



ANNOSENEGALIN AND ANNOGALENE: TWO CYTOTOXIC MONO-TETRAHYDROFURAN ACETOGENINS FROM *ANNONA SENEGALENSIS* AND *ANNONA CHERIMOLIA*

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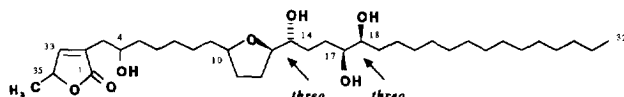
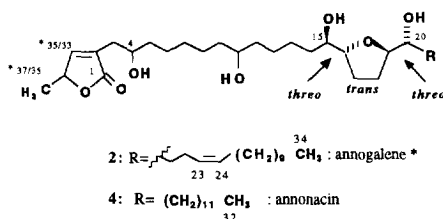
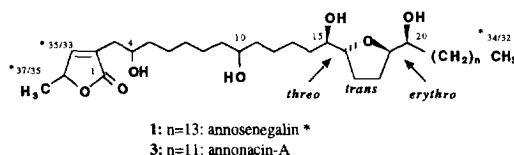
Key Word Index—*Annona senegalensis*; *A. cherimolia*; Annonaceae; acetogenins; mono-tetrahydrofuran γ -lactones; cytotoxicity.

Abstract—Two new cytotoxic mono-tetrahydrofuran acetogenins, annosenegalin and annogalene, have been isolated from the cytotoxic methanolic extract of *Annona senegalensis* and *A. cherimolia* seeds. Their structures were established on the basis of 1D and 2D NMR spectroscopic techniques. Annosenegalin belongs to the rare type of C_{37} mono-tetrahydrofuran acetogenins, and annogalene is an olefinic acetogenin.

INTRODUCTION

Annona senegalensis Pers. is a shrub native to Senegal and is used in traditional medicine [1]. In our previous study of *A. senegalensis* seeds, we isolated a new mono-tetrahydrofuran (mono-THF) γ -lactone acetogenin (senegalene) and four known bis-tetrahydrofuran (bis-THF) γ -lactone acetogenins (molvizarin,

asimicin, rolliniastatin-2 and squamocin) [2]. *Annona cherimolia* Mill. is a tropical tree native to Peru, which is used in traditional medicine as an insecticide and a parasiticide [3]. In our previous studies of *A. cherimolia* seeds, we isolated nine new bis- and mono-THF acetogenins (cherimolin-1, cherimolin-2, almunequin, itrabin, laherradurin, otivarin, jetein, molvizarin and motrillin), two known bis-THF acetogenins



(rolliniastatin-2 and squamocin) [4–9], and three 'iso'-acetogenins (isocherimolin-1, isomolvizarin-1 and isomolvizarin-2) [10]. All these compounds showed antiparasitic and cytotoxic activities [11–14]. Some of these acetogenins are very potent inhibitors of the NADH:ubiquinone oxidoreductase (complex I) activity of mammalian mitochondria [15].

We report here on the structural elucidation, on the basis of 2D NMR and mass spectroscopic methods, of two novel cytotoxic mono-THF acetogenins, annosenegalin (**1**) and annogalene (**2**), as well as the isolation of three known mono-THF acetogenins, annonacin-A (**3**), annonacin (**4**) and gigantetrocin (**5**), from *A. senegalensis* seeds, and of **1**, **3** and **5** from *A. cherimolia* seeds.

RESULTS AND DISCUSSION

The methanolic extract from *A. senegalensis* seeds showed antiparasitic activity against *Leishmania major*, *L. donovani* and *Trypanosoma brucei brucei* [14]. This extract (with 10% H₂O) was partitioned with hexane, and the aqueous methanolic fraction extracted with methyl chloride. The methylene chloride soluble portion showed antiparasitic activity (against *Leishmania* and *Trypanosoma*), and an LC₅₀ of $1.7 \times 10^{-2} \mu\text{g ml}^{-1}$ in the brine shrimp larvae (*Artemia salina*) assay. On fractionation by flash chromatography, 60H silica gel column chromatography and HPLC it gave five bioactive mono-THF γ -lactone acetogenins, two of which are new, annosenegalin (**1**) and annogalene (**2**), and three others being the known compounds annonacin-A (**3**) [16], annonacin (**4**) [17] and gigantetrocin (**5**) [18].

Compound **1** was assigned the molecular formula C₃₇H₆₈O₇ {FAB mass spectrum (+LiCl), m/z 631 [M + Li]⁺ and m/z 625 [MH]⁺}. It gave a positive reaction with Kedde reagent and an absorption at 1745 cm⁻¹ in the IR spectrum, characteristic of an α,β -unsaturated γ -lactone acetogenin [13]. The presence of four hydroxyl groups in **1** was suggested by a broad hydroxyl absorption at 3400 cm⁻¹ in the IR spectrum, the sequential loss of four molecules of H₂O

from the [MLi]⁺ (FAB mass spectrum (+LiCl)), and the formation of a tetraacetyl derivative on treatment with acetic anhydride–pyridine.

The 1D (¹H and ¹³C NMR and spin echo) and 2D homo- and hetero-nuclear correlations (COSY 45 and XH CORR) of **1** were typical of α,α' -dihydroxylated mono-THF γ -lactone acetogenins [19]. The signals for an α,β -unsaturated γ -methyl γ -lactone (4-pentanolide) system was observed in the ¹H NMR spectrum (δ 7.17, H-35; δ 5.05, H-36; δ 1.42, CH₃-37) and in the ¹³C NMR spectrum (δ 151.83, CH-35; δ 78.00, CH-36; δ 19.13, CH₃-37) of **1**. Compound **1** showed an ABX spin system characteristic of the presence of a hydroxyl group at C-4 [20]: δ 3.82/69.96 (CH-4) and δ 2.41, 2.53/33.43 (CH₂-3).

The remaining five groups associated with oxygen-bearing carbon atoms belong to a hydroxyl group in the alkyl chain, and a mono-THF α,α' -dihydroxylated system, respectively. A multiplet (1H) at δ 3.57, which correlated with a methine carbon signal at δ 71.70, was characteristic of a hydroxyl group in the alkyl chain similar to that observed for **3** at C-10 [21]. Typical resonances in the ¹H NMR spectrum about δ 3.82 and 3.40, correlated with oxygenated carbons at δ 83.24, 82.26 and 71.70 and at δ 74.31, respectively, and correspond to the rare mono-THF α,α' -dihydroxylated system with the relative configuration *threo/trans/erythro* (Fig. 1). Other examples, **3** [16] and jetein [7], a β -hydroxy- γ -methyl γ -lactone acetogenin, belong to this class. In nature, the more frequent relative configuration shown in mono-THF α,α' -dihydroxylated acetogenins is *threo/trans/threo* i.e. of the annonacin type [19].

Analysis of the mass fragmentation of **1** (Fig. 2) showed that annosenegalin possessed a similar structure to **3**, with a hydroxyl group at C-10, and a mono-THF α,α' -dihydroxylated system between C-15 and C-20. However, **1** had two more methylene groups than **3** in the alkyl chain. Annosenegalin is the first tetrahydroxylated C₃₇ mono-THF acetogenin with a *threo/trans/erythro* relative configuration.

Compound **2** was assigned the molecular formula C₃₇H₆₆O₇ {FAB mass spectrometry (+LiCl), m/z 629

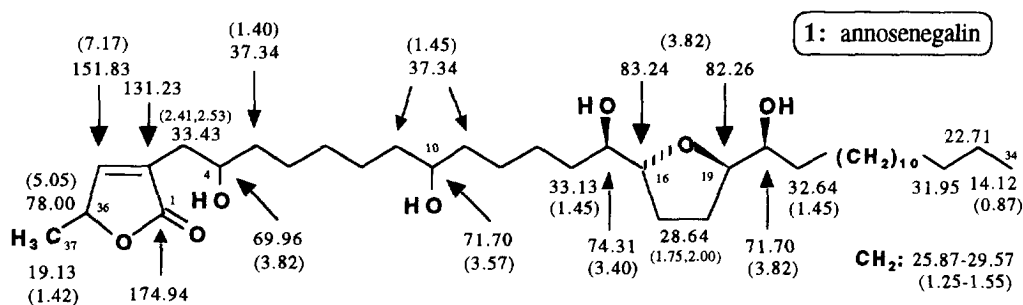


Fig. 1. Results of ¹³C NMR (¹H NMR in parentheses) and 2D NMR experiments of annosenegalin (**1**). ¹H NMR



The remaining NMR signals for **2** were characteristic of a mono-THF γ -lactone acetogenin [19]; however, two sp^2 methine resonances in the ^{13}C NMR spectrum

The position of the double bond was determined by mass spectrometry and 2D NMR (Figs 2 and 3). Fragment ions obtained by FAB mass spectrometry + Li at m/z 361 and 403 suggested the presence of a double bond between the mono-THF system and the



terminal CH₃-34. The 400 MHz COSY spectrum of **2** revealed correlations between the olefinic protons (δ 5.33 and 5.38) and two methylenes (δ 2.21 and 2.05, respectively). A multiplet at δ 2.21 was correlated with resonances at δ 1.45, and this was correlated with the methine hydroxyl carbon belonging to the mono-THF α,α' -dihydroxylated system (δ 3.38 at position 20). The other methylene multiplet, α to olefinic group (δ 2.05), was correlated with the resonances of methylenes of the alkyl chain (δ 1.3).

Thus, the double bond of **2** was unambiguously determined to be at the C-23/24 position i.e. two carbons from the mono-THF α,α' -dihydroxylated system similar to that of bullatencin [22], squamosten-A [23] and gonionenin [24]. Selective decoupling of methylenes 22 and 25 indicated that the double bond has a *cis* configuration ($J_{23,24} = 10$ Hz). Compound **2** is a new C₃₇ mono-THF acetogenin isomer to squamosten-A [23] and gonionenin [24]. Recently, an olefinic mono-THF acetogenin, xylomatenin, which is very similar to annogalene, has been reported from *Xylopia aromatica* [25].

Some of the mono-THF acetogenins isolated from *A. senegalensis* seeds (**1–5**) were isolated from *A. cherimolia* seeds: annosenegalin (**1**), annonacin-A (**3**) and gigantetrocin (**5**).

EXPERIMENTAL

Optical rotations: 589 nm; IR: film; ¹H NMR and ¹³C NMR: CDCl₃ sol; EIMS: Nermag-Sidar; FAB MS: Kratos MS-80.

Plant material. Seeds of *A. senegalensis* Pers. were obtained from ripe fruits collected in Senegal in July 1990. The seeds of *A. cherimolia* Mill. were obtained from fruits ('cherimoya') collected in December 1989 in 'cherimoyo-vale', Granada Coast (Spain). Voucher specimens are deposited in the herbarium of the Faculty of Pharmacy and Medicine of Dakar in Senegal (reference AL 282) and in the herbarium of the Department of Botany, University of Valencia, Spain (reference VF 10463), respectively.

Extraction and isolation. Dried and powdered seeds of *A. senegalensis* (2.3 kg) were macerated with MeOH at room temp. The conc MeOH extract (155 g, A) was partitioned between hexane and 10% aq MeOH. After removal of the hexane fr. (47 g, B), the aq. MeOH sol was partially evap under red. pres. and extracted with CH₂Cl₂. The CH₂Cl₂ soluble portion was dried and conc (56 g, C) and fractionated by silica gel flash chromatography (Merck 9385; elution with CH₂Cl₂-AcOEt-MeOH, 20:19:1) and by 60H silica gel CC (Merck 7736; elution with CH₂Cl₂-MeOH 24:1). Extraction and fractionation were monitored by Kedde reagent, and the brine shrimp (*Artemia salina*) test. Five bioactive mono-THF γ -lactone acetogenins were iso-

1 (5 mg), **3** (15 mg) and **5** (12 mg) were isolated from *A. cherimolia* seeds (1 kg).

Annosenegalin (1). Obtained in an amorphous form by prep. HPLC: μ -Bondapak C₁₈; MeOH-H₂O 17:3; *R*_f 24.5 min.; C₃₇H₆₈O₇; [α]_D +15° (CHCl₃; *c* 0.27); IR $\nu_{\max}$ (film) cm⁻¹: 3400, 2910, 2845, 1745; EIMS, *m/z*: 397, 379, 361, 343, 327, 309 (100%), 297, 291, 273, 241, 227, 223, 141, 111; FABMS (NBa + LiCl), *m/z*: 631 [M + Li]⁺ (100%), 625 [MH]⁺, 613 [MLi - H₂O]⁺, 595 [MLi - 2 H₂O]⁺, 577 [MLi - 3H₂O]⁺, 559 [MLi - 4H₂O]⁺ (see Fig. 2); ¹H NMR (200 MHz, CDCl₃) and ¹³C NMR (50 MHz, CDCl₃): Fig. 1.

Annogalene (2). Obtained in an amorphous form by prep HPLC: μ -Bondapak C₁₈; MeOH-H₂O 17:3; *R*_f 21.8 min.; C₃₇H₆₆O₇; [α]_D +14° (CHCl₃; *c* 0.31); IR $\nu_{\max}$ (film) cm⁻¹: 3350, 2910, 2845, 1750; EIMS, *m/z*: 397, 379, 361, 343, 327, 309 (100%), 297, 295, 291, 273, 241, 225, 223, 141, 111; FABMS (NBa + LiCl), *m/z*: 629 [M + Li]⁺ (100%), 623 [MH]⁺, 611 [MLi - H₂O]⁺, 593 [MLi - 2H₂O]⁺, 575 [MLi - 3H₂O]⁺, 559 [MLi - 4H₂O]⁺, (see Fig. 2); ¹H NMR (400 MHz, CDCl₃) and ¹³C NMR (100 MHz, CDCl₃): Fig. 3.

Annonacin-A (3). Obtained in an amorphous form by prep. HPLC: μ -Bondapak C₁₈; MeOH-H₂O 17:3; *R*_f 14.5 min.; C₃₅H₆₄O₇; IR $\nu_{\max}$ (film) cm⁻¹: 3400, 2910, 2845, 1745; FABMS (NBa + LiCl), *m/z*: 603 [M + Li]⁺ (100%), 597 [MH]⁺, 585 [MLi - H₂O]⁺, 567 [MLi - 2H₂O]⁺, 549 [MLi - 3H₂O]⁺, 531 [MLi - 4H₂O]⁺, 491, 433, 403, 391, 361; see ref. [16].

Annonacin (4). Obtained in an amorphous form by prep. HPLC: μ -Bondapak C₁₈; MeOH-H₂O 17:3; *R*_f 17.2 min; C₃₅H₆₄O₇; IR $\nu_{\max}$ (film) cm⁻¹: 3400, 2910, 2845, 1745; FABMS (NBa + LiCl), *m/z*: 603 [M + Li]⁺ (100%); see ref. [17].

Gigantetrocin (5). Obtained in an amorphous form by prep. HPLC: μ -Bondapak C₁₈; MeOH-H₂O 17:3; *R*_f 19.3 min; C₃₅H₆₄O₇; IR $\nu_{\max}$ (film) cm⁻¹: 3400, 2910, 2845, 1745; FABMS (NBa + LiCl), *m/z*: 603 [M + Li]⁺ (100%), 597 [MH]⁺, 585 [MLi - H₂O]⁺, 567 [MLi - 2H₂O]⁺, 549 [MLi - 3H₂O]⁺, 531 [MLi - 4H₂O]⁺, 491, 461, 405, 375, 317, 287; see ref. [18].

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REFERENCES

1. Kerharo, J. and Adam, J. G. (1974) *La Pharmacopée Sénégalaise traditionnelle. Plantes Médicinales et Toxiques*, pp. 147–148. Vigot, Paris.

4. Cortes, D., Ríos, J. L., Villar, A. and Valverde, S. (1984) *Tetrahedron Letters* **25**, 3199.
5. Ríos, J. L., Cortes, D. and Valverde, S. (1989) *Planta Med.* **55**, 321.
6. Cortes, D., Myint, S. H. and Hocquemiller, R. (1991) *Tetrahedron* **38**, 8195.
7. Cortes, D., Myint, S. H., Leboeuf, M. and Cavé, A. (1991) *Tetrahedron Letters* **32**, 6133.
8. Cortes, D., Myint, S. H., Dupont, B. and Davoust, D. (1993) *Phytochemistry* **32**, 1475.
9. Cortes, D., Figadère, B. and Cavé, A. (1993) *Phytochemistry* **32**, 1467.
10. Duret, P., Gromek, D., Hocquemiller, R., Cavé, A. and Cortes, D. (1994) *J. Nat. Prod.* **57**, 911.
11. Rupprecht, J. K., Hui, Y. H. and McLaughlin, J. L. (1990) *J. Nat. Prod.* **53**, 237.
12. Bories, C., Loiseau, P., Cortes, D., Myint, S. H., Hocquemiller, R., Gayral, P., Cavé, A. and Laurens, A. (1991) *Planta Med.* **57**, 434.
13. Cavé, A., Cortes, D., Figadère, B., Hocquemiller, R., Laprévote, O., Laurens, A. and Leboeuf, M. (1993) in *Recent Advances in Phytochemistry* (Downum, K. R., Romeo, J. and Stafford, H. P., eds), Vol. 27, pp. 167–202. Plenum Press, New York.
14. Sahpaz, S., Bories, Ch., Loiseau, P. M., Cortes, D., Hocquemiller, R., Laurens, A. and Cavé, A. (1994) *Planta Med.* **60**, 538.
15. Degli Esposti, M., Ghelli, A., Ratta, M., Cortes, D. and Estornell, E. (1994) *Biochem. J.* **301**, 161.
16. Lieb, F., Nonfon, M., Wachendorff-Neumann, U. and Wendisch, D. (1990) *Planta Med.* **56**, 317.
17. McCloud, T. G., Smith, D. L., Chang, C. J. and Cassady, J. M. (1987) *Experientia* **43**, 947.
18. Fang, X. P., Rupprecht, J. K., Alkofahi, A., Hui, Y. H., Liu, Y. M., Smith, D. L., Wood, K. V. and McLaughlin, J. L. (1991) *Heterocycles* **32**, 11.
19. Myint, S. H., Cortes, D., Laurens, A., Hocquemiller, R., Leboeuf, M., Cavé, A., Cotte, J. and Quéro, A. M. (1991) *Phytochemistry* **30**, 3335.
20. Cortes, D., Myint, S. H., Laurens, A., Hocquemiller, R., Leboeuf, M. and Cavé, A. (1991) *Can. J. Chem.* **69**, 8.
21. Fang, X. P., Anderson, J. E., Smith, D. L., Wood, K. V. and McLaughlin, J. L. (1992) *Heterocycles* **34**, 1075.
22. Hui, Y. H., Wood, K. V. and McLaughlin, J. L. (1992) *Nat. Toxins* **1**, 4.
23. Araya, H., Hara, N., Fujimoto, Y., Srivastava, A. and Sahai, M. (1994) *Chem. Pharm. Bull.* **42**, 388.
24. Gu, Z. M., Fang, X. P., Zeng, L., Song, R., Ng, J. H., Wood, K. V., Smith, D. L. and McLaughlin, J. L. (1994) *J. Org. Chem.* **59**, 3472.
25. Colman-Sainzarbitoria, T., Gu, Z. M. and McLaughlin, J. L. (1994) *J. Nat. Prod.* **57**, 1661.