

Phytochemistry, Vol. 42, No. 1, pp. 135–137, 1996 Copyright © 1996 Elsevier Science Ltd Printed in Great Britain. All rights reserved 0031-9422/96 \$15,00 + 0.00

A 2,5-DIMETHOXYTETRAHYDROFURAN FROM HEMEROCALLIS FULVA VAR. KWANSO

TENJI KONISHI,* TOMOHIRO INOUE,† SHIU KIYOSAWA and YASUHIRO FUJIWARA

Kyoto Pharmaceutical University, Nakauchi-cho, Misasagi Yamashina-ku, Kyoto 607, Japan; †Nihon Medi-Physics Co. Ltd, Rokutanji-cho 9-8, Nishinomiya City, Hyogo 662, Japan

(Received in revised form 10 October 1995)

Key Word Index—*Hemerocallis fulva* var. *kwanso*; Liliaceae; 2,5-dimethoxytetrahydrofuran; Ehrlich reaction; NOE.

Abstract—A 2,5-dimethoxytetrahydrofuran, fulvanol, has been extracted from *Hemerocallis fulva* var. *kwanso* along with fulvanines[‡], 2,5-dihydrofuryl- γ -lactams. The structure of fulvanol has been established as 3-hydroxymethyl-2,5-dimethoxy-3,4-dihydroxytetrahydrofuran by the ¹H and ¹³C NMR spectra. The structure of fulvanol is closely related to the branched-chain tetrofuranose apiose, occurring in the form of UDP-glycoside or as other cell components. [‡Fulvanines were first named for 2,5-dihydrofuryl- γ -lactam derivatives. The authors are concerned about possible confusion with fulvanin, cf. *J. Nat. Prod.* **56**, 527 (1993), which is a name given to a new sesquiterpene.]

INTRODUCTION

In the course of extensive studies on plant constituents which react with Ehrlich's reagent, the 2,5-dihydrofuryl- γ -lactams, fulvanine A, B, C, D and E, have been extracted from *Hemerocallis fulva* L. var. kwanso R_{EGEL} [1, 2]. Furthermore, the methanol extract from the fresh aerial part of the plant was chromatographed to give fulvanol as a minor constituent. We now report on the isolation and characterization of fulvanol.

RESULTS AND DISCUSSION

Fulvanol (1), reddish purple with the Ehrlich test, was obtained as a syrup having $[\alpha]_D = -3.6^{\circ}$ (MeOH). The ¹H NMR spectrum (CD₃OD) showed eleven non-D₂O exchangeable protons, a pattern of chemical shifts identical to that with alkoxy alkyl groups, indicating a vicinal coupling system due to two methine protons at δ 4.90 and 3.88 with a singlet methine proton at δ 4.76. and the six protons appeared as two sets of equivalent methoxy proton signals at δ 3.36 and 3.43, and two proton signals of methylene coupled each other at δ 3.57 and 3.93. Further ¹H NMR spectrum (DMSO-d_e) exhibited three hydroxy proton signals at δ 3.80, 4.27 and 5.05. In the double resonance experiment, irradiation of the signal at δ 4.72 sharpened the methylene signals at δ 3.36 and 3.43, and likewise, irradiation at δ 5.05 transformed the doublet signal at δ 3.88 from a show any changeable proton signal corresponding to the irradiation of hydroxy proton.

In general, furanosyl anomeric protons resonate between 4.9 and 5.0 ppm in glycosides, while in aldoses at 5.2-5.4 ppm. The remaining protons resonate between 3.6 and 4.4 ppm [3]. Therefore, the analogous proton shift values δ 4.76 and 4.90 refer to anomeric protons of a furanosylglycoside ring, and the presence of two methoxy functions; hence the basic structure of this compound is 2,5-dimethoxytetrahydrofuran. These results were indicated by 13C NMR spectrum in which C-2 and C-5, with attached methoxy group, appeared in the low field range at δ 110.4 and 112.3, and the residual carbon signals at δ 82.4 and 78.2 were assigned to C-3 and C-4, respectively, in regard to the orientation of exo-hydroxymethyl or hydroxy function. Accordingly, fulvanol is assumed to be 3-hydroxymethyl - 2,5 - dimethoxy - 3,4 - dihydroxytetrahydrofuran

It is known that in *erythro*-D form of apiofuranose, the methylene protons of 3-hydroxymethyl group are magnetically equivalent, but in the *threo* form, these protons are non-equivalent [4]. Therefore, the configurations of two hydroxy groups at C-3 and C-4 were presumed to be oriented *trans*, based on the geminal coupled 3-hydroxymethyl protons. Stereochemistry of 1 was determined by the nuclear Overhauser effect (NOE) data: decoupling of the methoxy protons at δ 3.36 and 3.43 gave the significant NOE on the proton at δ 4.76 and 4.90, respectively. In addition, both experi-

T. Konishi et al.

Fig. 1. Structure, NOE (a) and conformation (b) of 1.

proton at δ 3.88, and a negative NOE appeared on the proton at δ 3.57, as a result of geminal coupled relationship with the affected proton.

The tetrahydrofuran ring is flexible, and the interpretation of vicinal spin-coupling in the ring in terms of ring conformation is difficult. However, on the basis of significant information regarding apiofuranose ring conformation, some insight was obtained indicating a more *quasi-axial* orientation of 3-hydroxymethyl group and *cis*-orientation between O-1 and C-5 in L-furanoside 2T_3 conformation in these compounds [5]. The structure of 1 was analogous to α -L-threo-apiofuranose having 3-hydroxymethyl-1,2,3-trihydroxy substituent groups. Since 1 was regarded as 1,4-cis-

dimethoxy derivative of α -L-threo-apiose, and the dihedral angle, θ about 133° was calculated from $J_{\rm H4,H5}$ (4.15 Hz, trans) in "Karplus" relationship [6], 2T_3 conformation was presumed for 1 with a quasi-axial orientation of 3-hydroxymethyl group.

Fulvanol (1), is an unusual substance in that the branched-chain methyl-α-L-apioside is methoxylated at 4-position of the endomethylene. 2,5-cis-Dimethoxytetrahydrofurans have been synthesized from dihydrofuran for the study of stereochemistry of the various aldofuranose [7], but fulvanol is an unprecedented product in nature. It is noteworthy from the biochemical view that the structure of 1 is closely related to the branched aldofuranose, playing an integral role in the

Table 1. H NMR spectral data for fulvanol (1) and apiose (J_{H-H} Hz, in

Position*					Apiose (D ₂ O) ⁴	
	1 (MeOH)		1 (DMSO $-d_6$)		α-L-threo	β-D-erythro
2(4)	4.76 s	3.36 s	4.63 s	3.23 s	n.d.	3.85
		(OCH ₃)		(OCH ₃)	n.d.	4.13
3 (3)				3.80 br		
				(OH)		
4(2)	3.88 d (4.2)		3.69 br d (4.0)	5.05 br	4.03	3.88
				(OH)		
5(1)	4.90 d (4.2)	3.43 s	4.77 d (4.0)	3.31 s	5.26	5.28
		(OCH ₃)		(OCH ₃)		
6(3')	3.57 d (11.5)		3.38 br d (11.3)		3.70	3.65
	3.73 d (11.5)		3.49 br d (11.3)		3.81	

^{*}Numbering of apiose is indicated in parentheses. n.d., not determined.

Table 2. 13C NMR spectral data for fulvanol (1) and apiose4

Position*			Apios	Apiose (D ₂ O)		
	Fulv	anol (MeOH)	α-L-threo	β-D-erythro		
2(4)	110.38	55.82 (OCH ₃)	75.7	74.6		
3(3)	82.35		82.7	80.5		
4(2)	78.20		82.0	78.9		
4 / 4 3						

cell function of plants and 3-hydroxymethyl-2,5-dihydrofuranyl ring of fulvanines and oxypinnatanine in *H. fulva* var. *kwanso* [1].

EXPERIMENTAL

Optical rotation: 20°, 0.1 dm cell path length; positive FABMS: accelerating voltage 10 KV, emission current 10 mA; ¹H NMR: 300 MHz, TMS as int. standard; ¹H–¹H NOE: see ref. [8].

Isolation of fulvanol (1). The water solution of the methanol extract (98 g) from a fresh aerial part of *H. fulva* var. *kwanso* (6.5 kg) was made to flow a Diaion (HP-20) column followed by elution with methanol. The methanol eluate (9.6 g) was chromatographed on Sephadex LH-20 (MeOH) to give four fractions (A, 1.20 g; B, 1.35 g; C, 3.65 g; D, 3.80 g) [1]. Fr. A was subjected in turn to column chromatographies on silica gel (CHCl₃-MeOH-H₂O, 10:0.2:0.02; CHCl₃-MeOH, 50:1) to yield crude fulvanol, monitored by TLC with the Ehrlich reaction. The crude fr. (34 mg) was followed by recycling prep. gel partition chromatography (GS-310, MeOH) to give fulvanol (1) (5.0 mg).

Fulvanol (1). Syruplike, $[\alpha]_D = 3.64^\circ$ (0.4, MeOH).

Positive FABMS m/z: 217 [M + Na]⁺, 176 [M - H₂O]. ¹H and ¹³C NMR: Tables 1 and 2.

Acknowledgement—We thank Miss K. Oda, Kyoto Pharmaceutical University, for the FABMS data.

REFERENCES

- Inoue, T., Iwagoe, K., Konishi, T., Kiyosawa, S. and Fujiwara, Y. (1990) Chem. Pharm. Bull. 38, 3187.
- Inoue, T., Konishi, T., Kiyosawa, S. and Fujiwara, Y. (1994) Chem. Pharm. Bull. 42, 154.
- Serianni, A. S. and Barker, R. (1984) J. Org. Chem. 49, 3292.
- 4. Snyder, J. R. and Serianni, A. S. (1987) Carbohydrate Research 166, 85.
- 5. Angyal, S. J. (1979) Carbohydrate Research, 77, 37.
- 6. Karplus, M. J. (1959) Chem. Phys. 30, 11.
- Srogl, J., Janda, M. and Stibor, I. (1973) Collect. Czech. Chem. Commun. 38, 3666.
- 8. Iwagoe, K., Kakae, T., Konishi, T., Kiyosawa, S., Fujiwara, Y., Shimada, Y., Miyahara, K. and Kawasaki, T. (1989) *Chem. Pharm. Bull.* 37, 124.