

S0031-9422(96)00111-2

TWO XANTHONES FROM ROOTS OF CRATOXYLUM FORMOSANUM

MUNEKAZU IINUMA,* HIDEKI TOSA, TETSURO ITO, TOSHIYUKI TANAKA and DOMINGO A. MADULID[‡]

Department of Pharmacognosy, Gifu Pharmaceutical University, 6-1 Mitahora-Higashi 5 chome, Gifu 502, Japan; †Department of Education, Culture and Sports, National Museum, Executive House Building, P. Burgos Street, P.O. Box 2659, Manila, Philippines

(Received 14 November 1995)

Key Word Index—*Cratoxylum formosanum*; Guttiferae; 2,7-dihydroxy-1,8-dimethoxyxanthone; 1,4,7-trihydroxy-8-methoxyxanthone; 1,4,7-trihydroxyxanthone.

Abstract—From the roots of *Cratoxylum formosanum*, two new xanthones, 2,7-dihydroxy-1,8-dimethoxyxanthone and 1,4,7-trihydroxy-8-methoxyxanthone, were isolated, in addition to seven known xanthones and two flavonoids. Among the xanthones, 1,4,7-trihydroxyxanthone was the first isolation from the natural sources. Structures were determined by spectral analyses.

INTRODUCTION

The genus *Cratoxylum* has about six species [1] which are distributed mainly in Southeast Asia. Some species have been used as traditional medicines [2] and the occurrence of flavonoids [3], triterpenoids [4] and xanthones [5] has been reported. In our search for biologically active compounds in Guttiferous plants [6–8], we now report the isolation and structural determination of two new xanthones with a simple oxygenated pattern in the roots of *C. formosanum*.

RESULTS AND DISCUSSION

Roots collected in the Philippines were dried, ground and extracted with benzene, acetone and 70% MeOH, successively. The benzene and acetone extracts were repeatedly chromatographed on silica gel and Sephadex LH-20 to give 1-7, 10 and 11 (from the acetone extract) and 8 and 9 (from the benzene extract).

Compound 1 was obtained as pale yellow needles; FeCl₃ and Gibb's tests were negative. The high resolution EI-mass spectrum showed the [M]⁺ at m/z 288.0646, corresponding to $C_{15}H_{12}O_6$. The ¹H NMR spectrum showed signals for *ortho*-coupled protons [δ 7.15 and 7.33 (d, J = 8.8 Hz)], a methoxyl group [δ 8.90 (br s)]. The IR (1660 cm⁻¹) and ¹³C NMR spectra (δ 175.9) suggested the presence of a carbonyl group. In the ¹³C NMR spectrum, signals due to quaternary carbon were significantly attenuated, indicating that 1 was a xanthone derivative with a symmetrical axis in its structure; thus, the number of hydroxyl and methoxyl groups and of hydrogens was duplicated. In the ¹³C NMR

spectrum, the methoxyl carbons appeared at δ 62.1, indicating that both *ortho*-positions of the methoxyl were substituted [9]. Furthermore, the quaternary carbons with an *O*-function were observed at δ 146.1, 147.1 and 150.8, respectively, which suggested that 1 had a 1,3,4-trioxygenated benzene ring. Thus, the structure of 1 was determined to be 2,7-dihydroxy-1,8-dimethoxyxanthone. This deduced structure was clearly supported by the HMBC spectrum (Fig. 1). To the best of our knowledge, this is the first isolation of a symmetrically substituted xanthone.

Compound 2, a yellow amorphous solid, gave a positive FeCl₃ test. The $[M]^+$ at m/z 274.0470 in the HREI-mass spectrum corresponded to the molecular formula C₁₄H₁₀O₆. The UV and IR spectra suggested that 2 was also a xanthone derivative. In the ¹H NMR spectrum the presence of three hydroxyls [δ 8.17 (2H, br s) and 12.08 (1H, s, chelated)] and a methoxyl group $[\delta \ 3.82 \ (3H, s)]$ was suggested, in addition to two sets of ortho-coupled protons [δ 6.46 and 7.12 (1H each, d, J = 8.8 Hz) and $\delta 7.15$ and 7.33 (1H each, d, J =9.2 Hz)]. In the ¹³C NMR spectrum, the methoxyl group appeared at δ 62.4, indicating that both orthopositions of the methoxyl group were occupied by substituents. All protonated carbons were assigned from the CH COSY spectrum (Table 1). In the HMBC spectrum of 2 (Fig. 1), the chelated hydroxyl group was correlated to an aromatic carbon at δ 109.5, which gave a cross-peak to the *ortho*-coupled proton at δ 6.46 in the CH COSY spectrum. These results indicated the partial structure of 2 to be a 1,4-dihydroxyxanthone derivative. The positions of other substituents were determined as follows. In the 13C NMR spectrum, aromatic carbons with an O-function were observed at δ 146.3, 147.8 and 151.5, respectively, which suggested the presence of another 1,3,4-trioxygenated

^{*}Author to whom correspondence should be addressed.

1196 M. IINUMA et al.

Table 1. ¹³C NMR spectral data of compounds 1-3

CNo.	1	2	3
1	146.1	155.0	154.7
2	147.1	109.5	109.3
3	123.4	123.8	124.0
4	113.8	137.7	138.1
5	113.8	114.6	120.3
6	123.4	125.3	126.1
7	147.1	147.8	155.0
8	146.1	146.3	109.3
9	175.9	183.1	183.0
4a	150.8	144.6	145.2
8a	117.7	116.2	121.9
9a	117.7	110.1	109.5
10a	150.8	151.5	151.0
OMe	62.1	62.4	

All carbons assigned with the aid of HMQC and HMBC spectra.

benzene ring. When the chemical shift of the methoxyl group in the ¹³C NMR spectrum was taken into account, the structure of **2** was elucidated to be 1,4,7-trihydroxy-8-methoxyxanthone. This structure was supported by the correlations observed in the HMBC spectrum (Fig. 1).

Compound 3, a yellow amorphous solid, also reacted positively with FeCl₃. The HREI-mass spectrum showed the [M]⁺ at m/z 244.0358, which corresponds to C₁₃H₈O₅. UV and IR absorptions showed that 3 was a xanthone. In the ¹H NMR spectrum, the presence of three hydroxyl groups [δ 8.35, 8.92 (1H each, br s) and 11.93 (1H, s, chelated)] and ortho-coupled protons [δ 6.62 and 7.29 (1H each, d, J = 8.8 Hz)], in addition to a 1,3,4-trisubstituted benzene ring [δ 7.42 (1H, dd, J = 8.7, 3.0 Hz), 7.52 (1H, d, J = 8.7 Hz) and 7.61 (1H, d, J = 3.0 Hz)] were detected. In the HMBC spectrum (Fig. 1), the chelated hydroxyl group gave a cross-peak

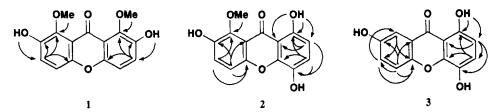


Fig. 1. HMBC spectra (J = 10 Hz) of compounds 1-3.

with an aromatic carbon at δ 109.3. In the CH COSY spectrum, this aromatic carbon was correlated to one of the *ortho*-coupled protons (δ 6.62). These results indicated that **3** was a 1,4-dihydroxyxanthone derivative. The position of remaining hydroxyl group was determined as follows. Comparison of the ¹H and ¹³C NMR spectral data with those of **5** (1,7-dihydroxyxanthone) isolated from *Harungana madagascariensis* [6]; both spectral data based on a 7-hydroxyxanthone moiety were superimposable on each other. Thus, the structure of **3** was concluded to be 1,4,7-trihydroxyxanthone, which was further supported by the HMBC spectrum (Fig. 1). Although this xanthone was previously synthesized [10], this is the first report of its occurrence as a natural compound.

Compound 7 was a xanthone with two hydroxyls and two methoxyl groups. The structure was considered to be either 3,8-dihydroxy-1,2-dimethoxy- or 1,7dihydroxy-5,6-dimethoxyxanthone. To clarify the substitution, 7 was derivatised to form a permethyl ether (7a), because 1,2,3,8-tetramethoxy- and 1,5,6,7-tetramethoxyxanthone (13) are available. The latter xanthone (13) was derived from 1,5,6-trihydroxy-7methoxyxanthone isolated from H. madagascariensis [6], the former prepared synthetically by condensation of 2,6-dimethoxybenzoic acid and 3,4,5-trimethoxyphenol. The ¹H NMR spectral data of 7a was identical to the former. Therefore, the structure of 7 was 3,8dihydroxy-1,2-dimethoxyxanthone which has previously been isolated from roots of Polygala nvikensis (Polygalaceae) [11].

Compounds 4-6 and 8-11 were identified as 1,7-dihydroxy-4-methoxy- (4), 1,7-dihydroxy- (5), 1,2,3,4,8-pentamethoxyxanthone (6), macluraxanthone (8), 1,7-dihydroxy-8-methoxyxanthone (9), (-)-epicatechin (10) and astilbin (11), respectively, by spectroscopic analysis.

EXPERIMENTAL

Plant material. Roots of *C. formosanum* (Jack) Dyer were collected in the Philippines, in August, 1993. Voucher specimens are deposited in the Philippine National Herbarium and the herbarium of Gifu Pharmaceutical University.

Extraction and isolation. Dried and ground roots (915 g) were extracted under reflux with benzene (21×12 hr × 3) (which is a considered after removal of solvent, 15 g), acetone (21×12 hr × 3) (16 g) and 70% MeOH (21×12 hr × 3) (48 g), successively. The benzene extract (7 g) was chromatographed on silica gel eluted with a benzene–Me₂CO system. The benzene eluent was recrystallized from n-hexane to give 8 (11 mg). The benzene–Me₂CO (10:1) eluent was further chromatographed on Sephadex LH-20 eluting with CHCl₃–Me₂CO (1:1) to give 4 frs. The fourth fr. was subjected to prep. TLC with n-hexane–EtOH (20:1) to give 9 (2 mg). The Me₂CO extract (14 g) was subjected to silica gel CC eluting with a benzene–Me₂CO system to give 9 frs (frs 1–9). Fr. 3 was further chromato-

graphed on Sephadex LH-20 eluting with MeOH to give 4 frs. The fourth fr. was recrystallized from benzene-Me, CO to give 4 (10 mg). After the amorphous crystals were filtered off, the filtrate was purified using prep. TLC with benzene-Me, CO (10:1) to give 5 (10 mg). Fr. 4 was further chromatographed on Sephadex LH-20 eluting with CHCl₃-Me₂CO (1:1) and prep. TLC with n-hexane-EtOAc-MeOH (8:2:1) to give 7 (5 mg). Fr. 5 was purified by Sephadex LH-20 using CHCl₃-Me₃CO (1:1) to give 5 frs. The first and fifth frs were subjected to prep. TLC usng n-hexane-EtOAc (5:1) and CHCl₃-MeOH (20:1), respectively, to give 6 (8 mg) (from the first fr.) and 2 (8 mg), 3 (5 mg) (from the fifth fr.). Fr. 7 was recrystallized from benzene-Me, CO to give 9 (30 mg). After the amorphous precipitations were filtered off, the filtrate was chromatographed on Sephadex LH-20 with CHCl₃- Me_2CO (1:1) to give 1 (10 mg). Fr. 8 was subjected to vacuum liquid chromatography on silica gel using a CHCl₃-MeOH system. From the CHCl₃-MeOH (10:1) eluent, 10 (220 mg) was obtained.

Compound 1 (2,7-dihydroxy-1,8-dimethoxyxanthone). Mp 243–244° (n-hexane–EtOAc), pale yellow needles. HREI-MS m/z 288.0646 for $C_{15}H_{12}O_6$ (calcd 288.0634); EI-MS m/z (rel. int.): 288 [M] $^+$ (100), 273 (22), 245 (25), 213 (20), 202 (14), 149 (9), 123 (5), 79 (4). UV λ (nm, MeOH): 207, 235 sh, 250 sh, 282, 305 sh, 350. IR ν (cm $^{-1}$, KBr): 3380, 1660, 1623. 1 H NMR (400 MHz, acetone- d_6): δ 3.94 (6H, s, OMe-C-1, 8), 7.15 (2H, d, J = 8.8 Hz, H-4, 5), 7.33 (2H, d, J = 8.8 Hz, H-3, 6), 8.00 (2H, br s, OH-C-2, 7).

Compound 2 (1,4,7-trihydroxy-8-methoxyxanthone). Yellow amorphous. HREI-MS m/z 274.0470 for $C_{14}H_{10}O_6$ (calcd 274.0477); EI-MS m/z (rel. int.): 274 [M] $^-$ (100), 256 (81), 244 (9), 231 (53), 228 (17), 200 (8), 149 (5), 123 (5), 102 (5), 79 (4). UV λ (nm, MeOH): 205, 238, 269, 333, 410. IR ν (cm $^{-1}$, KBr): 3390, 1650, 1615, 1600. 1 H NMR (400 MHz, acetone- d_6): δ 3.82 (3H, s, OMe-C-8), 6.46 (1H, d, J = 8.8 Hz, H-2), 7.12 (1H, d, J = 8.8 Hz, H-3), 7.15 (1H, d, J = 9.2 Hz, H-5), 7.33 (1H, d, J = 9.2 Hz, H-6), 8.17 (2H, br s, OH-C-4, 7), 12.08 (1H, s, OH-C-1).

Compound 3 (1,4,7-trihydroxyxanthone). Yellow amorphous. HREI-MS m/z 244.0358 for $C_{13}H_8O_5$ (calcd 244.0372); EI-MS m/z (rel. int.): 244 [M]⁺ (100), 243 (14), 187 (5), 131 (5). UV λ (nm, MeOH): 235, 268. IR ν (cm⁻¹, KBr): 3390, 1655, 1615, 1600. ¹H NMR (400 MHz, acetone- d_6): δ 6.62 (1H, d, J = 8.8 Hz, H-2), 7.29 (1H, d, J = 8.8 Hz, H-3), 7.42 (1H, dd, J = 8.7, 3.0 Hz, H-6), 7.52 (1H, d, J = 8.7 Hz, H-5), 7.61 (1H, d, J = 3.0 Hz, H-8), 8.35, 8.92 (1H, br s, OH-C-4, 7), 11.93 (1H, s, OH-C-1).

Methylation of 7. Compound 7 (2 mg) was methylated with MeI (0.5 ml) and K_2CO_3 (1 g) in Me₂CO (5 ml) under reflux for 5 hr. The reaction mixt. was poured into dil. HCl (200 ml) and extracted with EtOAc (20 ml). After evapn of EtOAc, the residue was purified by prep. TLC (n-hexane-EtOAc-MeOH, 8:2:1) to give 1,2,3,8-tetramethoxyxanthone (7a) (1 mg).

1198 M. IINUMA et al.

Synthesis of 7a. 2,6-Dimethoxybenzoic acid (500 mg) was dissolved in SOCl₂ (1.5 g) and heated under reflux for 2 hr. After evapn of SOCl₂, the reaction mixt. was dissolved in dry Et₂O (200 ml) and 3,4,5-trimethoxyphenol (500 mg) and AlCl, (1 g) added. After the mixt. was stirred at room temp. for 48 hr, the solvent was evapd under red. pres. and the residue was poured into dil. HCl. The acidified suspension was extracted with EtOAc and solvent evapd. The resulting mixt. was subjected to silica gel CC eluting with benzene-Me₂CO (50:1) to give 2-hydroxy-4,5,6,2',6'pentamethoxybenzophenone (12) (680 mg). Pale yellow amorphous. EI-MS m/z (rel. int.): 348 [M] (51), 317 (98), 287 (11), 210 (100), 195 (81), 167 (40), 165 (14). ¹H NMR (400 MHz, acetone- d_6): δ 3.31 (3H, s, OMe-C-5), 3.64 (3H, s, OMe-C-6), 3.73 (6H, s, OMe-C-2',6'), 3.93 (3H, s, OMe-C-4), 6.32 (1H, s, H-3), 6.70 (2H, d, J = 8.3 Hz, H-3',5'), 7.30 (1H, t, J =8.3 Hz, H-4'), 13.39 (1H, s, OH-C-2). A part of 12 (100 mg) was treated with 20% NaOH (10 ml) in MeOH (5 ml) under reflux for 10 hr. Usual work-up of the reaction mixt. gave 7a (70 mg). Colourless oil. EI-MS m/z (rel. int.): 316 [M]⁺ (28), 301 (100), 258 (22), 243 (10), 151 (7). UV λ (nm, MeOH): 220, 247, 253 sh, 293, 344. IR ν (cm⁻¹, KBr): 2950, 1655, 1605, 1595. H NMR (400 MHz, acetone- d_6): δ 3.81 (3H, s, OMe-C-2), 3.92 (3H, s, OMe-C-8), 3.93 (3H, s, OMe-C-1), 3.99 (3H, s, OMe-C-3), 6.78 (1H, s, H-4), 6.89 (1H, br d, J = 8.3 Hz, H-7), 6.94 (1H, br d, J = 8.3 Hz,H-5), 7.58 (1H, t, J = 8.3 Hz, H-6).

Methylation of 1,5,6-trihydroxy-7-methoxyxanthone. 1,5,6-Trihydroxy-7-methoxyxanthone (5 mg), isolated from the roots of H. madagascariensis [6], was methylated with MeI (1 ml) and K_2CO_3 (1 g) in Me₂CO (10 ml) under reflux for 5 hr. The reaction

mixt. was purified by prep. TLC (n-hexane-EtOAc-MeOH, 8:2:1) to give 1,5,6,7-tetramethoxyxanthone (13) (4 mg). Colourless amorphous. 1 H NMR (400 MHz, acetone- d_6): δ 3.94, 3.95, 3.98 and 4.04 (3H each, s, OMe-C-1,5,6,7), 6.95 (1H, br d, J = 8.3 Hz, H-2), 7.14 (1H, br d, J = 8.3 Hz, H-4), 7.52 (1H, d, J = 8.7 Hz, H-5), 7.39 (1H, s, H-8), 7.68 (1H, t, t = 8.3 Hz, H-3).

REFERENCES

- Bennet, G. J. and Lee, H. (1989) Phytochemistry 28, 967.
- Quisumbung, E. (1978) Medicinal Plants of the Philippines, p. 620. JMC Press, Quezon City, Philippines.
- 3. Kitanov, G., Assenov, I. and Dam, T. V. (1988) *Pharmazie* **43**, 879.
- Bennet, G. J., Harrison, L. J., Sia, G. and Sim, K. (1993) Phytochemistry 32, 1245.
- Sia, G., Bennet, G. J., Harrison, L. J. and Sim, K. (1995) *Phytochemistry* 38, 1521.
- 6. Iinuma, M., Tosa, H., Tanaka, T. and Yonemori, S. (1995) *Phytochemistry* 38, 725.
- Iinuma, M., Tosa, H., Ito, T., Tanaka, T. and Aqil, M. (1995) Phytochemistry 40, 267.
- 8. Iinuma, M., Tosa, H., Tanaka, T., Asai, F. and Shimano, R. (1995) *Phytochemistry* 39, 945.
- Miura, I., Hostettmann, K. and Nakanishi, K. (1978) Nouv. J. Chim. 2, 653.
- 10. Mahfouz, N. M. A., Hambloch, H., Omar, N. M. and Frahm, A. W. (1990) *Arch. Pharm.* **323**, 163.
- Marston, A., Hamburger, M., Sordat-Diserens, I., Msonthi, J. D. and Hostettmann, K. (1993) *Phyto-chemistry* 33, 809.