



PHYLLOCLADANES (13β-KAURANES) FROM PLECTRANTHUS AMBIGUUS

GUI LIU and PETER RÜEDI*

Organisch-chemisches Institut der Universität Zürich, Winterthurerstrasse 190, CH-8057 Zürich, Switzerland

(Received in revised form 18 September 1995)

Key Word Index—Plectranthus ambiguus; Labiatae; phyllocladane (13 β -kaurane) skeleton; structure elucidation; chemical correlation.

Abstract—Six new diterpenoids of the rare phyllocladane (13β -kaurane) type have been isolated from *Plectranthus ambiguus*. Their structures were determined on the basis of their spectral properties and by correlation with 16R,17-dihydroxyphyllocladane-3-one (calliterpenone).

INTRODUCTION

In continuation of our current research concerning biologically active constituents of African and Asian medicinal plants [1], we investigated the labiate *Plectranthus ambiguus* [2] with respect to antioxidative activity and inhibition of the enzymes of arachidonic acid metabolism. Antioxidant activity guided fractionation of the airdried plant material (see Experimental) and afforded the main antioxidant principle of this species, the quite rare 5,6-dihydroxy-7,4'-dimethoxyflavone (1) (ladanein or 4',7-dimethylscutellarein) [3–7]. The mother liquor of this fraction contained the tetracyclic diterpenoids 2a, 2b, 3a, 3b, 4 and 5a as the main components. The present paper reports on the isolation and structural determination of these six novel phyllocladanes (13 β -kauranes).

RESULTS AND DISCUSSION

Extraction of the aerial parts of *P. ambiguus* followed by partition (hexane-90% methanol) and column chromatography (Sephadex LH-20, silica gel) yielded from the main fraction exhibiting significant antioxidative activity the crystalline flavone 1 (0.026%). Subsequent column chromatography (silica gel) of the mother liquor and preparative HPLC (C-18) afforded the tetracyclic diterpenoids 2a (0.55%), 2b/3b (as a mixture, ca (6:1, 0.08%), 3a (0.29%), 4 (0.10%) and 5a (0.13%).

The molecular formula of compound **2a** was established as $C_{27}H_{42}O_6$ on the basis of its EI-mass spectrum and elemental analysis. Its substituents were assigned as $Me_2C = CHCO_2$ -, AcO- and $-CH_2OH$ based on the MS fragmentation pattern and the following spectral data: UV: 218 (4.15) nm (log ε); IR: 3200–3600 (br), 1746, 1718 and 1650 cm⁻¹; 1H NMR: δ 1.68 and 2.13 (each 3H, each d, $^4J = 1.2$ Hz), δ 5.55 (1H, m, t-like, $W_{1/2}$ ca 3 Hz); δ 2.11 (s, 3H); δ 3.62 and 3.76 (each 1H, ABX,

The mono-O-acetyl derivative **2b** (acetic anhydride-pyridine) still had IR absorption (3590 cm⁻¹) in the hydroxyl region and this was assigned to a tertiary hydroxyl group. The ¹H NMR spectrum showed an additional, primary acetoxy group (δ 2.10, 3H, s; 4.16 and 4.24, each 1H, AB, ²J = 11.3 Hz).

Compound 2a readily gave the acetonide 2c (acetone–CuSO₄ [8]). Its ¹H NMR spectrum showed an AB system for a methylene group at δ 3.90 and 4.06 the ²J = 8 Hz indicating a five-membered ring ketal [9]; hence, 2a had to be a 1,2-dihydroxy compound [8–10].

Compound 2a was easily oxidized (H_5IO_6 -methanol) to the ketone 2d. Its strong IR absorption (1745 cm⁻¹) is typical for five-membered ring ketones. Therefore, 2a is a tetracyclic diterpenoid with a five-membered ring D which is substituted with a primary and a tertiary hydroxyl group. These substituents can only be located at C-17 and C-16, respectively. Based on the low-field chemical shifts of H_2 -17 (AB at δ 3.62 and 3.76) and C-16 (δ 84.2) in 2a, the 16R configuration was tentatively assigned [11, 12].

The positions and the relative stereochemistry of the $Me_2C = CH-CO_2-$ and AcO- substituents were deduced by the ¹H NMR splitting pattern and double resonance experiments: the signals at $\delta 5.01$ (1H, d, ${}^3J = 2.7$ Hz, H_{eq}) and 5.25 (1H, ddd, ${}^3J = 12.4$, 4.5 and 2.7 Hz, H_{ax}) clearly showed a vicinal cis-relationship of the corresponding protons. When the doublet at $\delta 5.01$ was irradiated, only the signals at $\delta 5.25$ and 0.86 (Me-19) and 0.99 (Me-18) showed a NOE. Thus, these two substituents are located at the 2α and 3α position in ring A. Their relative arrangement was established by a heteronuclear ¹H, ¹³C-decoupling experiment: when the proton at $\delta 5.01$ (H-3) was irradiated, the signal of the acetoxy

J=10.8, 5.4 Hz, after adding D₂O: AB, $^2J=10.9$ Hz). The presence of three tertiary Me-signals at $\delta 0.86$, 0.99 and 1.01 (each 3H, each s) and the 13 C NMR data (Table 1) suggested that **2a** was a tetracyclic diterpenoid.

^{*}Author to whom correspondence should be addressed.

Table 1. ¹³C NMR spectral data (50 MHz, CDCl₃)

С	2a	2b/3b	3a	4	5a
1	37.7	38.0	37.9	32.7	44.3
2	66.8	66.9	67.8	22.4	70.2
3	77.0	77.2	77.1	78.2	209.2
4	37.8*	38.1*	38.1*	36.4*	48.3
5	49.5	49.7	49.8	50.3	55.7
6	19.1	19.2†	19.4	19.1 ^b	19.3†
7	40.6	40.7	40.9	41.1	40.4
8	43.1	43.6	43.5	43.6	43.0
9	56.0	56.2	56.2	56.3	56.9
10	38.7*	38.9*	39.0*	37.4*	38.1
11	19.1	19.3†	19.4	19.7†	20.6†
12	26.2	26.5	26.5	26.2	26.2
13	43.3	44.3	43.8	43.8	43.4
14	48.1	48.1	48.3	48.4	48.0
15	44.4	44.8	44.9	44.8	44.4*
16	84.2	82.5	84.5	84.6	84.2
17	65.0	67.6	65.5	65.5	65.0
18	27.6†	27.9‡	27.9	28.0	24.8
19	21.3‡	21.6§	21.7†	21.3‡	21.2‡
20	15.3	15.5	15.6	14.5	15.1
21	165.6	165.8/172.3	172.4		165.3
22	115.6	115.9/43.3	43.3		115.1
23	156.5	156.8/25.3	25.4		157.8
24	19.9‡	20.1§/22.3	22.4		20.1‡
25	27.1†	27.3‡/22.3	22.4		27.2
AcO	20.6‡	20.8§	21.0†	22.0‡	
	170.2	170.4	170.5	170.8	
		20.9§			
		171.1			

^{*†‡§}Assignments are interchangable.

carbonyl group (δ 170.2, dq, $^2J_{\rm CH}=7$ Hz and $^3J_{\rm CH}=4$ Hz) was simplified to a q ($^2J_{\rm CH}=7$ Hz), whereas after irradiation of the proton at δ 5.25 (H-2) the carbonyl signal of the senecioyl moiety (δ 165.6, m, $W_{1/2}$ ca 7 Hz) was significantly narrowed ($W_{1/2}$ ca 4 Hz), i.e. Me₂ C = CH-CO₂- is located at C-2 and AcO- at C-3.

The signal at δ 15.6 in the ¹³C NMR spectrum of **2a** corresponded to a methyl group and was assigned to Me-20. Due to this pronounced high-field chemical shift, the compound is considered to be a phyllocladane-type diterpenoid rather than a kaurane-type [13]. Hence, the structure of **2a** is formulated as $(16R)-2\alpha-(3-\text{methyl-2-butenoyloxy})-3\alpha-\text{acetoxyphyllocladane-16,17-diol}.$

The CI-mass spectrum of compound 3a indicated the molecular formula $C_{27}H_{44}O_6$, which suggested it to be a dihydro derivative of 2a. The ¹H and ¹³C NMR spectra are very similar to those of 2a (see Experimental and Table 1); they mainly lack signals corresponding to the senecioyloxy substituent. Extra signals appear at $\delta 0.92$ and 0.93 (each 3H, each d, $^3J = 6.4$ Hz) for two diastereotopic Me groups of a 2-propyl group [14], and the olefinic carbons are substituted by CH and CH₂ signals.

Acetylation (acetic anhydride-pyridine) of 3a yielded a mono-O-acetyl derivative (3b) and catalytic hydrogenation (Pd/C-methanol) of 2a afforded 3a, which proved to be identical in every respect with the natural product

 $(R_f, \text{ mp, } [\alpha]_D, \text{ mass, IR, }^1\text{H and }^{13}\text{C NMR spectra}).$ Therefore, the structure of **3a** is $(16R)-2\alpha-(3-\text{methyl-butanoyloxy})-3\alpha-\text{acetoxyphyllocladane-}16,17-diol.$

The diacetates **2b** and **3b** were also isolated as natural products (**2b**: **3b** ca 6:1, according to 1 H NMR). However, all attempts to separate the mixture were unsuccessful. Comparing the physical data with those of the pure mono-O-acetyl derivatives **2b** and **3b** prepared from either **2a** or **3a** fully confirmed the structures. Thus, the natural compounds are established as (16R)- 2α -(3-methyl-2-butenoyloxy)- 3α , 17-diacetoxy-16-hydroxyphyllocladane (**2b**) and (16R)- 2α -(3-methylbutanoyloxy)- 3α , 17-diacetoxy-16-hydroxyphyllocladane (**3b**), respectively.

Based on its CI-mass spectrum and ¹³CNMR data, the molecular formula C₂₂H₃₆O₄ can be assigned to compound 4. Its IR spectrum showed absorption bands for hydroxyl groups (3440 and 3325 cm⁻¹) and an ester (1740 cm⁻¹). Its ¹H NMR spectrum revealed three tertiary Me-groups (δ 0.85, 0.89 and 0.91, each 3H, each s), an ABX-system at $\delta 3.64$ and 3.78 (each 1H, J = 10.9 and 5.4 Hz, after adding D_2O : AB, $^2J = 10.9$ Hz, $-CH_2OH$), and an axially oriented acetoxy group (δ 2.08, 3H, s; 4.64, 1H, t, ${}^{3}J = 2.7$ Hz). These facts, together with the ¹³C NMR data (Table 1), indicate that 4 is an acetoxy substituted phyllocladane-16,17-diol. Its position at C-3 was determined by a NOE experiment: when the signal at δ 4.64 was irradiated, the signals of Me-18 (δ 0.89) and Me-19 (δ 0.85) showed a NOE, which establishes the structure of 4 as (16R)-3α-acetoxyphyllocladane-16,17diol.

Compound 5a has the molecular formula C₂₅H₃₈O₅ (CI-mass spectrum and elemental analysis). It exhibited the typical spectral features of a senecioyloxy derivative such as **2a** (δ 1.92, 2.18, each 3H, each d, ${}^4J = 1.2$ Hz), 5.79 (1H, m, t-like, $W_{1/2}$ ca 3 Hz) and an additional carbonyl group (IR 1705 cm⁻¹, 13 C NMR δ 209.2). In the 1 H NMR spectrum there are singlets for three tertiary Me-groups (δ 1.12, 1.16 and 1.27, each 3H, each s) and the ABXsystem for a CH₂OH substituent (δ3.65 and 3.80, each 1H, J = 10.8 and 4.7 Hz, after adding D_2O : AB, $^{2}J = 10.9$ Hz). The compound still has a tertiary hydroxyl group because acetylation (acetic anhydride-pyridine) afforded a monoacetate (5b), whose IR spectrum showed an OH-absorption band at 3580 cm⁻¹. Therefore, it is a senecioyloxy substituted 16,17-dihydroxyphyllocladanone derivative. The ¹³C NMR data supported this assumption (Table 1).

The chemical shift and the multiplicity of the oxymethine proton (δ 5.65, 1H, dd, $^3J = 13.4$ and 6.1 Hz, H_{ax}) clearly showed the ester substituent to be located in an equatorial position adjacent to a carbonyl group [8]. Because the signals of Me-18 and Me-19 are shifted paramagnetically (δ 1.16 and 1.27) compared with those of **2a** (δ 0.86, 0.99), this carbonyl group must be located at C-3 and the senecioyloxy substituent at C-2. Thus, the structure of **5a** is $(16R)-2\alpha-(3-\text{methyl-2-butenoyloxy})-16,17-dihydroxyphyllocladane-3-one.$

Phyllocladane-type (or 13β -kaurane) diterpenoids are rather rare in nature; most of the tetracyclic diterpenoids

$$R^1$$
 R^1
 R^2

2c OCOCH=CMe₂ OAc

$$R^1$$
 R^2

5a OCOCH=CMe₂ H5b OCOCH=CMe₂ Ac

belong to the ent-kaurane series. This fact prompted us to confirm the structures of the new natural products, which were based only on spectroscopic data, and, especially, to determine the absolute configuration unambiguously. Therefore, the main constituent (2a) was chemically transformed to the dihydroxyketone 6. The physical data, the chiroptical and spectral properties of the final product (6) corresponded to those of the known 16R,17-dihydroxyphyllocladane-3-one (calliterpenone) (6) [15–18]. Moreover, since several of the isolated compounds are correlated by interconversions, the new structures are established as 2a, 2b, 3a, 3b, 4 and 5a. A full account of the synthetic part will be published elsewhere.

EXPERIMENTAL

General. ¹H NMR: CDCl₃ at 300 or 400 MHz; ¹³C NMR: 50 MHz.

Plant material. Cuttings of P. ambiguus [2] (L.E. Codd, Acc.-Nr 411-67) from The Royal Botanical Gardens, Kew, U.K., were propagated and cultivated in the open in the surroundings of Zurich (1979). Plants from the original cuttings are kept in the greenhouse of our institute.

Extraction and isolation. Air-dried leaves and stems of P. ambiguus (350 g) were extracted with hexane (31, overnight) at room temp. and then re-extracted (×2) with Et₂O (31, 7 hr; 31, overnight) at room temp. The hexane extract was concd in vacuo to give a green semi-solid (1.55 g, 0.44%). The Et₂O extract was concd in vacuo to afford a green residue (9.01 g, 2.57%), which was partitioned between hexane (400 ml) and 90% MeOH (400 ml). The hexane layer was washed with 90% MeOH (200 ml) and the MeOH layers were combined and washed with hexane (3 × 200 ml) and then evapd in vacuo to afford a yellow solid (fr. A, 6.91 g, 1.97%). The hexane

extracts were combined and evapd in vacuo to give a green solid (fr. B, 2.02 g, 0.58%).

All frs were tested for their antioxidative activity using a modification of the method of ref. [19]. Among them, the main fr. A showed significant antioxidative activity.

Hexane-CH₂Cl₂ (1:6, 50 ml) was added to fr. A and the mixt. filtered. The filtrate was sepd on Sephadex LH-20 [increasing elution strength from hexane-CH₂Cl₂ (1:6), CH₂Cl₂, CH₂Cl₂-Me₂CO (1:1), Me₂CO] to give frs I-V. Only fr. III had significant antioxidative activity. Hexane-Et₂O (2:1, 100 ml) was added to III, which was filtered and the ppt. recrystallized from CH₂Cl₂-Me₂CO-MeOH to afford the flavone 1. The filtrate and the mother liquor displayed no antioxidative activity. They were combined and repetitive CC [silica gel, pentane-Et₂O (1:2), CH₂Cl₂-MeOH (50:1), CH₂Cl₂-MeOH (25:1)] yielded a mixt, of 2b and 3b and a solid fr. The latter was further sepd by prep. HPLC [column: Nucleosil C-18, 10 μ m, 25 × 2 cm, H₂O-MeOH (2.5:1), 17 ml min⁻¹; 70 bar] to give 2a, 3a and a mixt. of 4 and 5a, which was crystallized from Et₂O to yield 5a. The mother liquor was sepd by CC [silica gel, pentane-Et₂O (1:1), pentane- Et_2O (2:1), Et_2O to yield 4 and more 5a. CC of fr. II [silica gel, pentane, Et₂O-pentane (1:1)] afforded another portion of the mixt. of 2a and 3b. The following amounts of the pure compounds were isolated: 1 (0.092 g, 0.026%), 2a (1.940 g, 0.55%), 2b/3b (0.281 g, 0.55%)0.08%), 3a (1.015 g, 0.29%), 4 (0.364 g, 0.10%), and 5a (0.450 g, 0.13%).

5,6-Dihydroxy-4',7-dimethoxyflavone (1) (Ladanein or 4',7-dimethylscutellarein). Yellow needles (CH₂Cl₂-Me₂CO-MeOH), mp 213-215°. The spectral data (UV/VIS, IR, ¹H NMR, EI and CIMS) were in full agreement with the reported values [3-7].

(16R)-2\alpha-(3-Methyl-2-butenoyloxy)-3\alpha-Acetoxyphyllocladane-16,17-diol (2a). Needles (Et₂O-hexane), mp $88-92^{\circ}$; $[\alpha]_{D}^{25} = +5.7^{\circ}$ (c 0.575, CHCl₃); UV λ_{max}^{MeOH} nm (log ε): 218 (4.15); IR $v_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3200–3600 (br), 2940, 2870, 1746, 1718, 1650, 1452, 1375, 1240, 1142, 1075, 1048, 1032, 1010, 990; ¹H NMR: δ 0.86 (3H, s, Me-19), 0.99 (3H, s, Me-18), 1.01 (3H, s, Me-20), 1.86, 2.13 (each 3H, each d, ${}^{4}J = 1.2 \text{ Hz}$, Me-24, Me-25), 2.11 (3H, s, AcO), 3.62, 3.67 (each 1H, ABX, ${}^{2}J = 10.8, {}^{3}J_{17,OH} =$ 5.4 Hz, after adding D_2O : AB, $^2J = 10.9$ Hz, H_2 -17), 5.01 (1H, d, ${}^{3}J_{3,2} = 2.7 \text{ Hz}$, H-3), 5.25 (1H, ddd, ${}^{3}J_{2,\text{lax}}$ = 12.4, ${}^{3}J_{2,leq}$ = 4.5, ${}^{3}J_{2,3}$ = 2.7 Hz, H-2), 5.55 (1H, m, t-like, $W_{1/2}$ ca 3 Hz, H-22); ¹³C NMR: Table 1: EIMS 70 eV, m/z (rel. int.): 431 (25) [M - CH₂OH]⁺, 389 (6) $[M - CH_2OH - ketene]^+$, 362 (11): $[M - Me_2C =$ $CHCO_2H]^+$, 331 (21) $[M - CH_2OH - Me_2C]$ = $CHCO_2H]^+$, 320 (76) $[M - Me_2C] = CHCO_2H$ $- \text{ ketene}]^+, 302 (30) [M - Me_2C = CHCO_2H]$ - AcOH]⁺, 289 (20) [331 - ketene]⁺, 285 (21), 284 (22), $[302 - H₂O]^+$, 271 (100) $[289 - H₂O]^+$, 253 (53) $[271 - H_2O]^+$, 245 (11), 229 (14), 178 (10), 163 (11). Found: C, 69.91; H, 9.35. $C_{27}H_{42}O_6$ requires: C, 70.10; H, 9.15%.

(16R)-2α-(3-Methyl-2-butenoyloxy)-3α,17-diacetoxy-16-hydroxyphyllocladane (2b). A soln of 2a (30 mg) in dry pyridine and excess Ac₂O was kept overnight at room

temp. Work-up and CC [silica gel, pentane-Et₂O (1:2)] afforded **2b** (32 mg, 98%) as an oil. $[\alpha]_D^{25} + 10.0^{\circ}$ (c 1.085, CHCl₃); UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ε): 218 (4.14); IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm⁻¹: 3590, 2980, 2940, 2870, 1735 (br), 1648, 1452, 1375, 1248 (br), 1145, 1075, 1034, 990; 1 H NMR: δ 0.87 (3H, s, Me-19), 1.00 (3H, s, Me-18), 1.01 (3H, s, Me-20), 1.87, 2.14 (each 3H, each d, ${}^{4}J = 1.1$ Hz, Me-24, Me-25), 2.10, 2.11 (each 3H, each s, 2AcO), 4.16, 4.24 (each 1H, AB, ${}^{2}J = 11.3 \text{ Hz}$, H_{2} -17), 5.01 (1H, d, ${}^{3}J_{3,2} = 2.7 \text{ Hz}$, H-3), 5.25 (1H, ddd, ${}^{3}J_{2,lax} = 12.4$, ${}^{3}J_{2,leq} = 4.4$, $^{3}J_{2,3} = 2.7 \text{ Hz}, \text{ H-2}$), 5.55 (1H, m, t-like, $W_{1/2}$ ca 3 Hz, H-22); 13 C NMR: Table; CIMS (2-methylpropane), m/z(rel. int.): $505 (36) [M+1]^+$, 488 (21), 487 (100), $[M + 1 - H_2O]^+$, 463 (12) $[M + 1 - \text{ketene}]^+$, 445 (26) $[M + 1 - AcOH]^+$, 427 (10) $[M + 1 - H_2O - AcOH]^+$, 405 (34) $[M + 1 - Me_2C = CHCO_2H]^+$, 345 (47) [405]- AcOH]⁺, 327 (15) [345 - H₂O]⁺. Comparison of the R_f, UV, IR, ¹H, ¹³C NMR and MS proved this monoacetate to be identical in every respect with 2b, present in a mixt. with 3b.

Acetonide 2c. A mixt. of 2a (102 mg) in dry Me₂CO (10 ml) and CuSO₄ (150 mg) was heated under reflux for 5 hr under N₂. Work-up and CC [silica gel, hexane-CH₂Cl₂ (1:2)] afforded 2c (101 mg, 91%) as crystals: mp 70°. IR $v_{\text{max}}^{\text{KBr}}$ cm⁻¹: 2980, 2938, 2868, 1745, 1715, 1650, 1454, 1375, 1240, 1142, 1055, 1034, 988, 895, 850; ¹H NMR: δ 0.87 (3H, s, Me-19), 1.00 (3H, s, Me-18), 1.01 (3H, s, Me-20), 1.34, 1.38 (each 3H, each s, 2Me of the acetonide), 1.86, 2.13, (each 3H, each d, ${}^{4}J = 1.1$ Hz, Me-24, Me-25), 2.11 (3H, s, AcO), 2.26 (1H, dd, ${}^{2}J_{15\beta,15\alpha}$ = 14.6, ${}^{4}J_{15\beta,14\alpha}$ = 1.7 Hz, H- 15β), 3.90, 4.06 (each 1H, AB, ${}^{2}J = 8.0 \text{ Hz}$, H₂-17), 5.01 (1H, d, ${}^{3}J_{3,2} = 2.7 \text{ Hz}$, H-3), 5.25 (1H, ddd, ${}^{3}J_{2,lax} = 12.4$, ${}^{3}J_{2,leq} = 4.4$, ${}^{3}J_{2,3}$ = 2.7 Hz, H-2), 5.55 (1H, m, t-like, $W_{1/2}$ ca 3 Hz, H-22); CIMS (2-methylpropane), m/z (rel. int.): 503 (60) $[M + 1]^+$, 487 (9), 461 (10) $[M + 1 - \text{ketene}]^+$, 445 (37), 443 (15) $[M + 1 - AcOH]^+$, 403 (44) $[M + 1 - Me_2C]$ $= CHCO_2H)^+$, 386 (51), 385 (100) $[403 - H_2O]^+$, 344 (13), 343 (57) $[M + 1 - Me_2C = CHCO_2H)]^+$, 285 (36).

Oxidation of 2a to ketone 2d. A mixt. of 2a (31 mg) in MeOH (2 ml) and H₅IO₆ (17 mg) was stirred for 40 min at room temp., then poured into H2O and extracted with CH₂Cl₂. The organic phase was washed with H₂O, dried over MgSO₄, filtered, evapd and purified by CC (silica gel, CH₂Cl₂) to yield 2d (25 mg, 87%) as crystals, mp 47–50°; $[\alpha]_D + 50.4^\circ$ (c 0.28, CHCl₃); UV λ_{max}^{MeOH} nm $(\log \varepsilon)$: 218 (4.22); IR $v_{\text{max}}^{\text{KBr}}$ cm⁻¹: 2950, 2870, 1745, 1715, 1648, 1454, 1374, 1238, 1142, 1075, 1045, 1025, 1012; ¹H NMR: δ 0.91 (3H, s, Me-19), 1.02 (3H, s, Me-18), 1.03 (3H, s, Me-20), 1.88, 2.15 (each 3H, each d, ${}^4J = 1.1$ Hz, Me-24, Me-25), 2.13 (3H, s, AcO), 2.38 (1H, m, H-13), 2.66 (1H, dd, ${}^{2}J_{15\beta,15\alpha} = 18.6$, ${}^{4}J_{15\beta,14\alpha} = 3.6$ Hz, H-15 β), 5.04 (1H, d, ${}^{3}J_{3,2} = 2.7 \text{ Hz}$, H-3), 5.28 (1H, ddd, ${}^{3}J_{2,1ax} = 12.4, \ {}^{3}J_{2,1eq} = 4.4, \ {}^{3}J_{2,3} = 2.7 \text{ Hz}, \ \text{H-2}), 5.56$ (1H, m, t-like, $W_{1/2}$ ca 3 Hz, H-22); CIMS (NH₃), m/z (rel. int.): $448 (100) [M + 1 + NH_3]^+$, $406 (4) [M + 1 + NH_3]$ - ketene]+.

(16R)-2 α -(3-Methylbutanoyloxy)-3 α -acetoxyphyllocladane-16,17-diol (3a). Flakes (Et₂O-hexane), mp 35°; [α]_D⁵ + 2.5° (c 0.16, CHCl₃); IR ν_{max} KBr cm⁻¹: 3600–3200 (br), 2940, 2870, 1745 (br), 1465, 1455, 1374, 1295, 1236 (br), 1190, 1154, 1120, 1050, 1032, 1015, 988; 1 H NMR: δ 0.86 (3H, s, Me-19), 0.98 (3H, s, Me-18), 1.00 (3H, s, Me-20), 0.92, 0.93 (each 3H, each d, 3 J = 6.4 Hz, Me-24, Me-25), 2.11 (3H, s, AcO), 3.62, 3.76 (each 1H, AB, 2 J = 10.9 Hz, H₂-17), 4.97 (1H, d, 3 J_{3,2} = 2.7 Hz, H-3), 5.23 (1H, ddd, 3 J_{2,1ax} = 12.4, 3 J_{2,1eq} = 4.4, 3 J_{2,3} = 2.7 Hz, H-2); 13 C NMR: Table 1; CIMS (NH₃), m/z (rel. int.): 482 (100) [M + 1 + NH₃] +, 464 (6) [M] +, 451 (11), 450 (45) [M - CH₃OH] +, 440 (1) [M - ketene] +, 382 (14), 322 (9) [382 - AcOH] +.

Catalytic reduction of 2a. A mixt. of 2a (14 mg) and Pd/C (9 mg) in dry MeOH (3 ml) was shaken for 2 hr under H_2 at room temp. Work-up afforded a solid (13 mg, 93%) which proved to be identical in every respect with the natural product 3a (mp, mixed mp, R_f , $[\alpha]_D$, IR, ¹H, ¹³C NMR, MS).

 $(16R)-2\alpha-(3-Methylbutanoyloxy)-3\alpha,17-diacetoxy-16$ hydroxyphyllocladane (3b). A soln of 3a (10 mg) in dry pyridine and excess Ac₂O was kept overnight at room temp. Work-up and CC [silica gel, Et₂O-pentane (1:1)] afforded 3b (10 mg, 92%) as a viscous oil, $[\alpha]_D^{25} + 3.6^{\circ}$ (c 0.45, CHCl₃); IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm⁻¹: 3590, 2950, 2870, 1732, 1465, 1455, 1375, 1295, 1250, 1190, 1155, 1120, 1035, 988, 922; ¹H NMR: δ 0.87 (3H, s, Me-19), 0.94, 0.95 (each 3H, each d, ${}^{3}J = 6.3$ Hz, Me-24, Me-25), 0.99 (3H, s, Me-18), 1.01 (3H, s, Me-20), 2.12 (6H, s, 2AcO), 4.18, 4.25 (each 1H, AB, $^2J = 11.3 \text{ Hz}$, H_2 -17), 4.98 (1H, d, ${}^{3}J_{3,2} = 2.7 \text{ Hz}$, H-3), 5.25 (1H, ddd, = 12.4, ${}^{3}J_{2,1eq} = 4.4$, ${}^{3}J_{2,3} = 2.7$ Hz, H-2); ${}^{13}C$ NMR: Table 1; CIMS (NH₃), m/z (rel. int.): 524 (100) $[M + 1 + NH_3]^+$, 489 (31) $[M + 1 - H_2O]^+$, 464 (34), $[M - ketene]^+$, 429 (1) $[M + 1 - AcOH]^+$. Comparison of the R_f , ¹H, ¹³C NMR and MS proved this monoacetate to be identical in every respect with natural 3b.

(16R)-3α-Acetoxyphyllocladane-16,17-diol (4). Needles (Et_2O) , mp 122–123°; $[\alpha]_D^{25}$ – 32.3° (c 0.505, CHCl₃); IR $v_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3440, 3325, 2940, 2855, 1740, 1450, 1435, 1384, 1374, 1365, 1252, 1056, 1040, 1032, 1012, 990; ¹H NMR: $\delta 0.85$ (3H, s, Me-19), 0.89 (3H, s, Me-18), 0.91 (3H, s, Me-20), 2.08 (3H, s, AcO), 3.64, 3.78 (each 1H, ABX, $^{2}J = 10.9$, $^{3}J_{17,OH} = 5.4$ Hz, after adding $D_{2}O$: AB, $^{2}J = 10.9 \text{ Hz}, \text{ H}_{2}\text{-}17), 4.64 (1H, t, {}^{3}J_{3,2} = 2.7 \text{ Hz},$ H-3); 13 C NMR: Table 1; CIMS (NH₃) m/z: 382 (29) $[M + 1 + NH_3]^+$, 364 (19) $[M]^+$, 346 (5) $[M - H_2O]^+$, 304 (10) $[M - AcOH]^+$, 288 (21), 287 (100) $[M + 1 - H₂O - AcOH]^+,$ 273 (27), $[287 - H_2O]^+$

(16R)- 2α -(3-Methyl-2-butenoyloxy)-16,17-dihydroxy-phyllocladane-3-one (5a). Flakes (Et₂O), mp 175–177°; [α] $_{0}^{25}$ + 8.3° (c 1.20, CHCl₃). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ε): 219 (4.24); IR $\nu_{\text{max}}^{\text{KBr}}$ KBr cm $^{-1}$: 3525, 3580–3400 (br), 2930, 2860, 1720, 1705, 1648, 1480, 1458, 1390, 1380, 1235, 1158, 1125, 1075, 1068, 1050, 1023, 1008, 982, 968, 924, 870, 852; 1 H NMR: δ 1.12 (3H, s, Me-20), 1.16 (3H, s, Me-19), 1.27 (3H, s, Me-18), 1.92, 2.18 (each 3H, each d, ^{4}J = 1.2 Hz, Me-24, Me-25), 3.65, 3.80 (each 1H, ABX, ^{2}J = 10.8, $^{3}J_{17,\text{OH}}$ = 4.7 Hz, after adding D₂O: AB, ^{2}J = 10.9 Hz, H₂-17), 5.65 (1H, dd, $^{3}J_{2,\text{Lax}}$ =

13.4, ${}^{3}J_{2,1eq} = 6.1$ Hz, H-2), 5.79 (1H, m, t-like, $W_{1/2}$ ca 3 Hz, H-22); ${}^{13}C$ NMR: Table 1; CIMS (2-methylpropane), m/z (rel. int.): 419 (69) $[M+1]^{+}$, 401 (100) $[M+1-H_{2}O]^{+}$, 387 (4) $[M+1-MeOH]^{+}$, 336 (9) $[M+1-Me_{2}C=CHCO]^{+}$, 319 (12) $[M+1-Me_{2}C=CHCO_{2}H]^{+}$, 301 (40) $[M-H_{2}O-Me_{2}C=CHCO_{2}H]^{+}$, 283 (17) $[301-H_{2}O]^{+}$, 101 (6), 83 (49). Found: C, 72.02; H, 9.38. $C_{25}H_{38}O_{5}$ requires: C, 71.74; H, 9.15%.

Acetate 5b. A soln of 5a (30 mg) in dry pyridine and excess Ac₂O was kept overnight at room temp. After work-up and prep. TLC [silica gel, CH2Cl2-MeOH (25:1)], crystallization from hexane-Et₂O afforded 5b (30 mg, 91%) as plates, mp $136-138^{\circ}$; $[\alpha]_{D}^{25} + 5.9^{\circ}$ (c 0.17, CHCl₃); IR v_{max} KBr cm⁻¹: 3580, 2940, 2860, 1748, 1730, 1720, 1650, 1456, 1386, 1372, 1350, 1300, 1240 (br), 1146, 1124, 1068, 1030, 978, 918, 872, 850; ¹H NMR: δ 1.12 (3H, s, Me-20), 1.16 (3H, s, Me-19), 1.27 (3H, s, Me-18), 1.92, 2.18 (each 3H, each d, ${}^{4}J = 1.1$ Hz, Me-24, Me-25), 2.13 (3H, s, AcO), 4.19, 4.26 (each 1H, each d, AB, $^{2}J = 11.3 \text{ Hz}, \text{ H}_{2}\text{-}17), 5.65 \text{ (1H, } dd, {}^{3}J_{2,1ax} = 13.4,$ ${}^{3}J_{2,1eq} = 6.0 \text{ Hz}, \text{ H-2}, 5.79 (1H, m, t-like, } W_{1/2} ca$ 3 Hz, H-22); CIMS (NH₃), m/z (rel. int.): 478 (23) $[M + 1 + NH_3]^+$, 462 (25), 461 (100) $[M + 1]^+$, 443 (7) $[M + 1 - H_2O]^+$, 401 (7) $[M + 1 - AcOH]^+$, 378 (10) $[M + 1 + NH_3 - Me_2C = CHCO_2H]^+$, 362 (10), 361 (61) $[M + 1 - Me_2C = CH - CO_2H]^+$, 345 (11), 289 (7).

Acknowledgements—We thank Dr D. R. Hunt, The Royal Botanical Gardens at Kew, U.K., for cuttings of P. ambiguus. The work was supported by the Swiss National Foundation.

REFERENCES

- Bürgi, C. and Rüedi, P. (1993) Helv. Chim. Acta 76, 1890.
- Meyer, E. (1837) Commetariorum de Plantis Africae Australioris, p. 228; Thiselton-Dyer, T. W. (ed.). (1912) Flora Capensis, Vol. V, Sect. 1, p. 279; Codd, L. E. (1964) Bothalia 8, 159.
- 3. Markham, K. R., Hirshman, H., Kupfermann, H. and Ma, T. S. (1970) Mikrochim. Acta 590.
- Seshadri, T. R. and Sharma, P. (1973) Indian J. Chem. 11, 338.
- Okuda, T., Yoshida, I. and Ono, I. (1975) Phytochemistry 14, 1654.
- 6. Voirin, B. (1983) Phytochemistry 22, 2107.
- 7. Barberan, F. A. T., Hernandez, L., Ferreres, F. and Thomas, F. (1985) Planta Med. 51, 452.
- 8. Nomoto, K., Rüedi, P. and Eugster, C. H. (1976) *Helv. Chim. Acta* 59, 772.
- Cookson, R. C., Crabb, T. A., Frankel, J. J. and Hudec, J. (1966) Tetrahedron 22 (Suppl. 7), 355.
- Fujita, T., Takeda, Y. and Shingu, T. (1981) Heterocycles 16, 227.
- 11. Kitajima, J., Komori, T. and Kawasaki, T. (1982) *Chem. Pharm. Bull.* 30, 3912.

- Gustafson, K. R., Munro, M. H. G., Blunt, J. W., Cardellina, J. H., II, McMahon, J. B., Gulakowski, R. J., Cragg, G. M., Cox, P. A., Brinen, L. S., Clardy, J. and Boyd, M. R. (1991) Tetrahedron 47, 4547.
- Duc, D. K. M., Fétizon, M., Lazare, S., Grant, P. K., Nicholls, M. J., Liau, H. T. L., Francis, M. J., Poisson, J., Bernassau, J. M., Roque, N. F., Workulich, P. M. and Wenkert, E. (1981) Tetrahedron 37, 2371.
- Katti, S. B., Rüedi, P. and Eugster, C. H. (1982) Helv. Chim. Acta 65, 2189.
- Chatterjee, A., Desmukh, S. K. and Chandrasekharan, S. (1972) Tetrahedron 28, 4391.
- Ahmad, S. A. and Zaman, A. (1973) Tetrahedron Letters 24, 2179.
- 17. Fujita, E., Ochiai, M., Ichida, I., Chatterjee, A. and Desmukh, S. K. (1975) *Phytochemistry* 14, 2249.
- 18. Wong, W. H., Wei, C., Loke, S. E., and Mak, T. C. W. (1991) *Acta Crystallogr.* 47C, 906.
- 19. Whittern, C. C., Miller, E. E. and Pratt, D. E. (1984) J. Am. Oil. Chem. Soc. 61, 1075.