



CERCOSPORA BETICOLA TOXINS. DETERMINATION OF O_2^- · SCAVENGING ACTIVITY OF BETICOLIN-1*

CHRISTINE RUSTÉRUCCI, MARIE-LOUISE MILAT† and JEAN-PIERRE BLEIN

Laboratoire de Phytopharmacie et de Biochimie des Interactions Cellulaires, UA INRA-Université de Bourgogne, BV 1540, 21034 Dijon Cedex, France

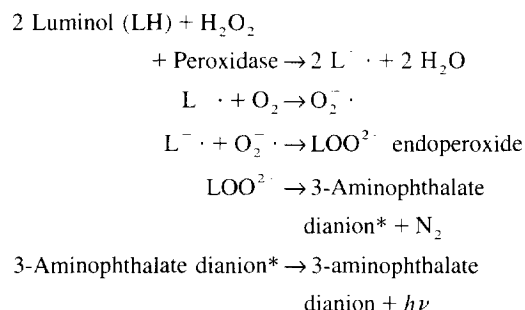
(Received in revised form 8 January 1996)

Key Word Index—*Cercospora beticola*; fungus; beticolin; superoxide anion (O_2^- ·); guaiacol; tiron; luminol; vitamin E.

Abstract—The O_2^- · scavenging properties of beticolin-1, a fungal toxin, have been studied using peroxidase-mediated luminol chemiluminescence. A comparison between beticolin-1, vitamin E and tiron is reported: beticolin-1 displays an anti-radical effect without inhibiting peroxidase activity in a larger range of concentrations (1×10^{-8} – 1×10^{-5} M) than vitamin E (3×10^{-7} – 1×10^{-5} M) or tiron (5×10^{-7} – 4×10^{-5} M). Maximal scavenging efficacy was higher for beticolin-1 and vitamin E than for tiron (88, 80 and 52%, respectively).

INTRODUCTION

Beticolins are yellow toxins produced by a phytopathogenic fungus, *Cercospora beticola*. They are constituted of partially hydrogenated anthraquinone and xanthone moieties (Fig. 1) [2–6]. Many chemical compounds containing phenolic groups are known for their antiradical activities [7–9]. This feature led us to search for a potential O_2^- · scavenging activity of beticolins. The peroxidase-mediated luminol chemiluminescence was used, since superoxide anions are generated through this reaction. This method is widely documented for determination of active oxygen species and the following scheme for the reaction has been proposed [10, 11]:



However, in such an experiment, a decrease in luminescence could also result either from an inhibition of peroxidase activity or from metabolism of H_2O_2 (catalase-like activity). In order to rule out these two

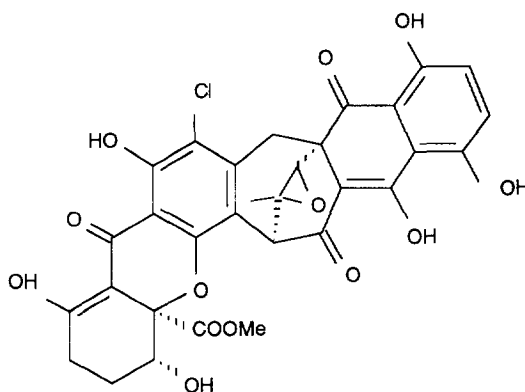


Fig. 1. Structure of beticolin-1.

possibilities and to point out the scavenging activity of a given compound, a simultaneous assay based on peroxidase-dependent guaiacol oxidation by H_2O_2 was performed [12]. In these conditions, guaiacol is oxidized in tetraguaiacol (determined spectrophotometrically) without involvement of O_2^- ·. Thus, an O_2^- · scavenger would be characterized by its inhibitory action of the luminol oxidation without any effect on the guaiacol one.

Using these dual experimental approaches, beticolin-1, one of the major compounds present in the fungal *C. beticola* extracts, appeared as an effective anti-radical molecule. Moreover, a comparison with two other antioxidants, vitamin E (presenting a large spectrum of anti-radical activity [13, 14]) and tiron (an O_2^- ·-specific scavenger [15, 16]) is reported.

*Part XIII in the series '*Cercospora beticola* Toxins'. For Part XII see ref. [1].

†Author to whom correspondence should be addressed.

RESULTS AND DISCUSSION

The effects of beticolin-1, vitamin E and tiron on the luminol oxidation (open symbols) and on the guaiacol oxidation (closed symbols) are shown in Figs. 2, 3 and 4, respectively. Results are expressed as percentage of respective controls.

Up to $ca\ 1 \times 10^{-8}$ M, beticolin-1 had no effect either on the luminol response or on the guaiacol one. At higher concentrations (1×10^{-8} – 1×10^{-5} M), beticolin-1 did not affect significantly the guaiacol oxidation whereas the luminol response was inhibited from 100 to 10% in a dose-dependent manner (Fig. 2). Between 1×10^{-9} and 3×10^{-6} M vitamin E concentrations, no change in guaiacol oxidation was detected. Between 3×10^{-7} to 3×10^{-6} M, vitamin E decreased luminol oxidation up to 20% of the control level (Fig. 3). Curves obtained with tiron (Fig. 4) presented similar shapes as the ones plotted in Fig. 3. However, the inhibition of guaiacol oxidation was noticeable from 1×10^{-5} M tiron concentration whereas luminol oxidation started to decrease from 5×10^{-7} M tiron concentration.

All these data show that beticolin-1, vitamin E and tiron exhibit an $O_2^{\cdot -}$ scavenging activity. In order to assess more accurately the potential scavenging activity of these three compounds, we plotted the curves of scavenging efficacy (Fig. 5) found by subtracting the values obtained by chemiluminescence from those obtained with guaiacol for each effector concentration. Two parameters could be analysed: the maximal efficacy and the concentration ranges where scavenging activity is observed without inhibiting peroxidase activity. First, the maximal efficacy for beticolin-1 and vitamin E were similar (88 and 80%, respectively) and better than that of tiron (52%). Moreover, these maxima were obtained for 1×10^{-5} M for beticolin-1 and vitamin E, and for 4×10^{-5} M for tiron. Second, the concentration ranges where scavenging activities are observed without inhibiting peroxidase activity are 1×10^{-8} – 1×10^{-5} M, 3×10^{-7} – 1×10^{-5} M and 5×10^{-7} – 4×10^{-5} M for beticolin-1, vitamin E and tiron, respectively (Fig. 5).

This comparison shows that antioxidant activity of beticolin-1 occurs at lower concentration than for

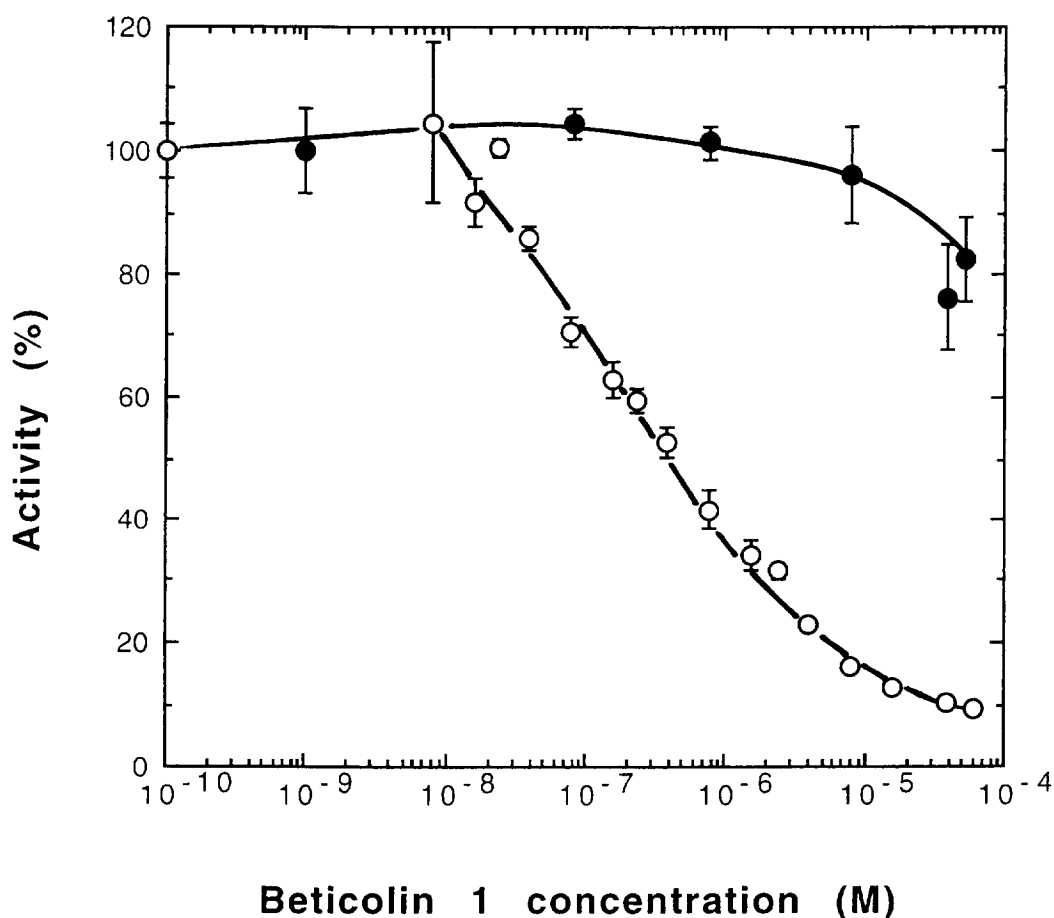


Fig. 2. Effect of beticolin-1 concentration on the peroxidase-mediated luminol chemiluminescence (○) and peroxidase activity (●) expressed as a percentage of the control containing the same concentration of DMSO (each point is the mean of three replicates).

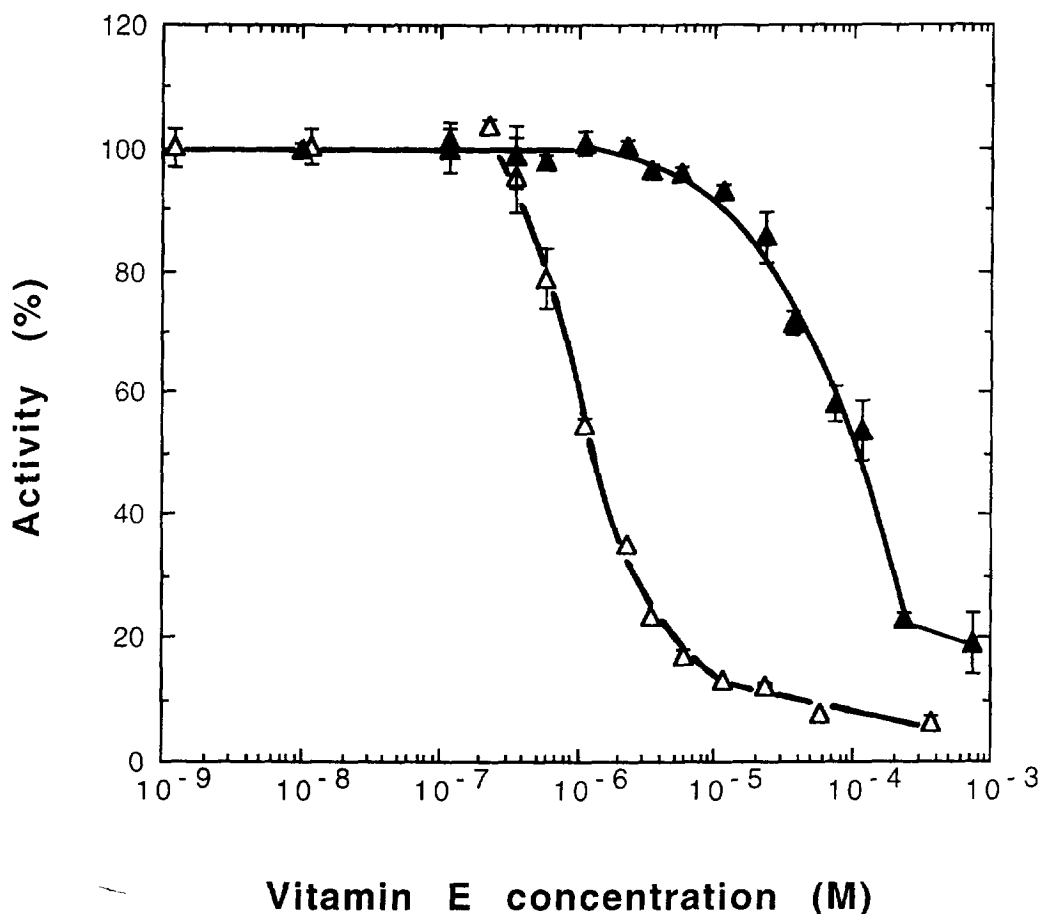


Fig. 3. Effect of vitamin E concentration on the peroxidase-mediated luminol chemiluminescence (Δ) and peroxidase activity (▲) expressed as a percentage of the control containing the same concentration of DMSO (each point is the mean of three replicates).

vitamin E and tiron. Moreover, the concentration range usable for scavenging activity is wider and the maximal scavenging activity higher for beticolin-1 than that for vitamin E or tiron. It was surprising to see that tiron in the millimolar range displays a low scavenging activity, concentrations which have been used to assess O₂⁻ production in biological systems [17, 18]. Peroxidase-mediated luminol chemiluminescence is a rapid and convenient method to analyse O₂⁻ production but, in order to search for the scavenging activity of a given compound, its effect on peroxidase activity has to be controlled.

EXPERIMENTAL

Reagents and chemicals. Peroxidase (EC 1.11.1.7) from horseradish (HRP), type VI A, 1100 diammonium salt substrate units mg⁻¹ solid, luminol (5-amino-2,3-dihydro-1,4-phthalazinedione), guaiacol (2-methoxyphenol) and vitamin E (dl-α-tocopherol) were obtained from Sigma. H₂O₂ (9% soln) was supplied by Gifrer Barbezat (France). Beticolin-1 was purified from a

mycelial extract of a *C. beticola* strain (CM) according to ref. [19].

Chemiluminescence (CL). This was measured with a Beckman LS 6000 TA scintillation counter in a single photon mode. HRP, H₂O₂ and tiron were dissolved in H₂O (0.45 μg ml⁻¹, 20 and 10 mM, respectively) and kept at 4°. Luminol (3 mM), vitamin E (0.23 mM) and beticolin-1 (0.16 mM) were dissolved in DMSO/H₂O. The final DMSO concn did not exceed 1%. Appropriate controls were run.

In the dark, in a scintillation vial containing 10 μl luminol, 500 μl MES buffer (175 mM mannitol, 0.5 mM CaCl₂, 0.5 mM K₂SO₄, 10 mM MES, pH 6.5), 50 μl H₂O₂ and 50 μl of the chemical assayed (beticolin-1, vitamin E or tiron), 50 μl of HRP were added. The final vol. was adjusted to 1 ml with H₂O. The pH of the final reaction mixt. was 6.5. CL measurements were recorded 40 sec after addition of HRP.

Guaiacol peroxidase assays. Measurements of *A* were performed with a Beckman DU 7400 spectrophotometer. Guaiacol (100 mM) was dissolved in DMSO/H₂O in such conditions that the final concn of

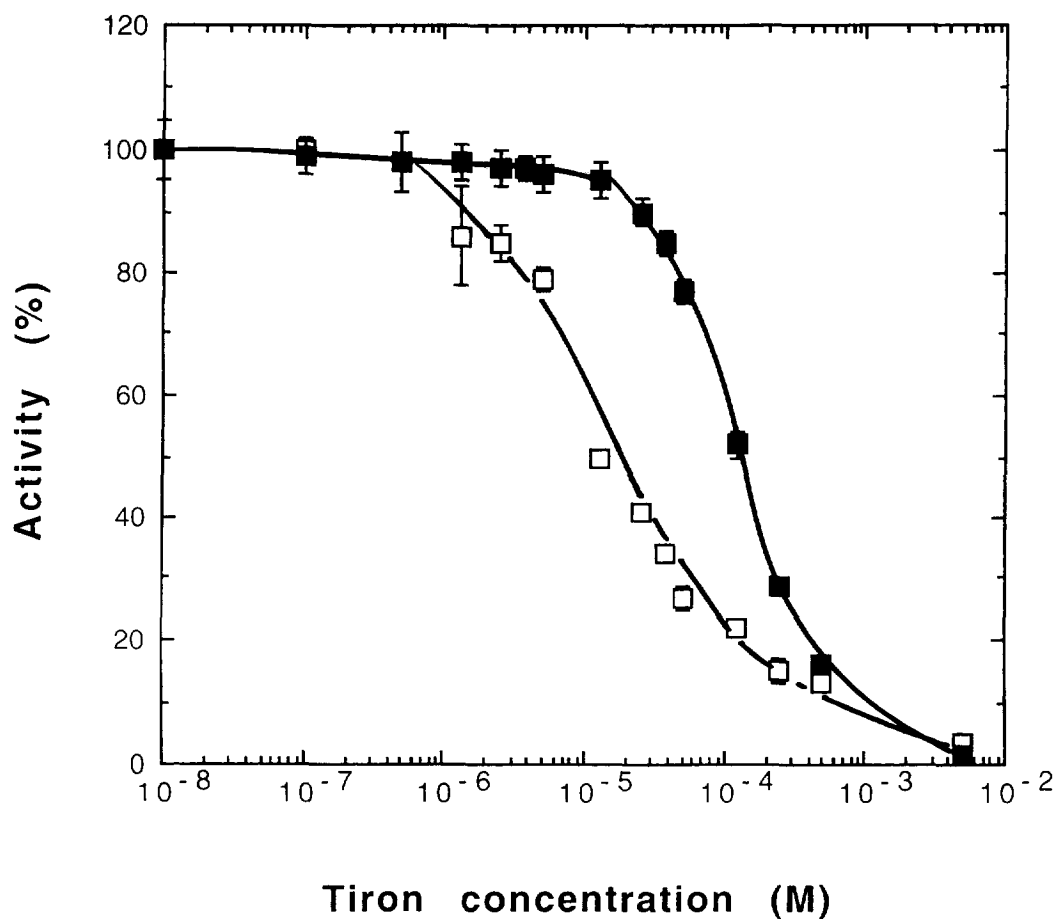


Fig. 4. Effect of tiron concentration on the peroxidase-mediated luminol chemiluminescence (□) and peroxidase activity (■) expressed as a percentage of the control containing the same concentration of DMSO (each point is the mean of three replicates).

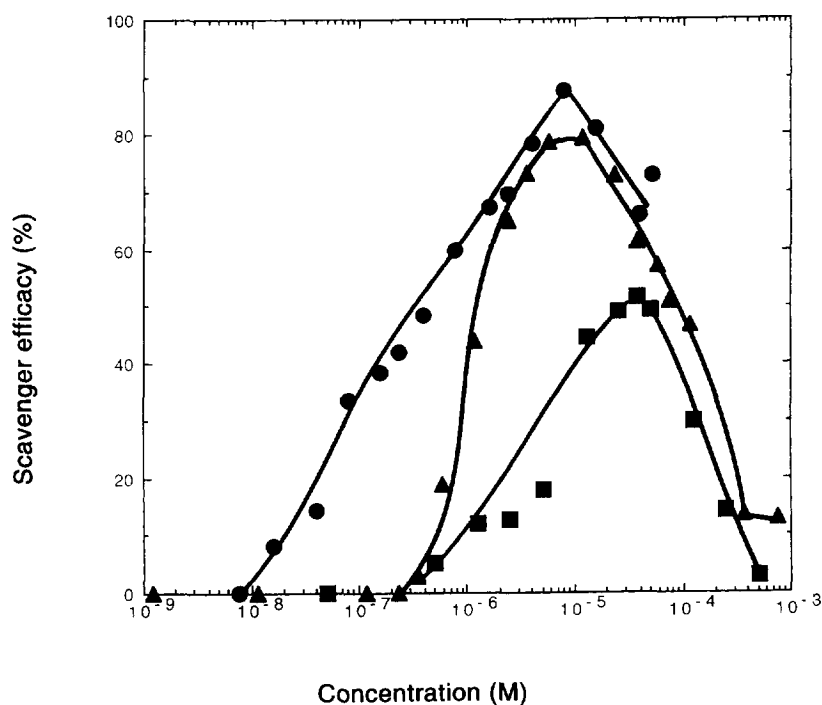


Fig. 5. Scavenging efficacy of beticolin-1 (●), vitamin E (▲) and tiron (■). Each plot was obtained by subtracting the value obtained by chemiluminescence assay from that obtained by guaiacol oxidation for each concentration assayed (open and closed symbols of Figs 2–4).

DMSO did not exceed 1%. In the dark, in the cuvette containing 50 μ l guaiacol, 500 μ l MES buffer, 50 μ l H₂O₂ and 50 μ l chemical assayed (beticolin-1, vitamin E or tiron), 50 μ l HRP were added. The final vol. was adjusted to 1 ml with H₂O. Oxidation of guaiacol was determined by measuring A at 470 nm, 10 min after HRP addition. Controls with and without DMSO have been performed.

Acknowledgements—We thank C. Descoins for his encouragement and the Conseil Régional de Bourgogne for financial support.

REFERENCES

1. Dinj, G.-Q., Maume, G., Milat, M.-L., Humbert, C., Blein, J.-P. and Maume, B. F. (1996) *Cell Biol. Int.* (in press).
2. Milat, M.-L., Prangé, T., Ducrot, P.-H., Tabet, J.-C., Einhorn, J., Blein, J.-P. and Lallemand, J.-Y. (1992) *J. Am. Chem. Soc.* **114**, 1478.
3. Milat, M.-L., Blein, J.-P., Einhorn, J., Tabet, J.-C., Ducrot, P.-H. and Lallemand, J.-Y. (1993) *Tetrahedron Letters* **34**, 1483.
4. Ducrot, P.-H., Lallemand, J.-Y., Milat, M.-L. and Blein, J.-P. (1994) *J. Chem. Soc., Chem. Commun.* 2215.
5. Ducrot, P.-H., Lallemand, J.-Y., Milat, M.-L. and Blein, J.-P. (1994) *Tetrahedron Letters* **35**, 8797.
6. Prangé, T., Neuman, A., Milat, M.-L. and Blein, J.-P. (1995) *Acta Crystallogr. B* **51**, 308.
7. Minami, H., Kinoshita, Y., Fukuyama, Y., Kodama, M., Yoshisawa, T., Sugaira, M., Nakagawa, K. and Tago, H. (1994) *Phytochemistry* **36**, 501.
8. Torel, J., Cillard, J. and Cillard, P. (1986) *Phytochemistry* **25**, 383.
9. Hidalgo, M. E., Fernandez, E., Quilhot, W. and Lissi, E. (1994) *Phytochemistry* **37**, 1585.
10. Misra, H. P. and Squatrito, P. M. (1982) *Arch. Biochem. Biophys.* **215**, 59.
11. Thorpe, G. H. G. and Kricka, L. J. (1986) in *Methods in Enzymology* (Deluca, M. A. and McElroy, W. D., eds), Vol. 133, p. 331. Academic Press, New York.
12. Maehly, A. C. and Chance, B. (1954) in *Methods of Biochemical Analysis* (Glick, D., ed.), Vol. 1, p. 357. Interscience, New York and London.
13. Range, P., Badeck, F. W., Plöchl, M. and Kohlmaier, G. H. (1993) *New Phytol.* **125**, 771.
14. Orth, A. B., Sfarra, A., Pell, E. J. and Tien, M. (1993) *Pestic. Biochem. Physiol.* **47**, 134.
15. Miller, R. W. and MacDowall, F. D. H. (1975) *Biochim. Biophys. Acta* **38**, 176.
16. Qiu, Q.-S., Liang, H.-G., Zheng, H.-J. and Chen, P. (1994) *Plant Sci.* **101**, 99.
17. Sanchez, L. M., Doke, N. and Kawakita, K. (1993) *Plant Sci.* **88**, 141.
18. Doke, N., Miura, Y., Sanchez, L. M. and Kawakita, K. (1994) in *Causes of Photooxidative Stress and Amelioration of Defense Systems in Plants* (Foyer, C. H. and Mullineaux, P. M., eds), p. 177. CRC Press, Boca Raton, FL.
19. Milat, M.-L. and Blein, J.-P. (1995) *J. Chromatogr. A* **699**, 277.