

PII: S0031-9422(96)00160-4

MAYOSIDE, AN OXANTHRONE FROM PICRAMNIA HIRSUTA

MARIA DEL R. HERNANDEZ-MEDEL,* OSCAR LOPEZ-MARQUEZ, ROSA SANTILLAN† and ANGEL TRIGOS‡

Instituto de Ciencias Básicas, Universidad Veracruzana, Av. Dos Vistas s/n, Carretera Xalapa-Las Trancas, 91000. Xalapa, Ver., México; †Departamento de Química, Centro de Investigación y de Estudios Avanzados, Instituto Politécnico Nacional, A.P. 14-740, México, D.F., 07000. México; ‡Instituto de Investigación y Posgrado, Departamento de Química y Biología, Escuela de Ciencias, Universidad de las Américas Puebla, A.P. 100, 72820. Cholula, Puebla. México

(Received in revised form 30 January 1996)

Key Word Index—Picramnia hirsuta; Simaroubaceae; oxanthrone, mayoside.

Abstract—A new oxanthrone named mayoside was isolated from the roots of *Picramnia hirsuta* and the structure established by X-ray diffraction. Copyright © 1996 Elsevier Science Ltd

INTRODUCTION

Picramnia is the largest and most complex genus in the Simaroubaceae and comprises 40 species in the American tropics. P. hirsuta appears to be restricted to rainforests near the Isthmus of Tehuantepec, Mexico, ranging from Catemaco, Veracruz, to the Uxpanapa region of Oaxaca and Veracruz [1]. Previous investigations of the genus Picramnia have resulted in the isolation of several anthraquinones and triterpenoids [2–5].

RESULTS AND DISCUSSION

The extracts were subjected to repeated column chromatography on silica gel to yield compounds 1-5. Compounds 1-4 were shown to be β -sitosterol [6], chrysophanol, emodin [7] and 7-hydroxycoumarin [8], respectively, by comparison with authentic samples and spectral data [9]. Compound 5 gave rise to a UV spectrum (218, 275, 366 and 373 nm) characteristic of a highly conjugated system, such as an anthraquinone, and the IR spectrum showed absorption bands at 3540, 3390, 1715 and 1635 cm⁻¹. The ¹H and ¹³C NMR spectra established the presence of 24 protons and 27 carbons. The GC-mass spectrum displayed a fragmentation ion at m/z 402 $[M-122]^+$ indicating the loss of benzoic acid. The presence of an emodin system was supported by the ¹H NMR spectrum (Table 1), which showed two singlets for chelated hydroxyl groups (δ 12.17 and 12.14) and the typical AB pattern for the protons in the H-5, H-7 and H-4, H-2 positions $(\delta 7.20, 6.82 \text{ and } 6.73, 6.09, \text{ respectively})$. The singlet at δ 10.44 (1H, D₂O exchangeable proton) was due to a

hydroxyl group in the 3 position. There were signals for nine aromatic protons from δ 6.09-7.80 and the COSY spectrum showed three separate aromatic rings; three D_2O exchangeable protons at δ 6.14, 5.18 and 5.01 were assigned to the hydroxyl groups in the sugar moiety; and a singlet at δ 5.70 was assigned to the proton on C-1'. The two proton singlet-shaped multiplet at δ 3.70 was assigned to the protons on C-2' and C-3' and the two proton singlet-shaped multiplet at δ 3.54 was assigned to the protons at C-4' and C-5'. The ¹³C NMR assignments are presented in Table 1. The spectrum showed 25 signals, including two CH signals at δ 129.01 and δ 129.33 which showed a double intensity characteristic of a monosubstituted benzene ring. The DEPT experiment was accounted for by one methyl, 14 methine, and 12 quaternary carbons. An interesting feature of the ¹³C NMR spectrum was the presence of one quaternary carbon resonance at δ 75.03 for C-10, indicating oxygenation at this position and hence the presence of an oxanthrone moiety. The presence of a C-glycosyde unit was confirmed by five CH signals at δ 94.52, 80.10, 71.67, 69.00 and 68.34. The full assignment of the ¹³C NMR signals was mainly based on comparison with those of anthrones [10-11] and oxanthrones [12-13] and also on HET-COR experiments (Table 1). The above information led to the conclusion that compound 5, named mayoside, has the structure shown in Fig. 1. The lyxose nature of the sugar moiety was established by single-crystal Xray diffraction.

EXPERIMENTAL

General. MP: uncorr.; TLC: silica gel (Merck 60 GF₂₅₄; 0.2 mm thickness); CC: silica gel (Merck, Kieselgel 60 particle size 0.063-0.200 mm and 0.040-0.063 mm); ¹H and ¹³C NMR; Varian Gemini 200

^{*}Author to whom correspondence should be addressed.

Table 1. 1	H NMR and	¹³ C NMR	spectral data for	r mavoside, 5	(DMSO- d_6 , δ , ppm)
------------	-----------	---------------------	-------------------	---------------	--------------------------------

С	δ	Multiplicity* DEPT	¹ H- ¹³ C† Connectivity	Multiplicity‡ ¹H NMR	¹ H- ¹ H† Connectivity
1	163.94	С			
1a	108.97	C			
2	101.63	CH	H-2 (δ 6.09)	d(2.0)	H-4
3	165.37	C			
4	105.94	CH	H-4 (δ 6.73)	d(2.0)	H-2
4a	146.17	C			
5	119.68	CH	H-5 (δ 7.20)	S	H-7 and H-11
5a	144.78	C			
6	150.92	C			
7	117.11	CH	H-7 (δ 6.82)	S	H-5 and H-11
8	161.25	C			
8a	113.54	C			
9	190.30	C			
10	75.03	C			
11	22.35	CH,	H-11 (δ 2.40)	S	H-5 and H-7
1'	94.52	CH	H-1' (δ 5.70)	S	H-2'
2'	69.00	CH	H-2' (δ 3.70)	S	H-1' and 2' OH
3'	71.67	CH	H-3′ (δ 3.70)	S	H-4' and 3' OH
4'	68.34	CH	H-4' (δ 3.54)	S	H-3' and 4' OH
5'	80.10	CH	H-5' (δ 3.54)	S	H-4'
1''(C=0)	163.58	C			
1"	129.18	C			
2" and 6"§	129.33	СН	$H-2''/H-6'' (\delta 7.80)$	d (7.6)	H-3"/H-5"
3" and 5"§	129.01	СН	$H-3''/H-5'' (\delta 7.55)$	dd (7.6 and 7.2)	H-2"/H-6" and H-4"
4"	133.81	CH	$H-4'' (\delta 7.70)$	d (7.2)	H-3"/H-5"

^{*}Multiplicity from DEPT ¹³C NMR experiment.

instrument in $CDCl_3$ and $DMSO-d_6$ with TMS as int. standard.

Plant material. The root of Picramnia hirsuta was collected at Valle de Uxpanapa, Veracruz, México, during November 1992. A voucher specimen is deposited at the Herbarium of the Centro de Investigaciones Biológicas, Universidad Veracruzana (XALU).

Isolation. The chipped roots $(1.60 \,\mathrm{kg})$ were soaked successively in $\mathrm{C_6H_{14}}$, $\mathrm{CHCl_3}$ and MeOH at room temp. and then filtered; the filtrates after removal of the solvents under red. pres. gave residues weighing 1.5 g, 3.2 g and 50.3 g, respectively.

The crude C_6H_{14} extract was chromatographed on silica gel. Elution with C_6H_{14} –CHCl $_3$ (4:1) yielded compound 1 (40 mg). The CHCl $_3$ extract was subjected to CC on silica gel; elution with C_6H_{14} –CHCl $_3$ (4:1) and C_6H_{14} –EtOAc (8:2, 7:3) gave compounds 2 (5 mg), 3 (7 mg) and 4 (12 mg). The crude MeOH extract (30 g) was chromatographed on silica gel, using CHCl $_3$, CHCl $_3$ –MeOH (19:1, 9:1, 17:3, 4:1) as eluents to yield compounds 3 (25 mg), 4 (15 mg) and 5 (50 mg).

Compounds 1–4 were identified as sitosterol, chrysophanol, emodin and 7-hydroxycoumarin, respec-

tively, by spectral data (¹H NMR and ¹³C NMR) and direct comparison with authentic samples [7–9].

Compound 5. Yellow needles (MeOH), mp 238–240°. UV λ_{max} (EtOH) nm: 218, 275, 366 and 373; IR (KBr) ν_{max} , (cm⁻¹): 3540 and 3390 (OH), 2940, 1715, 1635, 1610, 1250, 1155, 910, 850 and 710; GC–MS 70 eV, m/z (rel. int.): 524 (1) [M]⁺, 402 (5) [M – C₇-H₆O₂]⁺ 122 (75) C₇H₆O₂, 77 (100) C₆H₅; ¹H NMR and ¹³C NMR: Table 1.

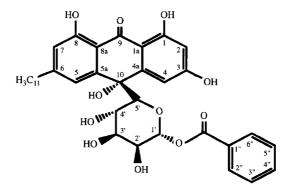
X-ray analysis. Suitable crystals of mayoside (5) were grown from MeOH as prisms. Intensity data were collected at room temp. on a CAD4-Enraf-Nonius diffractometer using MoKα radiation (λ = 0.71069 Å) with the $\omega/2\theta$ data collection method in the range 2–25°. Crystal data for 5: C₂₇H₂₄O₁₁, square prism, space group P1, a = 8.251 (3), b = 8.955 (3), c = 9.931 (2) Å, α = 61.83 (2), β = 70.50 (2), γ = 74.65 (3), °, V = 604.7 (3) ų, Z = 1. 1768 observed reflections [(Fo)² > 3σ(Fo)²], final R = 0.040, Rw = 0.039. The final fractional coordinates, thermal parameters, bond distances and angles have been deposited with the Cambridge Crystallographic Centre.

Acknowledgement—The authors thank Professor Mario Vázquez (Centro de Investigaciones Biológicas, Uni-

[†]Assignments were confirmed by ¹H COSY, DEPT and HETCOR experiments.

[‡]Multiplicity from ¹H NMR experiment. Coupling constants (*J* in Hz) in parentheses.

[§]The intensity for this signal was twice that of the other CH signals.



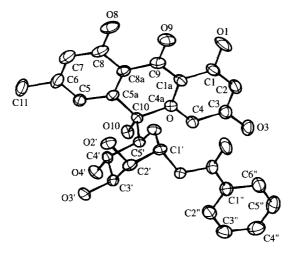


Fig. 1. Formula of mayoside (5), as determined by X-ray analysis.

versidad Veracruzana, México) for the identification of the plant.

REFERENCES

- 1. Thomas, W. W. (1988) Brittonia 40, 89.
- Arana, C. and Julca, B. (1986) Rev. Peru Bioquim. 8, 16.
- Leon, C. and Juan, J. (1975) Bol. Soc. Quim. Peru 41, 14.
- Herz, W., Santhanam, P. S. and Wahlberg, I. (1972) *Phytochemistry* 11, 3061.
- Solis, P. N., Gutierrez-Ravelo, A., Gonzalez A. G., Gupta, M. P. and Phillipson, J. D. (1995) Phytochemistry 38, 447.
- 6. Salama, A. M., Sánchez-López, M., Gutierrez, M. and Achenbach, H. (1987) Rev. Latinoamer. Quim.

- 18 (3), 132.
- Danielsen, K., Aksnes, D. W. and Francis, G W. (1992) Magn. Res. Chem. 30, 359.
- Duddeck, H., Kaiser, M. (1982) Org. Magn. Res. 20 (2), 55.
- 9. Wagner, H., Bladt, S. and Zgainski, E. M. (1984) *Plant Drug Analysis*. Springer-Verlag, New York.
- Manitto, P., Monti, D. and Speranza, G. (1990) J. Chem. Soc. Perkin Trans. 1, 1297.
- Manitto, P., Monti, D., Speranza, G., Mulinacci, N., Vincieri, F. F., Griffini, A. and Pifferi, G. (1993) J. Chem. Soc. Perkin Trans. 1, 1577.
- Adinolfi, M., Corsaro, M. M., Lanzetta, R., Parrilli, M. and Scopa, A. (1989) *Phytochemistry* 28 (1), 284.
- Yenesew, A., Dagne, E., Müller, M. and Steglich, W. (1994) Phytochemistry 37 (2), 525.