



DRIMANES FROM THE EPICUTICULAR WAX OF THE FERN *NEPHROLEPIS BISERRATA*

KARSTEN SIEMS, FRANKA WEIGT and ECKHARD WOLLENWEBER*

AnalytiCon Gesellschaft für Chemische Analytik und Consulting, Gustav-Meyer-Allee 25, D-13355 Berlin, Germany; *Institut für Botanik der Technischen Hochschule, Schnittspahnstrasse 3, D-64287 Darmstadt, Germany

(Received in revised form 29 August 1995)

Key Word Index—*Nephrolepis biserrata*; Davalliaceae; Pteridophyta; leaf wax; drimane sesquiterpenes; acetylated hemiacetals.

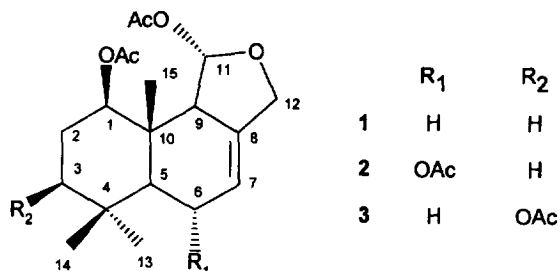
Abstract—From the thin epicuticular wax layer of the tropical fern *Nephrolepis biserrata*, three new drimane-type sesquiterpenes (1 β ,11 α -diacetoxy-11,12-epoxydrim-7-ene, 1 β ,6 α ,11 α -triacetoxy-11,12-epoxydrim-7-ene, 1 β ,3 β ,11 α -triacetoxy-11,12-epoxydrim-7-ene) were isolated. All these new compounds contain a hemiacetal esterified with acetic acid.

INTRODUCTION

Diterpenes and triterpenes have been found before as constituents of the epicuticular material accumulated on fronds of ferns belonging to various taxa [1-3]. They form either farinose exudates, often together with flavonoids, e.g. in *Cheilanthes* and *Notholaena* species, or thin wax layers that cause the glaucous appearance of fern fronds, e.g. in *Lophosora quadripinnata* and *Polypodium aureum*. In the tropical fern *Nephrolepis biserrata* (Sw.) Schott, it is the rachis of its large fronds that bears a thin wax layer.

RESULTS AND DISCUSSION

The epicuticular wax recovered from the leaf rachis of *N. biserrata* contains three drimane-type sesquiterpenes (1-3) with an unusual acetylated hemiacetal. The pres-



ence in all three of two acetates followed from typical singlets at about δ 2.0 in the ^1H NMR spectra together with singlets of about 170 ppm and quartets (about 21 ppm) in the ^{13}C NMR spectra. Beside these six ester carbons, the ^{13}C NMR spectrum of 3 showed 3q, 3t (one oxygen-substituted at 69.2 ppm), 6d (three oxygen-substituted below 70 ppm, and an olefinic at 116.2 ppm) and

3s (one olefinic). From these data, the presence of two carbocyclic rings could be deduced. Bicyclic sesquiterpene skeletons with two singlets in the ^{13}C NMR upfield 60 ppm are quite rare; one possibility is the drimane-skeleton. Spin decoupling established the sequence C-1 to C-3 and C-5 to C-11. The olefinic proton H-7 coupled allylically to H-9, H-12 and H-12₂. The stereochemistry at C-6 could be deduced from the NOE of H-6 to two methyl groups (H-14 and H-15). The coupling pattern (dd , $J=11$ Hz and $J=4$ Hz) of the signal at δ 4.67 allowed two positions for the third acetate: C-1 or C-3. NOE experiments (5% NOE from H-1 to H-9) and the coupling pattern of H-1 indicate that C-1 bears an equatorial acetate. The vicinal coupling constant of about 3 Hz (H-9 to H-11) is not sufficient for determination of the stereochemistry at C-11, because similar coupling constants were reported for acetates of drimeninol and isodrimeninol [4, 5]. The stereochemistry at C-11 was established by a strong NOE-effect, H-11 to the methyl group H-15 (12%). The ^1H NMR-spectrum of 1 is similar to the ^1H NMR-spectrum of 2, but obviously the acetate at C-6 is missing. There is one acetate at C-1 (5% NOE from axial H-1 to H-9) and another on the acetylated hemiacetal at C-11. 2 has acetates at C-1 and C-11 and an additional equatorial acetate at C-3. The coupling pattern and chemical shifts of H-1 and H-3 are very similar, but by NOE-experiments (effect from H-9 to H-1 and H-5, respectively, and from H-3 to H-5) they could be assigned unambiguously. All the ^{13}C NMR signals were assigned by HMQC. On EIMS none of the compounds 1-3 gave a molecular ion corresponding to its molecular formula deduced from the ^{13}C NMR spectra, but in each case a fragment corresponding to the loss of acetic acid present. The absolute stereochemistry of the new drimanes was not determined.

Table 1. ^1H NMR data of compounds 1–3 (CDCl_3 , 400 MHz, int. standard solvent peak = 7.26 ppm)

H	1	2	3
1 _{ax}	4.59 <i>dd</i> (12; 4)	4.67 <i>dd</i> (11; 4)	4.73 <i>dd</i> (12; 4)
2 _{eq}	1.72 <i>dddd</i> (13; 4; 4; 4)	1.77 <i>dddd</i> (13; 4; 4; 4)	1.97 <i>ddd</i> (12; 4; 4)
2 _{ax}	1.61 <i>dddd</i> (13; 13; 12; 4)	1.65 <i>m</i>	1.72 <i>ddd</i> (12; 12; 12; 12)
3 _{eq}	1.48 <i>ddd</i> (13; 4; 4)	1.43 <i>*</i>	—
3 _{ax}	1.41 <i>*</i>	1.48 <i>*</i>	4.68 <i>dd</i> (12; 4)
5	1.39 <i>dd</i> (12; 5)	1.67 <i>d</i> (10)	1.39 <i>dd</i> (11; 6)
6 ₁	2.03 <i>m</i>	5.55 <i>br d</i> (10)	2.12 <i>br d</i> (18)
6 ₂	2.19 <i>br d</i> (18)	—	2.24 <i>br d</i> (18)
7	5.59 <i>br s</i>	5.65 <i>br s</i>	5.60 <i>br s</i>
9	2.58 <i>br s</i>	2.72 <i>br s</i>	2.55 <i>br s</i>
11	6.30 <i>d</i> (3)	6.37 <i>d</i> (3)	6.28 <i>d</i> (3)
12 ₁	4.20 <i>br d</i> (12)	4.28 <i>br d</i> (12)	4.21 <i>br d</i> (11)
12 ₂	4.32 <i>br d</i> (12)	4.39 <i>dddd</i> (12; 1; 1; 1)	4.32 <i>dddd</i> (11; 2; 1; 1)
13	0.89 <i>s</i>	0.92 <i>s</i>	0.87 <i>s</i>
14	0.94 <i>s</i>	1.03 <i>s</i>	0.93 <i>s</i>
15	0.94 <i>s</i>	1.00 <i>s</i>	0.96 <i>s</i>
Ac-1	1.97 <i>s</i>	1.98 <i>s</i>	1.94 <i>s</i>
Ac-2	2.03 <i>s</i>	2.03 <i>s</i>	2.01 <i>s</i>
Ac-3	—	2.03 <i>s</i>	2.03 <i>s</i>

*Overlapping.

Drimanes are widespread in nature. They are already known from a few families of higher plants (e.g. *Drimys* sp. [6, 7] and *Polygonum* sp. [8]), from liverworts (e.g. *Porella cordeana* [9]) and from fungi (e.g. *Phoma asparagi* [10] and the mushroom *Marasmius oreades* [11]). Acetic esters of a drimane-hemiacetal were isolated from the nudibranch *Dendrodoris grandiflora* [12]. The function of drimanes, especially of polygodial or protected forms of aldehydes from nudibranches [12] as defensive agents and antifeedants is well investigated, as is their synthetic chemistry [13]. To the best of our knowledge, this is the first report of drimane sesquiterpenes from ferns. It is also the first report of any sesquiterpenes forming a wax layer on fern fronds.

EXPERIMENTAL

Nephrolepis biserrata was propagated in a greenhouse of the Botanischer Garten der TH Darmstadt. Pinnae were removed from the fronds and the rachis were rinsed with CHCl_3 to dissolve the waxy material. The fractions obtained by CC of the concentrated crude soln (silica gel, hexane–EtOAc gradient) were examined by ^1H NMR spectroscopy. Sesquiterpene-containing fractions were further sepd by prep TLC (petrol–Et₂O 3:2) to obtain pure compounds that exhibit R_f 0.36 (1), 0.22 (2), and 0.20 (3), respectively.

1 β ,11 α -Diacetoxy-11,12-epoxydrim-7-ene (1). MS m/z (rel. int.): 276 (100) $[\text{M} - \text{HOAc}]^+$, 261 (4) $[276 - \text{CH}_3]^+$, 217 (12) $[276 - \text{OAc}]^+$.

1 β ,6 α ,11 α -Triacetoxy-11,12-epoxydrim-7-ene (2). MS m/z (rel. int.): 334 (40) $[\text{M} - \text{HOAc}]^+$, 275 (5) $[334 - \text{OAc}]^+$.

Table 2. ^{13}C NMR data of compounds 1–3 (CDCl_3 , 100.6 MHz, int. standard CDCl_3 , 77.0 ppm)

C	1	2	3
1	81.4	80.2	77.5 <i>d</i>
2	24.2	24.2	29.3 <i>t</i>
3	39.6	39.8	76.2 <i>t/d</i>
4	32.7	33.7	37.2* <i>s</i>
5	48.9	53.1	46.7 <i>d</i>
6	23.3	70.3	22.5 <i>t/d</i>
7	117.6	116.2	117.0 <i>d</i>
8	135.1	140.6	134.9 <i>s</i>
9	57.9	57.3	57.7 <i>d</i>
10	37.4	42.1	37.4* <i>s</i>
11	98.8	98.6	98.2 <i>d</i>
12	69.6	69.2	69.4 <i>t</i>
13	32.3	33.0	27.1 <i>q</i>
14	21.3	22.0	15.5 <i>q</i>
15	9.1	10.8	9.1 <i>q</i>
Ac	21.3	21.1	20.8 <i>q</i>
Ac	21.3	21.0	20.8 <i>q</i>
Ac	—	21.4	21.0 <i>q</i>
Ac	170.5	170.2	169.7 <i>s</i>
Ac	171.1	170.5	170.0 <i>s</i>
Ac	—	170.6	170.9 <i>s</i>

*Interchangeable.

1 β ,3 β ,11 α -Triacetoxy-11,12-epoxydrim-7-ene (3). MS m/z (rel. int.): 334 (20) $[\text{M} - \text{HOAc}]^+$, 275 (43) $[334 - \text{OAc}]^+$.

For ^1H NMR and ^{13}C NMR data see Tables 1 and 2.

Acknowledgement—E. W. wishes to thank Mr Helmut Groh for the plant material and Mrs Marion Dörr for technical assistance. K. S. wishes to thank Dr Jakupovic, TU Berlin, for discussion of NMR spectra.

REFERENCES

1. Rüedi, P., Wollenweber, E., Marx, D. and Scheele, C. (1989) *Z. Naturforsch.* **44c**, 901.
2. Arriaga-Giner, F. J., Rumero, A. and Wollenweber, E. (1992) *Z. Naturforsch.* **47c**, 508.
3. Tanaka, N., Wada, H., Dietz, V. and Wollenweber, E. (1992) *Phytochemistry* **31**, 347.
4. Fukuyama, Y., Sato, T., Asakawa, Y. and Takemoto, T. (1982) *Phytochemistry* **21**, 2895.
5. Asakawa, Y., Toyota, M. and Takemoto, T. (1978) *Phytochemistry* **17**, 457.
6. Appel, H. H., Brooks, C. J. W. and Overton, K. H. (1959) *J. Org. Chem.* 3332.
7. Vichnewski, W., Kulanthaivel, P. and Herz, W. *Phytochemistry* (1986) **25**, 1476.
8. Jacobsson, U. and Muddathir, K. (1992) *Phytochemistry* **31**, 4207.
9. Harrigan, G. G., Ahmad, A., Baj, N., Glass, T. E., Gunatilaka, A. A. L. and Kingston, D. G. I. (1993) *J. Nat. Prod.* **56**, 921.
10. Ichihara, A., Sawamura, S. and Sakamura, S. (1984) *Tetrahedron Letters* **25**, 3209.
11. Ayer, W. A. and Craw, P. A. (1989) *Can. J. Chem.* **67**, 1371.
12. Cimino, G., Sodano, G. and Spinella, A. (1988) *J. Nat. Prod.* **51**, 1010.
13. Jansen, B. J. M. and de Groot, A. (1991) *Nat. Prod. Rep.* **8**, 319.