1	39.1
11	132.9	27.6	21.9
12	37.4	27.7	38.6
13	46.3	46.4	44.8
14	84.7	87.0	56.3 ^a
15	31.2	31.9	22.6
16	27.2	26.3	27.8
17	50.4	50.2	56.5 ^a
18	16.2	16.4	12.9
19	29.5	22.9	19.6
20	40.1	40.0	40.4
21	20.9	21.3	21.2
22	135.4	135.4	135.3
23	133.3	133.4	133.2
24	42.9	43.0	43.0
25	33.2	33.2	33.2
26	20.0	20.0	20.0
27	19.7	19.7	19.7
28	17.6	17.7	17.6

^aThe assignments may be reversed.

Table 2. Three-bond correlations (HMBC experiments) of the steroid ring of calvasterols A (1) and B (2)

H	C	
	1	2
1	2*, 3, 5, 9, 10*, 19	2*, 3, 9, 10*, 19
2	1*, 3*, 4, 10	1*, 3*, 10
4	2, 5*, 6, 10	2, 5*, 6, 10
7	5, 9, 14	5, 9, 14
11	8, 10, 12*, 13	8, 12*
12	9, 11*, 13*, 14, 18	9, 11*, 13*, 14, 18
15	13, 14*, 16*, 17	14*
16	14, 15*, 20	14, 17*, 20
17	18, 20*	15, 16*, 18, 21
18	112, 13*, 14, 17	12, 13*, 14, 17
19	1, 5, 9, 10*	1, 5, 9, 10*

*Two-bond correlation.

vacuo. The residue (13.5 g) was chromatographed on silica gel with C_6H_6 – Me_2CO (10:1) followed by LP-LC using *n*-hexane–EtOAc (5:1) to give a calvasterol B (9 α ,14 α -dihydroxyergosta-4,7,22-triene-3,6-dione) (2) (15 mg), and then *n*-hexane–EtOAc (4:1) to give calva-

sterol A (14 α -hydroxyergosta-4,7,9,22-tetraene-3,6-dione) (1) (10 mg).

Calvasterol A (1). Pale yellow needles (*n*-hexane– CHCl_3 , 1:1); mp 190–192° [α]_D²⁰ + 93° (CHCl_3 ; *c* 0.39). IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3450 (OH), 1660 (CO). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 250 (4.35), 340 (4.06). EI-MS (probe) 70 eV, *m/z* (rel. int.): 422.2815 [M]⁺ ($\text{C}_{28}\text{H}_{38}\text{O}_3$ requires 424.2820, 2), 404 [$\text{M} - \text{H}_2\text{O}$]⁺ (60), 280 (55), 265 (38). ^1H NMR (600 MHz, CDCl_3 , TMS as int. std.): δ 0.76 (3H, s, H-18), 0.84 (3H, *d*, *J* = 6.9 Hz, H-26), 0.85 (3H, *d*, *J* = 6.9 Hz, H-27), 0.93 (3H, *d*, *J* = 6.6 Hz, H-28), 1.04 (3H, *d*, *J* = 5.9 Hz, H-21), 1.44 (1H, *m*, H-16b), 1.46 (3H, s, H-19), 1.50 (1H, *m*, H-25), 1.77 (1H, *m*, H-15b), 1.89 (1H, *m*, H-24), 2.00 (2H, *m*, H-15a, H-16a), 2.09 (1H, *m*, H-20), 2.11 (1H, *m*, H-17), 2.28 (1H, *ddd*, *J* = 5.5, 13.6, 14.3 Hz, H-1a), 2.38 (1H, *dd*, *J* = 6.6, 18.3 Hz, H-12b), 2.42 (1H, *ddd*, *J* = 4.5, 5.1, 13.6 Hz, H-1b), 2.59 (1H, *ddd*, *J* = 4.5, 5.5, 19.4 Hz, H-2b), 2.65 (1H, *ddd*, *J* = 5.1, 14.3, 19.4 Hz, H-2a), 2.72 (1H, *dd*, *J* = 1.8, 18.3, H-12a), 5.22 (1H, *dd*, *J* = 7.7, 15.4 Hz), 5.30 (1H, *dd*, *J* = 7.7, 15.4 Hz, H-23), 6.17 (1H, s, H-7), 6.22 (1H, *dd*, *J* = 1.8, 6.6 Hz, H-11), 6.45 (1H, s, H-4) ^{13}C NMR: Table 1. CD (MeOH; *c* 7.1×10^{-5}): $\Delta\epsilon_{275} - 3.46$, $\Delta\epsilon_{234} + 1.57$.

Calvasterol B (2). Pale yellow needles; mp 173–175°; [α]_D²⁰ – 103° (CHCl_3 ; *c* 0.27). IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3450 (OH), 1660 (CO). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 266 (4.09). EI-MS (probe) 70 eV, *m/z* (rel. int.): 440.2932 [M]⁺ ($\text{C}_{28}\text{H}_{40}\text{O}_4$ requires 440.2926, 4), 422 [$\text{M} - \text{H}_2\text{O}$]⁺ (38), 297 (26), 229 (87). ^1H NMR (600 MHz, CDCl_3 , TMS): δ 0.77 (3H, s, H-18), 0.84 (3H, *d*, *J* = 6.9 Hz, H-26), 0.85 (3H, *d*, *J* = 6.9 Hz, H-27), 0.93 (3H, *d*, *J* = 6.9 Hz, H-28), 1.05 (3H, *d*, *J* = 6.6 Hz, H-21), 1.37 (3H, s, H-19), 1.46 (1H, *m*, H-16 β), 1.49 (1H, *m*, H-25), 1.65 (1H, *m*, H-15 β), 1.75 (1H, *br d*, *J* = 13.6 Hz, H-12 β), 1.82 (1H, *br d*, *J* = 13.9 Hz, H-1 β), 1.83 (1H, *m*, H-11 β), 1.88 (1H, *m*, H-24), 1.94 (1H, *m*, H-16 α), 1.97 (1H, *m*, H-15 α), 2.01 (1H, *m*, H-17), 2.09 (1H, *m*, H-11 α), 2.11 (1H, *m*, H-20), 2.18 (1H, *ddd*, *J* = 4.4, 13.2, 13.6 Hz, H-12 α), 2.46 (1H, *br s*, OH-9 α or 14 α), 2.50 (1H, *ddd*, *J* = 5.5, 13.9, 16.8 Hz, H-2 β), 2.59 (1H, *ddd*, *J* = 4.0, 5.1, 16.8 Hz, H-2 α), 2.79 (1H, *ddd*, *J* = 5.1, 13.9, 13.9 Hz, H-1 α), 4.22 (1H, *br s*, OH-9 α or 14 α), 5.20 (1H, *dd*, *J* = 8.4, 15.0 Hz, H-22), 5.29 (1H, *dd*, *J* = 7.7, 15.0 Hz, H-23), 6.19 (1H, s, H-7), 6.60 (1H, s, H-4). ^{13}C NMR: Table 1. CD (MeOH; *c* 6.8×10^{-5}): $\Delta\epsilon_{290} - 2.94$, $\Delta\epsilon_{248} + 4.30$.

Hydroxylation of ergosta-4,7,22-trien-3,6-dione (5). A 1 M K_2CO_3 soln (0.5 ml) was added to a stirred soln of ergosta-4,7,22-trien-3,6-dione (5) (37 mg) in Me_2CO (10 ml) and refluxed for 20 min. The reaction mixture was poured into ice- H_2O and extracted with CHCl_3 . The evapd residue was purified by LP-LC using *n*-hexane–EtOAc (5:1) to obtain 9 α ,14 α -dihydroxyergosta-4,7,22-trien-3,6-dione (2) (2 mg), [α]_D²⁰ – 87° (CHCl_3 ; *c* 0.06). This compound was identical with calvasterol B (2) on the basis of a comparison of the ^1H NMR and IR spectra and the optical rotation.

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REFERENCES

1. Kawahara, N., Sekita, S. and Satake, M. (1994) *Phytochemistry* (in press).
2. Chiang Su New Medical College (ed.), *Dictionary of Chinese Crude Drugs* (1977) p. 283. Shanghai Scientific Technologic Publisher, Shanghai.
3. Adler, J. O., Young, M. and Nes, W. R. (1977) *Lipids* **12**, 364.
4. Malorini, A., Minale, L. and Riccio, R. (1978) *Nouv. J. Chim.* **2**, 351.
5. Demarco, P. V., Farkas, E., Doddrell, E., Mylari, B. L. and Mulheirn, L. J. (1968) *J. Am. Chem. Soc.* **90**, 5480.