



## THREE ISOCOUMARINS FROM *CORIANDRUM SATIVUM*

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**Key Word Index**—*Coriandrum sativum*; Umbelliferae; isocoumarins; coriandrones C–E.

**Abstract**—Three new isocoumarins, coriandrones C–E, were isolated from whole plants of *Coriandrum sativum* and their structures established from spectral and chemical evidence.

### INTRODUCTION

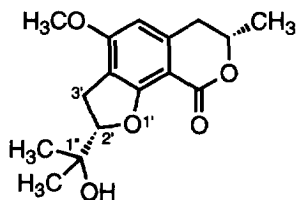
In a previous paper [1], we reported the isolation of two new isocoumarins, coriandrones A (4) and B, together with two known isocoumarins, coriandrin (5) and dihydrocoriandrin (6) [2] from the aerial parts of *Coriandrum sativum*. In the present work, we have isolated three more new isocoumarins, coriandrones C–E (1–3).

### RESULTS AND DISCUSSION

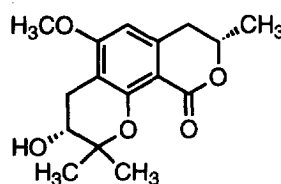
A methanolic extract of the whole plant yielded the three new compounds (1–3) after chromatographic purification, together with 20 other compounds, viz. coriandrones A (4) and B, coriandrin (5), dihydrocoriandrin (6), *p*-hydroxyphenethyl ferulate, (*R*)-(-)-4,  $\beta$ -dihydroxyphenethyl ferulate, umbelliferone, iso-

scopoletin, escletin dimethyl ether, daphnetin-8-*O*-glucoside, syringaldehyde, ferulic acid, veratric acid, *p*-hydroxycinnamic acid, *p*-hydroxybenzoic acid, 2-(4-hydroxyphenyl)-ethanol, 2-(4-hydroxyphenyl)-2-methoxyethanol, 1-(4-hydroxyphenyl)-1,2-ethanediol, kaempferol 3-*O*- $\alpha$ -L-[2,3-di-(*E*)-*p*-coumaroyl]rhamnopyranoside and kaempferol 3-*O*- $\alpha$ -L-[3-(*E*)-*p*-coumaroyl]rhamnopyranoside].

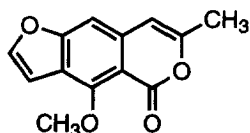
Compound 1 was isolated as needles and assigned the molecular formula  $C_{13}H_{10}O_5$  by HR mass spectrometry ( $m/z$  246.0530  $[M]^+$ ). The UV spectrum showed absorption maxima at 249.2, 277.0, 286.0, 297.7 and 334.4 nm, and the IR spectrum showed absorption bands at 3527, 3414, 3269, 1709, 1672, 1609 and 1568  $cm^{-1}$ , suggesting the presence of a hydroxyl group, an aromatic ring and an unsaturated lactone. The  $^1H$  NMR spectrum (Table 1) showed the presence of a 4,5,6-trisubstituted benzofuran ring [ $\delta$



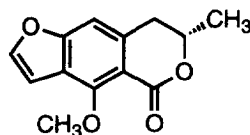
Coriandrone A (4)



Coriandrone B



Coriandrin (5)



Dihydrocoriandrin (6)

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Table 1. NMR spectral data for compounds **1** and **5** (values in parentheses are coupling constants in Hz;  $\delta$  in CDCl<sub>3</sub>, TMS)

	H		C	
	1	5	1	5
1			160.0	160.7
3			154.9	153.9
4	6.51 <i>s</i>	6.23 <i>q</i> (1.0)	103.9	104.2
4a			136.4	137.3
5	7.18 <i>d</i> (0.9)	7.10 <i>d</i> (1.0)	103.1	102.0
6			160.5	160.4
7			119.9	119.4
8			158.3	158.1
8a			107.6	107.2
2'	7.64 <i>d</i> (2.4)	7.61 <i>d</i> (2.4)	145.9	145.6
3'	7.09 <i>dd</i> (2.4, 0.9)	7.06 <i>dd</i> (2.4, 1.0)	106.4	106.2
OMe-8	4.25 <i>s</i>	4.23 <i>s</i>	61.8	61.8
CH <sub>3</sub> -3		2.25 <i>d</i> (1.0)		19.6
CH <sub>2</sub> -3	4.48 <i>s</i>		61.7	
OH	2.00 <i>br s</i>			

Assignments confirmed by <sup>1</sup>H-<sup>1</sup>H COSY, <sup>1</sup>H-<sup>13</sup>C COSY and NOE experiments.

7.64 (1H, *d*, *J* = 2.4 Hz), 7.18 (1H, *d*, *J* = 0.9 Hz) and 7.09 (1H, *dd*, *J* = 2.4 and 0.9 Hz), an olefinic proton [ $\delta$  6.51 (1H, *s*)] and a methoxyl group [ $\delta$  4.25 (3H, *s*)]. These functional groups were also identified by <sup>13</sup>C NMR (Table 1). These spectral data were closely

related to those of **5**, except for the presence of the signals assignable to a hydroxymethyl group [<sup>1</sup>H:  $\delta$  4.48 (2H, *s*), 2.00 (1H, *br s*), <sup>13</sup>C:  $\delta$  61.7] instead of signals due to a methyl group [<sup>1</sup>H:  $\delta$  2.25 (3H, *d*, *J* = 1.0 Hz), <sup>13</sup>C:  $\delta$  19.6]. In the NOE difference spectrum of **1**, on irradiation of the hydroxymethyl group at  $\delta$  4.48, a NOE was observed at the olefinic proton at  $\delta$  6.51. When the olefinic proton at  $\delta$  6.51 was irradiated, NOEs were observed at the aromatic proton at  $\delta$  7.18 and the hydroxymethyl group at  $\delta$  4.48. On irradiation of the methoxyl proton at  $\delta$  4.25, NOE was only observed at the  $\beta$ -proton of the furan ring at  $\delta$  7.09. In the coupled <sup>13</sup>C NMR spectrum of **1**, the carbonyl carbon signal at  $\delta$  160.0 was exhibited as a singlet signal. Thus, the structure of **1** was decided.

Compound **2** was isolated as prisms and assigned the molecular formula C<sub>18</sub>H<sub>24</sub>O<sub>7</sub> by HR mass spectrometry (*m/z* 352.1523 [M]<sup>+</sup>). The UV spectrum showed absorption maxima at 223.1, 272.8 and 300.0 nm, and the IR spectrum showed absorption bands at 3460, 1728, 1667, 1628, 1580 and 1520 cm<sup>-1</sup>, indicating the presence of a hydroxyl group, an aromatic ring and a lactone. The <sup>1</sup>H NMR spectrum (Table 2) exhibited signals assignable to a methoxyl group [ $\delta$  3.88 (3H, *s*)], a 2-acetoxy-3-hydroxy-3-methylbutyl group [ $\delta$  5.07 (1H, *dd*, *J* = 9.7 and 3.0 Hz), 3.06 (1H, *dd*, *J* = 14.2 and 9.7 Hz), 2.88 (1H, *dd*, *J* = 14.2 and 3.0 Hz), 1.90 (3H, *s*), 1.31 and 1.27 (each 3H, *s*)] and a hydroxyl

Table 2. NMR spectral data for compounds **2** and **4** (values in parentheses are coupling constants in Hz;  $\delta$  in CDCl<sub>3</sub>, TMS)

	H		C	
	2	4	2	4
1			170.7	164.2
3	4.67 <i>dqd</i> (10.3, 6.4, 4.5)	4.55 <i>dqd</i> (10.6, 6.3, 4.8)	75.9	74.8
4	2.93 <i>dd</i> (16.0, 10.3)	2.89 <i>dd</i> (16.1, 10.6)	35.3	36.2
	2.84 <i>dd</i> (16.0, 4.5)	2.77 <i>dd</i> (16.1, 4.8)		
4a			140.0	142.3
5	6.23 <i>s</i>	6.23 <i>s</i>	101.5	102.6
6			164.1	160.5
7			112.9	115.2
8			162.0	163.6
8a			102.3	102.5
CH <sub>3</sub> -3	1.50 <i>d</i> (6.4)	1.48 <i>d</i> (6.3)	20.9	21.0
OMe-6	3.88 <i>s</i>	3.87 <i>s</i>	56.2	55.9
OH-8	11.41 <i>s</i>			
1'	3.06 <i>dd</i> (14.2, 9.7)		23.3	
	2.88 <i>dd</i> (14.2, 3.0)			
2'	5.07 <i>dd</i> (9.7, 3.0)	4.80 <i>t</i> (9.2)	79.4	92.7
3'		3.04 <i>d</i> (9.2)	72.8	27.2
CH <sub>3</sub> CO	1.90 <i>s</i>		21.1	
CH <sub>3</sub> CO			171.3	
CH <sub>3</sub> -3'	1.31 <i>s</i>		26.7	
	1.27 <i>s</i>		25.4	
1''				71.8
CH <sub>3</sub> -1''		1.39 <i>s</i>		26.2
		1.21 <i>s</i>		23.6
OH-2''		2.75 <i>br s</i>		

Assignments confirmed by <sup>1</sup>H-<sup>1</sup>H COSY, <sup>1</sup>H-<sup>13</sup>C COSY and NOE experiments.

Table 3. NMR spectral data for compounds **3** and **6** (values in parentheses are coupling constants in Hz;  $\delta$  in CDCl<sub>3</sub>, TMS)

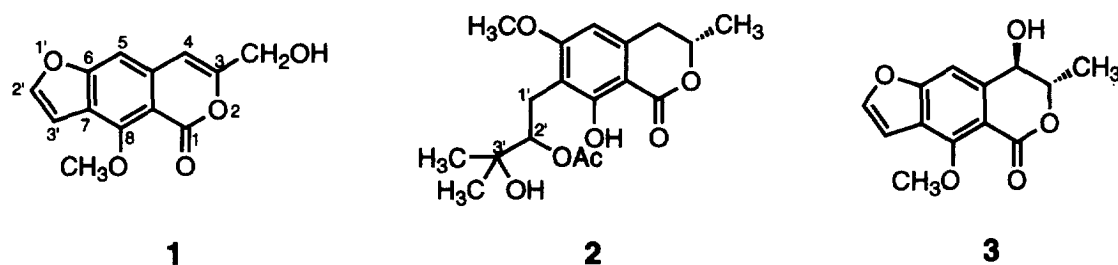
	H		C	
	3	6	3	6
1			162.3	163.7
3	4.40 <i>dq</i> (8.2, 6.4)	4.58 <i>dqd</i> (9.2, 6.4, 5.0)	79.1	74.7
4	4.67 <i>ddd</i> (8.2, 7.3, <0.1)	3.05 <i>dd</i> (16.1, 9.2)	70.8	37.0
		2.94 <i>dd</i> (16.1, 5.0)		
4a			140.9	138.2
5	7.44 <i>dd</i> (1.0, <0.1)	7.05 <i>d</i> (1.0)	103.5	105.2
6			159.8	159.3
7			120.1	119.4
8			157.8	157.8
8a			109.4	111.1
2'	7.63 <i>d</i> (2.3)	7.58 <i>d</i> (2.3)	145.7	145.1
3'	7.04 <i>dd</i> (2.3, 1.0)	7.00 <i>dd</i> (2.3, 1.0)	106.4	106.1
OMe-8	4.21 <i>s</i>	4.19 <i>s</i>	61.7	61.6
CH <sub>3</sub> -3	1.53 <i>d</i> (6.4)	1.51 <i>d</i> (6.4)	17.9	20.8
OH-4	2.36 <i>d</i> (7.3)			

Assignments confirmed by <sup>1</sup>H-<sup>1</sup>H COSY, <sup>1</sup>H-<sup>13</sup>C COSY and NOE experiments.

group chelated with a carbonyl group [ $\delta$  11.41 (1H, *s*)], in addition to signals due to a 3-methyl-6,7,8-trisubstituted 3,4-dihydro-isocoumarin [ $\delta$  6.23 (1H, *s*), 4.67 (1H, *dqd*,  $J$  = 10.3, 6.4 and 4.5 Hz), 2.93 (1H, *dd*,  $J$  = 16.0 and 10.3 Hz), 2.84 (1H, *dd*,  $J$  = 16.0 and 4.5 Hz) and 1.50 (3H, *d*,  $J$  = 6.4 Hz)] as shown in that of **4**. From the above spectral data, the structure of **2** was established. This structure was confirmed by a NOE experiment (data not shown). The absolute configuration at C-3 was concluded to be *S* from the fact that the circular dichroic spectrum of **2** showed a positive Cotton effect ascribed to the *K*-absorption band at 273 nm [1, 3-6].

Compound **3** was isolated as colourless needles and assigned the molecular formula C<sub>13</sub>H<sub>12</sub>O<sub>5</sub> by HR mass spectrometry ( $m/z$  248.0676 [M]<sup>+</sup>). The UV spectrum showed absorption maxima at 232.0, 259.0sh and 306.2 nm, and the IR spectrum showed absorption bonds at 3332, 1702, 1693, 1613, 1592, 1542 and 1477 cm<sup>-1</sup>, suggesting the presence of a hydroxyl group, an aromatic ring and a lactone. The <sup>1</sup>H NMR spectrum (Table 3) exhibited signals due to a 4,5,6-trisubstituted benzofuran ring [ $\delta$  7.63 (1H, *d*,  $J$  = 2.3 Hz), 7.44 (1H, *dd*,  $J$  = 1.0 and <0.1 Hz), and 7.04

(1H, *dd*,  $J$  = 2.3 and 1.0 Hz)], a methine proton linked to an oxygen atom [ $\delta$  4.40 (1H, *dq*,  $J$  = 8.2 and 6.4 Hz)], a methyl group [ $\delta$  1.53 (3H, *d*,  $J$  = 6.4 Hz)] and a methoxyl group [ $\delta$  4.21 (3H, *s*)]. These signals were closely related to those of **6**, except for the presence of signals assignable to a benzylmethine proton [ $\delta$  4.67 (1H, *ddd*,  $J$  = 8.2, 7.3 and <0.1 Hz)] coupled with a hydroxyl group [ $\delta$  2.36 (1H, *d*,  $J$  = 7.3 Hz)], instead of the signal due to a benzylmethylene proton [ $\delta$  3.05 (1H, *dd*,  $J$  = 16.1 and 9.2 Hz), 2.94 (1H, *dd*,  $J$  = 16.1 and 5.0 Hz)]. In the NOE difference spectrum of **3**, on irradiation of the methyl group at  $\delta$  1.53, a NOE was observed at a benzylmethine proton at  $\delta$  4.67. When the benzylmethine proton at  $\delta$  4.67 was irradiated, NOEs were observed at the aromatic proton at  $\delta$  7.44 and the methyl proton at  $\delta$  1.53. On irradiation of the methoxyl proton at  $\delta$  4.21, NOE was observed only at the  $\beta$ -proton of the furan ring at  $\delta$  7.04. Thus, the structure of **3** was decided as shown in the formula. The absolute configurations of C-3 and C-4 were concluded to be *S* and *R*, respectively, from the coupling constant between C-3 and C-4 protons ( $J$  = 8.2 Hz) in the <sup>1</sup>H NMR spectrum of **3**, and from analysis of the circular dichroic spectrum of **3** [7, 8].



## EXPERIMENTAL

**General.** Mps: uncorr. EIMS: 70 eV.  $^1\text{H}$  and  $^{13}\text{C}$  NMR: 300 and 75.4 MHz with TMS as int. standard. CC: Merck silica gel 60 (70–230 mesh) and Sephadex LH-20. TLC: Merck silica gel 60 F<sub>254</sub> (0.25 mm) and Whatman silica gel 150A PLK5F (1 mm). Spots and bands were detected by UV irradiation (254 and 365 nm).

**Plant material.** Plants of *C. sativum* L. were cultivated and collected in the botanical garden of the Osaka University of Pharmaceutical Sciences in June 1993. A voucher specimen is deposited at this university.

**Extraction and isolation.** Air-dried whole plants (26 kg) were chopped into small pieces and extracted with MeOH (600 l  $\times$  2) under reflux. The combined MeOH extracts were concd to dryness *in vacuo*. The residue obtained (3 kg) was subjected to CC on silica gel, eluting successively with *n*-hexane–EtOAc and  $\text{CHCl}_3$ –MeOH mixts of increasing polarity. The 15% EtOAc eluates were rechromatographed on silica gel with *n*-hexane–EtOAc (6:1), and Sephadex LH-20 with MeOH, to give **5** (11.9 mg) and **6** (9.4 mg). The 25% EtOAc eluates were rechromatographed on silica gel with *n*-hexane–EtOAc (4:1) followed by Sephadex LH-20 with MeOH to give **2** (2.3 mg), **3** (16.0 mg), 2-(4-hydroxyphenyl)-2-methoxyethanol (324.3 mg), *p*-hydroxyphenethyl ferulate (31.1 mg), umbelliferone (2.1 mg), isoscapoletin (2.5 mg) and 2-(4-hydroxyphenyl)-ethanol (6.4 mg). The 50% EtOAc eluates were rechromatographed on silica gel with *n*-hexane–EtOAc (1:1) and Sephadex LH-20 with MeOH to give **1** (3.2 mg), (*R*)-(-)-4, $\beta$ -hydroxyphenethyl ferulate (73.0 mg), escletin dimethyl ether (4.7 mg) and syringaldehyde (1.9 mg). The 5% MeOH eluates were rechromatographed on silica gel with  $\text{CHCl}_3$ –MeOH (20:1), after Sephadex LH-20 with MeOH, to give **4** (334.1 mg), coriandrone B (35.2 mg), kaempferol 3-*O*- $\alpha$ -L-[2,3-di-(*E*)-*p*-coumaroylrhamnopyranoside] (220.0 mg), kaempferol 3-*O*- $\alpha$ -L-[3-(*E*)-*p*-coumaroylrhamnopyranoside] (49.1 mg), 1-(4-hydroxyphenyl)-1,2-ethandiol (98.3 mg), ferulic acid (275.6 mg), veratric acid (7.0 mg), *p*-hydroxycinnamic acid (30.5 mg) and *p*-hydroxybenzoic acid (23.7 mg). The 10% MeOH eluates were rechromatographed on

silica gel with  $\text{CHCl}_3$ –MeOH (10:1), followed by Sephadex LH-20 with MeOH, to give daphnetin-8-*O*-glucoside (3.3 mg).

**Coriandrone C (1).** Needles, mp 142–143°. UV  $\lambda_{\text{max}}^{\text{MeOH}}$  nm (log  $\epsilon$ ): 334.4 (3.57), 297.7 (4.11), 286.0 (4.02), 277.0 (3.86), 249.2 (4.44). IR  $\nu_{\text{max}}^{\text{KBr}}$   $\text{cm}^{-1}$ : 3527, 3414, 3269, 1709, 1672, 1609, 1568. HR-MS  $m/z$ : 246.0530  $[\text{M}]^+$  (calc. for  $\text{C}_{13}\text{H}_{10}\text{O}_5$  246.0528).  $^1\text{H}$  and  $^{13}\text{C}$  NMR in Table 1.

**Coriandrone D (2).** Prisms, mp 172–173°. UV  $\lambda_{\text{max}}^{\text{MeOH}}$  nm (log  $\epsilon$ ): 300.0sh (3.57), 272.8 (4.06), 223.1 (4.34). IR  $\nu_{\text{max}}^{\text{KBr}}$   $\text{cm}^{-1}$ : 3460, 1728, 1667, 1628, 1580, 1520. CD (MeOH,  $c$   $2.84 \times 10^{-5}$ )  $\Delta\epsilon^{23}$  (nm): 0 (295), +1.81 (273), 0 (253), -0.43 (246), 0 (240). HR-MS  $m/z$ : 352.1523  $[\text{M}]^+$  (calc. for  $\text{C}_{18}\text{H}_{24}\text{O}_7$  352.1522).  $^1\text{H}$  and  $^{13}\text{C}$  NMR in Table 2.

**Coriandrone E (3).** Needles, mp 148–150°. UV  $\lambda_{\text{max}}^{\text{MeOH}}$  nm (log  $\epsilon$ ): 306.2 (3.33), 259.0sh (3.84), 232.0 (4.59). IR  $\nu_{\text{max}}^{\text{KBr}}$   $\text{cm}^{-1}$ : 3332, 1702, 1693, 1613, 1592, 1542, 1477. HR-MS  $m/z$ : 248.0676  $[\text{M}]^+$  (calc. for  $\text{C}_{13}\text{H}_{12}\text{O}_5$  248.0685). CD (MeOH,  $c$   $1.79 \times 10^{-4}$ )  $\Delta\epsilon^{23}$  (nm): 0 (346), +0.220 (310), +0.102 (284), +0.135 (276), +0.119 (271), +0.440 (260), +0.846 (245), +0.694 (240).  $^1\text{H}$  and  $^{13}\text{C}$  NMR in Table 3.

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