

FAREANINE AND FAREANOL FROM LEAVES OF *MEDICOSMA FAREANA*

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Abstract—Two new compounds, named fareanine (1-methyl-3ξ-hydroxy-2,2-dimethoxy-3ξ-methoxycarbonyl-1,4H-quinoline-[2,3: b]-cyclopentan-4,5-dione) and fareanol (1ξ,2-dihydroxy-1ξ-(4-hydroxy-3,5-dimethoxyphenyl)-ethane, together with the known compounds 1,3,4-trimethoxy-10-methyl acridone, normelicopicine, melicopidine, and *p*-hydroxybenzaldehyde, have been isolated from leaves of *Medicosma fareana*. The identity of the new compounds was established from their spectroscopic data. Fareanine appears to be the product of fission of the C-ring of a normal acridone precursor. Copyright © 1996 Elsevier Science Ltd

INTRODUCTION

Medicosma fareana is a small tree occurring in the rain forest areas of North-Eastern Queensland, Australia, between sea-level and *ca* 800 m [1]. Previous phytochemical work on this species resulted in the isolation of the furoquinoline alkaloids, skimmianine and acronycidine, the acridones, melicopine, melicopidine and melicopicine, and the common triterpene, lupeol [2]. In the present study, we report the isolation of four known compounds (*p*-hydroxybenzaldehyde and 1–3) and the structural elucidation of two new compounds, fareanine (4) and fareanol (7), from the leaves of *M. fareana*.

RESULTS AND DISCUSSION

The ethanol extract of leaves of *M. fareana* was suspended in water and successively re-extracted with hexane, chloroform, ethylacetate and butanol. Vacuum liquid chromatography (VLC) followed by column and preparative TLC of the chloroform fraction gave the known compounds 1,3,4-trimethoxy-10-methylacridone (1) [3], normelicopicine (2) [4], melicopidine (3) [2, 5, 6], *p*-hydroxybenzaldehyde and two new compounds, fareanine (4) and fareanol (7).

The high-resolution EI and FAB mass spectra of compound 4 analysed for $[M]^+$ 347 ($C_{17}N_1O_7$). The IR spectrum showed a broad band between 3500 and 3000 cm^{-1} (OH), and bands in the carbonyl region at 1713 (ester) and 1620 and 1618 cm^{-1} , characteristic of α,β -unsaturated carbonyls. The 1H and ^{13}C NMR spectra revealed typical signals for the *N*-methyl-4-

quinolone part of the acridone skeleton unsubstituted in the aromatic ring [3–6] (Table 1). The structural assignments and the chemical shift values of the A and B rings were further substantiated by HMBC studies (Table 2).

The C ring of alkaloid 4, which differs from that of all known acridones, was established from HMBC and NOESY studies. In the HMBC spectrum (Table 2), a 3J interaction between the protons of two methoxyl substituents (δ 3.53 and 3.59) and a carbon signal at δ 99.6 required the methoxyls to be placed geminal on an *sp*³

Table 1. 1H and ^{13}C data for compound 4 (400 MHz, $CDCl_3$)

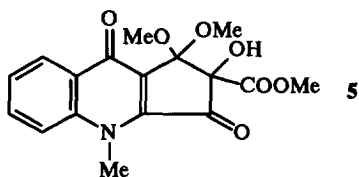
H/C	δ_H	σ_C
1		186.9
2		81.8
3		168.7
4		99.6
5	7.59 <i>d</i> (8.4)	116.3
6	7.77 <i>td</i> (7.1, 1.6)	133.8
7	7.51 <i>td</i> (8.0, 0.9)	126.1
8	8.52 <i>dd</i> (8.0, 1.4)	128.1
9		173.5
10		
11		164.4
12		116.3
13		129.2
14		141.7
<i>N</i> -Me	3.88 <i>s</i>	36.0
OMe-4 _A	3.53 <i>s</i>	52.5
OMe-4 _B	3.59 <i>s</i>	53.3
Ester-Me	3.80 <i>s</i>	54.1
OH-2	4.53 <i>br s</i>	

Coupling constants (Hz) in parentheses.

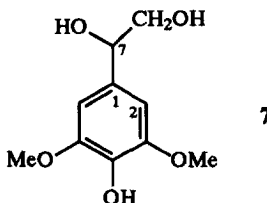
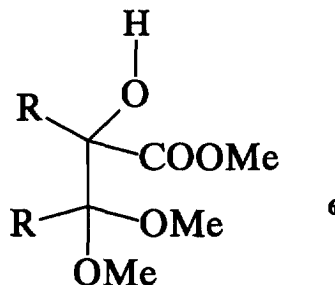
*Author to whom correspondence should be addressed.

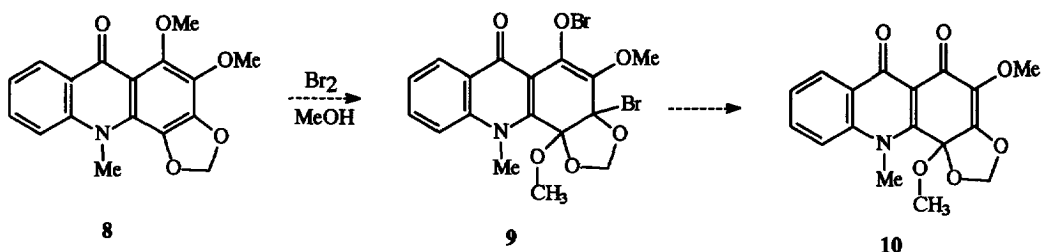
H	1J	2J	3J
5		133.8	126.1, 129.2
6		116.3	128.1, 141.7
7		133.8	116.3, 129.2
8	128.1		133.8, 141.7, 173.5
<i>N</i> -Me	36.0		141.1, 164.4
OMe-4 _A	52.5		99.6
OMe-4 _B	53.3		99.6
Ester-Me	54.1		168.7
OH-2		81.8	99.6, 168.7

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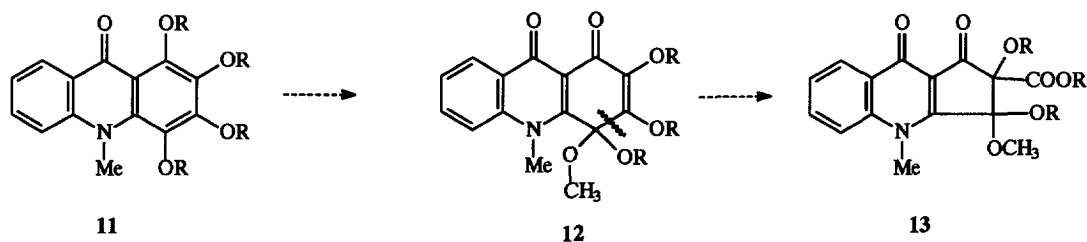


Alkaloid **4** (trivial name fareanine) is likely to be a degraded product of the highly oxygenated C-ring of acridone alkaloids of the type isolated in the present and in previous studies on this species. A possible route to the formation of compound **4** is shown in Scheme 1b and receives support from synthetic studies. In their attempt to find a convenient method of degrading the oxygenated ring of acridones, Prager and Thredgold [7] brominated moelicopine (**8**) and this yielded compound **10**, presumably through the unstable intermediate **9** (Scheme 1a). Their oxidation–reduction reactions did not lead to compound **4**, but similar degradation pathways are likely to be involved in the formation of **4**, (cf. Scheme 1b). Following the formation of an





Scheme 1a



Scheme 1b

intermediate, such as **12**, the C₃–C₄ bond of the C-ring must be broken with oxidation of C-3 and recyclization through C₂–C₄ leading to compounds such as **13** or **4**.

The high-resolution EI-mass spectrum of the second new compound **7** revealed an empirical formula of C₁₀H₁₄O₅. The ¹H and ¹³C NMR spectra (Table 3) revealed signals for one primary and one secondary alcohol functional group. The broad hydroxyl doublet signal in the ¹H NMR spectrum showed coupling to the oxymethine which, in turn, showed coupling to the two oxymethylene protons (¹H–¹H COSY). The ¹H NMR further revealed signals for two, chemically equivalent, aromatic protons, one olefinic hydroxyl and two identical methoxys. Because the ¹³C NMR also showed signals in the aromatic region for only four carbons signals (one methine), compound **7** must possess a symmetrical aromatic skeleton. The two methoxys

must then be assigned at C-3 and C-5 and the hydroxyl at C-4 position.

Both compounds **4** and **7** were isolated as a result of bioassay-guided separation. Their biological activity will be discussed elsewhere.

EXPERIMENTAL

General. Silica gel (Merck 7749) was used for VLC and silica gel 60 PF₂₅₄ for prep. TLC. Spots and bands were detected by spraying with 5% vanillin in concd H₂SO₄. UV: EtOH and IR: KBr. ¹H, ¹³C, ¹H–¹H COSY, NOESY (mixing time *d*₈ = 0.8 s) and HMBC (*d*₆ set for *ca* *J* = 7 Hz) NMR spectra were recorded on a Bruker AMX-400 instrument. Chemical shifts are reported in ppm relative to CDCl₃. EI-MS were obtained by direct probe insertion at 70 eV.

Plant material. Leaves of *M. fareana* (F. Muell) T. G. Hartley were collected from Northern Queensland in 1991. A voucher specimen TGH-15147, was identified by Dr T. G. Hartley and deposited at the Australian National Herbarium, Canberra.

Extraction and isolation. Ground leaves (590 g) were Soxhlet-extracted to exhaustion. Removal of solvent yielded a residue (90 g) which was suspended in H₂O and subjected to successive partitioning against hexane, CHCl₃, EtOAc and MeOH. The CHCl₃ fr. (14 g) was further subjected to VLC over silica gel, eluting with hexane and hexane–CHCl₃ mixts of increasing polarity and finally CHCl₃ containing increasing amounts of MeOH. The hexane–CHCl₃ (1:9) eluent was collected and chlorophyll removed by elution through Sephadex LH 20 (CHCl₃–MeOH, 1:1).

Table 3. ¹H and ¹³C chemical shift data for compound **7** (400 MHz, CDCl₃)

H/C	¹ H	¹³ C
1		132.0
2	6.59 <i>s</i>	103.0
3		147.4
4		137.6
5		147.4
6	6.59 <i>s</i>	103.0
7	3.10 <i>m</i>	86.3
8	3.90, 4.30 <i>m</i>	72.0
OMe-3, OMe-5	3.90 <i>s</i>	56.5
OH-4	5.50 <i>br s</i>	
OH-7	5.50 <i>br d</i> (4.4)	

Coupling constant (Hz) in parentheses.

The resulting gum was subjected to repetitive prep. TLC (silica gel, CHCl_3 -MeOH, 24:1) to give 1,3,4-trimethoxy-10-methyl acridone (**1**, 5 mg, 0.0009%), normelicopicine (**2**, 6 mg, 0.001%) and melicopidone (**3**, 50 mg, 0.009%). Similar treatment of the CHCl_3 VLC eluent gave *p*-hydroxybenzaldehyde (4 mg, 0.0007%), fareanine (**4**, 8 mg, 0.0013%) and fareanol (**7**, 6 mg, 0.001%).

1,3,4-Trimethoxy-10-methyl acridone (**1**), normelicopicine (**2**), melicopidine (**3**) *p*-hydroxybenzaldehyde. All gave UV, IR, EI-MS and ^1H NMR data in close agreement with those reported [3-6].

Fareanine (1-methyl-3 ξ -hydroxy-2,2-dimethoxy-3 ξ -methoxycarbonyl-1,4*H*-quinoline-[2,3:*b*]-cyclopentan-4,5-dione, **4**). Gum. $[\alpha]_D^{20}$ (CHCl_3 , *c* 0.7). UV λ_{max} , nm: 305. IR ν_{max} cm^{-1} : 3450 *br*, 3020, 1713, 1620, 1618, 1602, 1538, 1506, 1463, 1421, 1074 and 1046. ^1H and ^{13}C NMR (400 MHz, CDCl_3 : see Table 1). (HREI-MS: Found 347.1229. calcd for $\text{C}_{17}\text{H}_{17}\text{NO}_7$, 347.1005). EI-MS *m/z* (rel. int.): 347 $[\text{M}]^+$ (30.4), 332 (23.8), 287 (26.5), 272 (100) 244 (60), 77 (67.5).

Fareanol (1 ξ ,2-dihydroxy-1 ξ -(4-hydroxy-3,5-dimethoxyphenyl, **7**). Gum. $[\alpha]_D^{18}$ (CHCl_3 , *c* 0.4). UV λ_{max} , nm: 265, 340 *sh*. IR ν_{max} cm^{-1} : 3398 *br*,

2929, 1616, 1521, 1456, 1521, 1215, 1115. ^1H and ^{13}C NMR (400 MHz, CDCl_3 : see Table 3). (HREI-MS: Found 214.0312. calcd 214.0841 for $\text{C}_{10}\text{H}_{14}\text{O}_5$). EI-MS *m/z* (rel. int.): 214 $[\text{M}]^+$ (10.9), 213 (14.2), 201 (22.7), 199 (15.1), 193 (99.8), 93 (100).

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REFERENCES

1. Hartley, T. G. (1985) *Aust. J. Bot.* **33**, 27.
2. Price, J. R. (1949) *Aust. J. Sci. Res.* **2a**, 249.
3. Funayama, S. and Cordell, G. A. (1984) *J. Nat. Prod.* **47**, 285.
4. Couge, B., Tillequin, F., Koch, M. and Sevenet, T. (1980) *Plant Med. Phytother.* **14**, 208.
5. Bert, M. and Michel-Plat, M. K. (1974) *Phytochemistry* **13**, 301.
6. Bergenthal, D., Mester, I., Rozsa, Z. and Reisch, J. (1979) *Phytochemistry* **18**, 161.
7. Prager, R. H. and Thredgold, H. M. (1967) *Aust. J. Chem.* **21**, 229.