

27-NOR-TRITERPENOID GLYCOSIDES FROM *ADINA RUBELLA*

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**Key Word Index**—*Adina rubella*; Rubiaceae; saponins; 3 $\beta$ -hydroxy-27-nor-olean-13-en-28-oic acid glycosides; pyrocincholic acid; rubelloside C and D.

**Abstract**—Two new 27-nor-triterpenoid glycosides, pyrocincholic acid-3 $\beta$ -O- $\beta$ -D-glucopyranosyl-(1 $\rightarrow$ 2)- $\beta$ -D-fucopyranoside, pyrocincholic acid-3 $\beta$ -O- $\alpha$ -L-rhamnopyranosyl-28-[ $\beta$ -D-glucopyranosyl-(1 $\rightarrow$ 6)- $\beta$ -D-glucopyranosyl] ester, named rubelloside C and D, respectively, were isolated from roots of *Adina rubella*. Their structures were elucidated on the basis of spectral data.

## INTRODUCTION

*Adina rubella* Hance, a Chinese folk medicine, has been reported to contain rubelloside A and B [1]. These compounds were isolated from the ether fraction of the ethanol extract of its roots. The ethyl acetate fraction has been shown to contain two new nor-triterpenoid glycosides, pyrocincholic acid-3 $\beta$ -O- $\beta$ -D-glucopyranosyl-(1 $\rightarrow$ 2)- $\beta$ -D-fucopyranoside (**1**), pyrocincholic acid-3 $\beta$ -O- $\alpha$ -L-rhamnopyranosyl-28-[ $\beta$ -D-glucopyranosyl-(1 $\rightarrow$ 6)- $\beta$ -D-glucopyranosyl] ester (**2**), named rubelloside C and D, respectively.

## RESULTS AND DISCUSSION

The nor-triterpenoid glycoside **1** was obtained as a powder. The molecular formula was determined as C<sub>41</sub>H<sub>66</sub>O<sub>12</sub> by <sup>13</sup>C NMR DEPT (Table 1) and FAB-mass spectral data (*m/z* 773 [M + Na]<sup>+</sup>). The presence of a nor-triterpenoid glycoside containing two sugar subunits was revealed by its <sup>1</sup>H (Table 2) and <sup>13</sup>C NMR spectra and FAB-mass spectral data. In the <sup>1</sup>H NMR spectrum, there are characteristic signals of nor-triterpenoid, but no olefinic proton resonances. The <sup>13</sup>C NMR spectrum of the aglycone showed 29 carbon signals including two quaternary olefinic carbons ( $\delta$  137.1, 130.7). A comparison of the <sup>13</sup>C NMR spectra of **1** and pyrocincholic acid-3 $\beta$ -O- $\beta$ -D-deoxyglucopyranosyl-28-[ $\beta$ -D-glucopyranosyl] ester (**3**) [2] revealed that the carbon signals of the aglycones were almost identical except for the chemical shift of the carboxylic carbon of **1**, (180.0 ppm rather than 176.5 ppm) showing that it was not glycosidated. These data indicated that the aglycone of compound **1** was

pyrocincholic acid (**4**) and the glycosidation site was C-3. Acid hydrolysis gave D-glucose and D-fucose. The <sup>1</sup>H and <sup>13</sup>C NMR spectra were assigned by <sup>1</sup>H-<sup>1</sup>H COSY, TOCSY, NOESY, HMQC and HMBC experiments. The coupling constants of the <sup>1</sup>H NMR signals of the sugar moieties suggested a  $\beta$ -D-glucopyranose and  $\beta$ -D-fucopyranose (Table 2). The <sup>13</sup>C NMR signals of the glucose and methyl- $\beta$ -D-glucopyranoside ( $\delta$  106.1, 76.6, 78.2, 71.6, 77.9, 62.5) [3] were almost identical and those of the fucose of **1** and  $\beta$ -D-fucopyranose ( $\delta$  106.3, 73.0, 75.5, 72.8, 71.3, 17.4) [4] were similar, except that C-1 was shifted upfield by 1.1 ppm and C-2 downfield by 8.9 ppm indicating that the fucose was attached to C-3 and the glucose was attached to C-2'. In the NOESY spectrum there were cross-peaks between H-1' and H-3, H-1'' and H-2', confirming the linkage mode. From these considerations, the structure of **1** was deduced to be pyrocincholic acid-3 $\beta$ -O- $\beta$ -D-glucopyranosyl-(1 $\rightarrow$ 2)- $\beta$ -D-fucopyranoside.

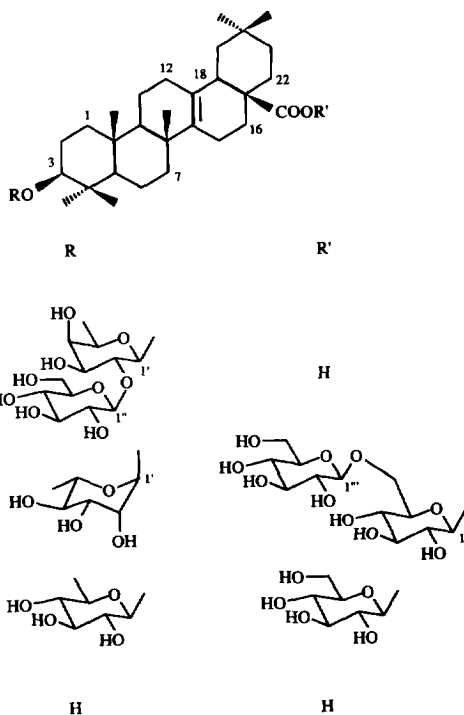
Compound **2** was obtained as a powder whose molecular formula, C<sub>47</sub>H<sub>76</sub>O<sub>17</sub>, was determined by <sup>13</sup>C NMR DEPT and FAB mass spectral data (*m/z* 935 [M + Na]<sup>+</sup>). The <sup>13</sup>C NMR signals of the aglycone were very similar to those of **3**, indicating that the aglycone and the glycosidation sites were identical. Acid hydrolysis gave L-rhamnose and D-glucose. In the <sup>13</sup>C NMR spectrum, the carbon signals of the rhamnose were consistent with those of methyl rhamnopyranoside ( $\delta$  102.1, 71.2, 71.5, 73.3, 69.5, 17.9) [3]. There were two glucose units. A comparison of their carbon signals with those of methyl- $\beta$ -D-glucopyranoside ( $\delta$  106.1, 76.6, 78.2, 71.6, 77.9, 62.5) [3] revealed that one of them ( $\delta$  95.7, 74.2, 78.8 or 78.4, 69.8, 78.0, 69.8) was attached to the aglycone at C-28 while the other glucose ( $\delta$  105.3, 75.2, 78.4 or 78.8, 71.0, 78.4 or 78.8, 62.9) or the rhamnose was attached to it at C-6''. The

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Table 1.  $^{13}\text{C}$  NMR spectral data for compounds **1** and **2**

C	1	2
1	38.7	38.5
2	27.0	26.1
3	89.2	88.9
4	40.0	39.6
5	56.1	55.6
6	18.9	19.0
7	39.8	39.4
8	38.1	38.2
9	56.7	56.9
10	37.4	37.4
11	18.3	18.1
12	32.3	32.1
13	130.7	130.4
14	137.1	137.0
15	21.3	21.0
16	24.4	24.3
17	45.3	45.8
18	39.8	39.6
19	41.8	41.6
20	30.9	30.7
21	34.8	34.5
22	31.8	31.4
23	28.3	28.3
24	16.7	16.7
25	16.7	16.7
26	20.3	21.0
28	180.0	176.7
29	32.6	32.4
30	25.3	25.1
1'	105.2	104.4
2'	81.9	72.5
3'	75.4	73.0
4'	72.5	74.2
5'	71.0	69.8
6'	17.4	18.5
1''	106.1	95.7
2''	76.8	74.2
3''	77.7	78.8*
4''	71.9	69.8
5''	78.1	78.0
6''	62.9	69.8
1'''		105.3
2'''		75.2
3'''		78.4*
4'''		71.0
5'''		78.4*
6'''		62.9

\*Assignments may be interchanged within each column.



tached to the inner glucose at C-6''. This was consistent with the data for a reported 28-*O*-gentiobiosyl moiety (inner glucose:  $\delta$  95.7, 74.0, 78.7\*, 71.4, 77.3, 69.7; terminal glucose:  $\delta$  105.1, 75.2, 78.4\*, 71.9, 78.7\*, 62.9\*; assignments may be interchanged) [5]. From these results, the structure of **2** was established as pyrocincholic acid-3-*O*- $\alpha$ -L-rhamnopyranosyl-28- $[\beta$ -D-glucopyranosyl-(1  $\rightarrow$  6)- $\beta$ -D-glucopyranosyl] ester.

#### EXPERIMENTAL

General procedures were carried out as described in our previous paper [1]. The EtOAc fr. (143.5 g) was chromatographed on a silica gel column using  $\text{CHCl}_3$ -MeOH as eluent. The frs eluted with  $\text{CHCl}_3$ -MeOH (1:2) were further chromatographed on a silica gel column eluting with EtOAc-MeOH (7:1), **1** (21 mg) and **2** (18 mg) were obtained consecutively.

**Rubelloside C (1).** Powder, mp 232–235°,  $[\alpha]_{\text{D}}^{20}$  –99.4° (MeOH; *c* 0.053). IR  $\nu_{\text{max}}$   $\text{cm}^{-1}$ : 3400, 2940, 1700, 1650, 1460, 1380, 1360, 1300, 1180, 1070. FAB-MS  $m/z$ : 773  $[\text{M} + \text{Na}]^+$ .  $^{13}\text{C}$  and  $^1\text{H}$  NMR: Tables 1 and 2.

**Rubelloside D (2).** Powder, mp 225–226°.  $[\alpha]_{\text{D}}^{20}$  –47.1° (MeOH; *c* 0.034). IR  $\nu_{\text{max}}$   $\text{cm}^{-1}$ : 3400, 2940, 1730, 1640, 1450, 1380, 1250, 1060. FAB-MS  $m/z$ : 935  $[\text{M} + \text{Na}]^+$ .  $^{13}\text{C}$  and  $^1\text{H}$  NMR: Tables 1 and 2.

**Acid hydrolysis of 1 and 2.** **1** and **2** (10 mg, respectively) were submitted to acid hydrolysis in the usual way. The sugars were identified with authentic samples by PC.

proton signals of the sugar moieties were assigned from the  $^1\text{H}$ - $^1\text{H}$  COSY spectrum. The coupling constants were consistent with the above results. On irradiation of H-3 a NOE was observed at H-1', a *vice versa* effect also indicating that it was the rhamnose that was attached to C-3. Thus, the terminal glucose was at-

Table 2.  $^1\text{H}$  NMR spectral data for compounds **1** and **2**

H	1	2
3	3.31 <i>dd</i> (11.6, 4.2)	3.28 <i>dd</i> (11.5, 4.2)
18	2.85 <i>dd</i> (11.7, 3.9)	2.71 <i>dd</i> (11.6, 3.3)
23	1.30 <i>s</i>	1.25 <i>s</i> *
24	1.10 <i>s</i>	0.98 <i>s</i> *
25	0.77 <i>s</i>	0.94 <i>s</i> *
26	0.95 <i>s</i>	0.91 <i>s</i> *
29	0.99 <i>s</i>	0.82 <i>s</i> *
30	1.00 <i>s</i>	0.78 <i>s</i> *
1'	4.78 <i>d</i> (7.5)	5.33 <i>br s</i>
2'	4.54 <i>dd</i> (9.3, 7.5)	4.54 <i>br s</i>
3'	4.15 <i>dd</i> (9.3, 3.3)	4.48 <i>overlap</i>
4'	4.00 <i>br d</i> (3.3)	4.30 <i>overlap</i>
5'	3.75 <i>overlap</i>	4.30 <i>overlap</i>
6'	1.53 <i>d</i> (6.3)	1.69 <i>d</i> (5.5)
1''	5.21 <i>d</i> (7.6)	6.24 <i>d</i> (8.0)
2''	4.10 <i>dd</i> (9.1, 7.6)	4.11 <i>t</i> (8.0)
3''	4.15 <i>t</i> (9.1)	4.30 <i>overlap</i>
4''	4.30 <i>t</i> (9.1)	4.15 <i>overlap</i>
5''	3.75 <i>overlap</i>	3.90 <i>overlap</i>
6''	4.40 <i>d</i> (3.4), 4.40 <i>d</i> (3.4)	4.48 <i>overlap</i> , 4.30 <i>m</i>
1'''		5.00 <i>d</i> (7.7)
2'''		4.00 <i>t</i> (7.7)
3'''		4.15 <i>overlap</i>
4'''		4.30 <i>overlap</i>
5'''		4.08 <i>overlap</i>
6'''		4.30 <i>overlap</i> , 4.72 <i>d</i> (10.2)

Coupling constants (*J* in Hz) are given in parentheses; the assignments were based on  $^1\text{H}$ -COSY (**1**, **2**), NOESY (**1**), TOCSY (**1**), HMQC (**1**) and HMBC (**1**).

\*Assignments may be interchanged within each column.

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