



4-QUINOLINONE ALKALOIDS FROM *DICTYOLOMA PERUVIANA*

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Abstract—The stem-bark of *Dictyoloma peruviana* yielded two new piperidino [1,2-a] 4-quinolinones, dictyolomide A and dictyolomide B. Their structures were established by NMR spectroscopy.

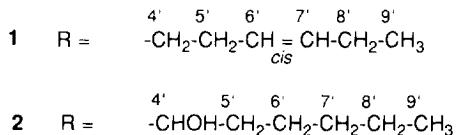
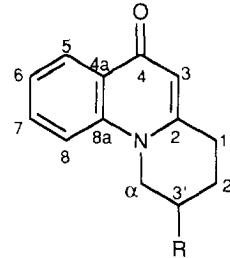
INTRODUCTION

Only two species belong to the genus *Dictyoloma*: *D. incanescens* (syn. *D. vandellianum*) and *D. peruviana* and they are widespread in Bolivia. *Dictyoloma incanescens* is known to contain indole alkaloids [1, 2], 2-quinolinone alkaloids [1], prenylated chromones [3] and limonoids [1]. *Dictyoloma peruviana* is a small tree used in folk medicine for the treatment of leishmaniasis; to the best of our knowledge, it has not been the subject of phytochemical investigations. As part of a cooperative programme on the search for new antileishmaniasis agents from natural sources, bioassay-guided separations were conducted which led to the isolation and structural elucidation of two 4-quinolinone alkaloids from the stem bark of this species.

RESULTS AND DISCUSSION

Dictyoloma peruviana was collected in the Carrasco province of Bolivia. *In vitro* antileishmanial activity tests were performed on various strains of promastigote forms of *Leishmania* species. Crude extracts and pure compounds, aseptically dissolved in liquid medium and DMSO (final concentration of DMSO less than 0.1%), were assayed in triplicate as previously described [4]. Biological activity was concentrated in the ethyl acetate fraction, which contained two compounds (named dictyolomides A and B), for which structures **1** and **2** are proposed. Antileishmanial activity could be attributed to these compounds which induced complete lysis of parasites at 100 µg ml⁻¹ (Table 1).

The two compounds belonged to the same class of molecules and mass spectrometry allowed determination of *M_r* of 281 (C₁₉H₂₃NO) for **1** and 299 (C₁₉H₂₅NO₂) for



2. The 4-quinolone skeleton [5] was recognized on the basis of UV spectra and ¹H NMR signals at 6.30 ppm (H-3) and 8.47 ppm (H-5 deshielded by a *peri*-carbonyl group). The chemical shifts of C-4 carbonyls at 176.8 ppm in **1** and 176.7 ppm in **2** are characteristic of a vinyllogous amide.

On the basis of the hypothesis of **1** being a 4-quinolone, its elemental composition indicates the presence of two supplementary unsaturations. Their location and the proposed structure **1** is based on the assignment of ¹H and ¹³C NMR spectra with the help of 2-D correlation experiments. These unsaturations are located in the C₁₀H₁₉ fragment anchored at positions 1 and 2 of the quinolone. The ¹H NMR spectrum of **1** displayed five signals, corresponding to the protons of rings A and B of the quinolone. HMQC and HMBC experiments allowed identification of the carbon signals for these rings. Proton signals not included in the quinolone system were observed at δ 5.45 and 5.35 (ethylenic protons attached to

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Table 1. *In vitro* activity of *D. peruviana* extracts and of alkaloids **1** and **2** on promastigote forms of *Leishmania* species

Extract	Strain*	Concentration ($\mu\text{g ml}^{-1}$)†			
		100	50	25	10
Petrol	<i>L. a.</i>	+	0	0	—
	<i>L. b.</i>	++	++	0	—
Ethyl acetate	<i>L. a.</i>	+++	++	0	—
	<i>L. b.</i>	+++	+	0	—
Alcoholic	<i>L. a.</i>	++	+	0	—
	<i>L. b.</i>	+	+	0	—
Alkaloid mixture	<i>L. a.</i>	+++	+++	++	—
	<i>L. b.</i>	++	++	0	—
Alkaloid 1	<i>L. a.</i>	+++	+++	++	—
	<i>L. b.</i>	++	+	0	—
Alkaloid 2	<i>L. a.</i>	+++	+	0	—
	<i>L. b.</i>	++	+	+	—
Pentamidine	<i>L. a.</i>	—	—	—	+++
	<i>L. b.</i>	—	—	—	+++

* *L. a.* = *Leishmania amazonensis*; *L. b.* = *Leishmania brasiliensis*.

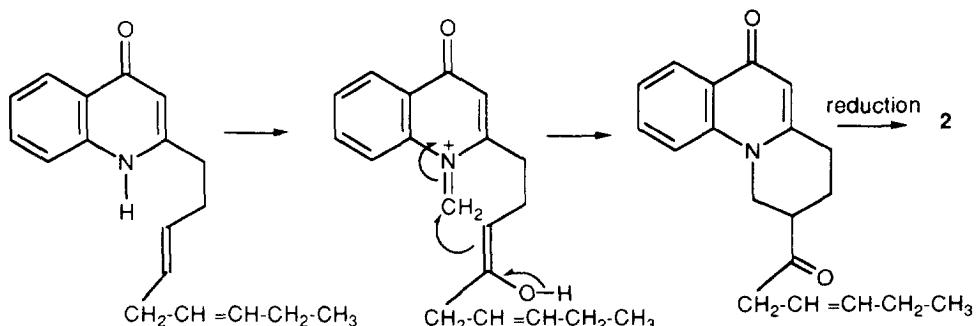
† 0: Promastigotes identical to control; +: 75% promastigotes, with a few degenerative forms; ++: 50% promastigotes, with a few degenerative forms; +++: no promastigotes, total lysis of parasites.

Table 2. ^{13}C NMR data and HMBC correlations of alkaloids **1** and **2**

C	HMBC correlations			
	1	2	1	2
2	152.1	152.7	H-3, H- α , H-1'	H-3, H- α , H-1', H-2'
3	109.7	109.0	H-1'	H-1'
4	176.7	176.8	H-5	H-5
4a	126.5	126.3	H-3, H-6,	H-3, H-6,
5	126.8	126.3	H-7, H-8	H-7, H-8
6	124.0	123.8	H-7, H-8	H-8
7	132.0	131.9	H-5, H-6, H-8	H-5
8	114.8	115.2	H-6	H-6
8a	141.3	141.3	H-5, H-7, H α	H-5, H-7, H α
α	51.2	47.2	H-4'	H-4'
1'	30.0	29.9	H-3	H-3, H-2'
2'	25.0	22.2	H- α , H-1', H-4'	H- α , H-1'
3'	33.8	40.0	H- α , H-1', H-5'	H- α , H-1', H-2'
4'	34.0	73.4	H-6', H-7'	H- α
5'	24.1	34.7	H-4', H-6', H-7'	
6'	127.8	25.2	H-4', H-5', H-8'	H-4'
7'	133.0	31.8	H-5', H-8', H-9'	H-8', H-9'
8'	20.6	22.6	H-6', H-7', H-9'	H-7', H-9'
9'	14.5	14.0	H-8'	H-8'

carbons at δ 127.8 and 133.0), at δ 4.30 (*dd*, J = 12.5 and 5 Hz) and 3.59 (*dd*, J = 12.5 and 10.5 Hz) as the AB part of an ABX system, between δ 3.0 and 1.45 (11H, multiplets) and at δ 1.00 (*t*, J = 7 Hz) for one terminal methyl. The ^{13}C NMR spectrum showed 19 carbons, 11 down-field of chloroform, with the nine sp^2 carbons of the 4-quinolinone part and the two supplementary ethylenic

methines at δ 127.8 and 133.0 and, at higher field, one methyl, one methine sp^3 and six methylenes (Table 2). The ABX system was assigned to a $\text{N}-\text{CH}_2-\text{CH}-$ moiety with attachment of the nitrogen based on the chemical shift of the methylene (δ 51.2). Two adjacent methylenes were detected in the COSY experiment at δ 3.0 ppm (*m*, 2H), 2.05 (*m*, 1H) and 1.45 (*m*, 1H). Coupling of the latter



Scheme 1. Proposed rate for biogenesis of alkaloids 1 and 2.

methylene with the aliphatic CH could not be determined because of overlap but the other methylene was coupled at long-range with H-3 of the quinolone. This CH₂-CH₂ fragment is probably attached to C-2 of the quinolone. The HMBC experiment allowed linkage of these fragments through observation of long-range couplings between the N-CH₂ and quaternary carbon atoms of the quinolone (C-8a at δ 141.3 and C-2 at δ 152.1) and between H-1' of the CH₂-CH₂ fragment and C-2. The number of unsaturations and the presence of one double bond imply one ring and one chain for the non-quinolone fragment; the chain must therefore be attached to the aliphatic methine. The composition of the chain was determined as C₆H₁₁ from the base peak in the mass spectrum (*m/z* 198); the count of methylenes (three) indicates that the chain is linear and contains a disubstituted double bond. Identification of the chain requires the presence of a supplementary ring as a piperidine substituted at C-3' (alkaloid numbering) by the alkenyl chain. Final location of the unsaturation in the chain is based on observation of a long-range correlation between the protons of the methyl (triplet at δ 1.0, *J* = 7 Hz and, therefore, CH₃-CH₂-) and an ethylenic carbon at δ 133.0. Compound 1, dictyolomide A, is therefore a piperidino [1,2-a] 4-quinolinone substituted by a 3-hexenyl chain at C-3'. The *cis*-configuration of the double bond is deduced from the chemical shift of the terminal methylene (δ 20.6) shielded as in the *cis*-hex-3-ene at δ 20.7.

The second compound (2) showed a [M]⁺ at *m/z* 299, which corresponds to a molecular formula of C₁₉H₂₅NO₂. This suggested that 2 is a hydroxyl derivative of 1. The ¹H and ¹³C NMR spectra of 2 displayed many similarities with those of 1 (Table 2). The major differences were the absence of the ethylenic system in the linear chain and the presence of a hydroxymethine group at δ _H 3.60 and δ _C 73.4 in 2. This proton shows coupling with H-3' of the piperido ring in the COSY spectrum and a cross-peak with C- α in the HMBC experiment; the reverse correlation between one of the H- α protons (δ 3.96) and this hydroxymethine carbon is also observed. Thus, the chain is hydroxylated at position 4' and the appendage fragment corresponds to a hexan-1-ol in alkaloid 2.

The isolation in two other species of Rutaceae of 4-quinolinone alkaloids with an alkyl chain at C-2, 2-(3'-6'-nonadienyl)-4-quinolone [6] and 1-methyl-2-nonyl-4-quinolone [7], allows us to propose a biogenetic scheme for compounds 1 and 2 involving a Mannich reaction for the formation of the third ring (Scheme 1). This reaction would leave a carbonyl function at position 4', which is further functionalized in compound 2.

EXPERIMENTAL

UV spectra measured in MeOH. ¹H and ¹³C NMR were recorded at 300 and 75 MHz respectively, in CDCl₃. 2-D experiments were performed using standard Bruker microprograms. Hardware modifications of the spectrometer allowed acquisition of C-H correlations in the reverse mode (HMQC and HMBC (d_4 = 70 msec)).

Plant material. *Dictyoloma peruviana* Planchon was collected at 'Valle del Sacta', Cochabamba area, Carrasco province (Bolivia), in November 1991. A voucher specimen is deposited at the Herbarium of San Andrés University in La Paz.

Extraction and purification of alkaloids. Ground stem bark (1097 g) was wetted with 50% NH₄OH and extracted (*×* 7) with CHCl₃. The extracts were concd (200 ml) under red. pres. then extracted with 5% HCl. The aq. phase was made alkaline with 50% NH₄OH and extracted with CHCl₃. The CHCl₃ layer was washed with H₂O, dried (Na₂SO₄) and then evapd *in vacuo* to give 1.38 g crude alkaloid mixt. (1.25 g kg⁻¹). The CHCl₃ residue was chromatographed on a column of silica gel (48 g) eluting with increasing percentages of CHCl₃-MeOH: 49:1, 97.3, 19:1 and 9:1. From the CHCl₃-MeOH (97:3) eluate, alkaloid 1 was obtained in frs 75-86; the CHCl₃-MeOH (19:1) eluate, fr. 135-150, yielded alkaloid 2.

Dictyolomide A (1). $[\alpha]_D$ + 21° (*c* 0.6, CHCl₃). EIMS *m/z* (rel. int.): 281 ([M]⁺, 63), 266 (8), 252 (10), 226 (27), 212 (33), 198 (100), 144 (70). ¹H NMR: δ 1.00 (*t*, *J* = 7 Hz, 3H, H-9'), 1.45 (*m*, H-2'), 1.57 (*m*, 2H, H-4'), 2.05, (*m*, 4H, H-2' + H-3' + Hs-8'), 2.20 (*br*, *q*, *J* = 7 Hz, 2H, H-5'), 3.00 (*m*, 2H, H-1'), 3.59 (*dd*, *J* = 12.5 and 10.5 Hz, H- α), 4.30 (*dd*, *J* = 12.5 and 5 Hz, H- α), 5.35 (*m*, H-6'), 5.45 (*m*, H-7'), 6.30 (*br* *s*, H-3), 7.42 (*t*, *J* = 8.5 Hz, H-6), 7.59 (*d*,

$J = 8.5$ Hz, H-8), 7.68 (*td*, $J = 8.5$ and 1.5 Hz, H-7), 8.47 (*dd*, $J = 8.5$ and 1.5 Hz, H-5). ^{13}C NMR: see Table 2.

Dictyolomide B (**2**). $[\alpha]_D + 32^\circ$ (*c* 0.9, CHCl_3). EIMS m/z (rel. int.): 299 ($[\text{M}]^+$, 1), 279 (15), 222 (5), 167 (40). ^1H NMR: δ 0.90 (*t*, $J = 7$ Hz, 3H, H-9'), 1.30 (*m*, 2H, H-8'), 1.45–1.75 (*m*, 7H, H-2 + Hs-5' + Hs-6' + Hs-7'), 1.96 (*dd*, $J = 12.5$ and 4 Hz, H-2'), 2.08 (*m*, $W_{1/2} \approx 35$ Hz, H-3'), 2.92 (*m*, 2H, H-1'), 3.60 (*m*, $W_{1/2} \approx 15$ Hz, H-4'), 3.96 (*dd*, $J = 12.5$ and 10.5 Hz, H- α), 4.35 (*dd*, $J = 12.5$ and 5 Hz, H- α), 6.20 (*br s*, H-3), 7.36 (*t*, $J = 7.5$ Hz, H-6), 7.60 (*m*, H-7 + H-8), 8.40 (*d*, $J = 7.5$ Hz, H-5). ^{13}C NMR: see Table 2.

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