

TEIXIDOL, AN ABEO-TAXANE FROM EUROPEAN YEW, *TAXUS BACCATA*\*

JAVIER SOTO, MONICA FUENTES and LUIS CASTEDO†

Departamento de Química Orgánica, Facultad de Química, Universidad de Santiago de Compostela y Sección de Alcaloides del C.S.I.C., 15706 Santiago de Compostela, Spain

(Received in revised form 5 February 1996)

**Key Word Index**—*Taxus baccata*; Taxaceae; diterpenoids; 11(15→1)abeo-taxanes.

**Abstract**—A new taxoid, teixidol, was isolated from the leaves of *Taxus baccata*. Its structure, which features an 11(15→1)abeo-taxane skeleton consisting of a 5/7/6 membered ring system, was established by spectroscopic analysis. Copyright © 1996 Elsevier Science Ltd

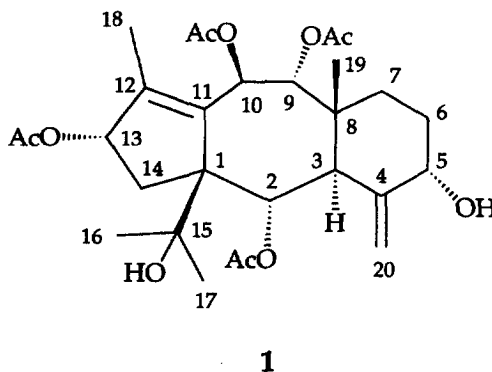
## INTRODUCTION

In our search for novel, potent and more selective antitumour agents, we have recently investigated the taxoid contents of the leaves of a Galician specimen of *Taxus baccata*. In addition to the known taxoids 10-deacetylbaccatine III [1], baccatine III [2], taxuspine F [3], decinnamoyl-1-hydroxytaxinine J [4], 5-cinnamoyl-10-acetyltaxine I [5] and deaminoacyltaxine A [6], we also isolated a new 11(15→1)abeo-taxane [7], teixidol (1). This new taxoid is the 10,13-diacetylated derivative of a taxoid originally assigned a baccatine structure [8] and more recently shown to have a 11(15→1)abeo-taxane skeleton [9].

## RESULTS AND DISCUSSION

A methanolic extract of *T. baccata* leaves collected at Santiago de Compostela provided, in addition to several known taxoids, a new member of the 11(15→1)abeo-taxane family that we have named teixidol (1).

Compound 1 was obtained as an amorphous white solid of mp 159°. A molecular formula of  $C_{28}H_{40}O_{10}$  was established on the basis of its  $^{13}C$  NMR spectrum and an EI mass spectral peak for  $[M - 2xAc]^+$  at  $m/z$  418. IR bands at 3600–3400 and  $1730\text{ cm}^{-1}$  revealed the presence of hydroxyl and ester groups.  $^1H$  and  $^{13}C$  NMR signals were assigned by DEPT, HMQC, COSY and NOE techniques, and are indicated in Table 1. The  $^{13}C$  NMR and DEPT spectra showed signals due to six oxygenated, four secondary (one  $sp^2$  and three  $sp^3$ ), eight primary, one tertiary and five quaternary (three  $sp^2$  and two  $sp^3$ ) carbons, and to four ester carbonyl



groups. The presence of three quaternary signals at  $\delta$  68.7, 42.4 and 75.4 is characteristic of an 11(15→1)abeo-taxane skeleton [7, 9]. The  $^1H$  NMR spectrum showed singlets for four methyl groups at  $\delta$  0.93, 1.15, 1.17 and 1.84, and singlets for four acetyl methyls at  $\delta$  1.97, 1.98, 2.0 and 2.05. The protonated carbons were identified by HMQC and COSY experiments. The relative stereochemistry was elucidated by means of a NOESY experiment [10] (Fig. 1).

The structures of the other six taxoids isolated were determined by comparing their physical and spectral data with those reported in the literature [1–6].

## EXPERIMENTAL

**General.** Mps: uncorr.  $^1H$  and  $^{13}C$  NMR spectra were recorded in  $CDCl_3$  and  $CD_3OD$  on Bruker AMX-300 and AMX-500 spectrometers with TMS as int. standard; the NOE data and 2D  $^1H$ – $^1H$  and  $^{13}C$ – $^1H$  correlation spectra were obtained using standard Bruker software. EIMS: direct inlet, 70 ev. UV: Hewlett Packard 8452A diode array spectrophotometer. IR: Midac Prospect spectrophotometer. HPLC: Waters 600 E system controller and Waters 490 UV detector.

\*Dedicated to Prof. Antonio González González in celebration of his half-century of contribution to natural product chemistry.

†Author to whom correspondence should be addressed.

Table 1.  $^1\text{H}$  and  $^{13}\text{C}$  NMR data of teixidol (1) (300 MHz,  $\text{CDCl}_3$ ,  $\delta$  values with TMS as internal standard,  $J$  in Hz)

No.	$^{13}\text{C}$	$^1\text{H}$
1	68.74 s	
2	68.58 d	5.90 d (9.2)
3	43.65 d	3.42 d (9.2)
4	146.55 s	
5	75.50 d	4.23 s br
6	37.51 t	1.6 m br
7	30.38 d	1.73 m br
8	42.40 s	
9	76.89 d	5.73 d (10)
10	69.10 d	6.15 d (10)
11	146.83 s	
12	136.02 s	
13	79.23 d	5.5 t (7.15)
14	36.81 t	2.35 dd (7.5; 15) 1.9 dd (7.4; 15)
15	75.37 s	
16	25.37 q	1.17 s
17	27.65 q	1.15 s
18	11.09 q	1.84 s
19	17.05 q	0.93 s
20	111.30 t	5.08 s 4.44 s
Ac	171.11 s	
	170.93 s	
	169.88 s	
	168.55 s	
Ac	21.73 t	2.05 s
	21.11 t	2.0 s
	20.79 t	1.98 s
	20.65 t	1.97 s

**Plant material.** *Taxus baccatas* leaves were collected at Santiago de Compostela in March 1992.

**Extraction and separation.** Leaves (5 kg) were extracted with MeOH (16 L) at room temp. The methanolic extract was partitioned between  $\text{CH}_2\text{Cl}_2$  and  $\text{H}_2\text{O}$ , and the solvent was evapd from the organic fr. under red. pres. to give a residue (164 g) that was suspended in MeOH– $\text{H}_2\text{O}$  and loaded on to a reversed-phase chromatography column (Merck Lichroprep RP-18, 25–40  $\mu\text{m}$ ) that was then successively eluted with MeOH– $\text{H}_2\text{O}$  (1:1), MeOH and  $\text{CH}_2\text{Cl}_2$  to give frs I, II and III, respectively.

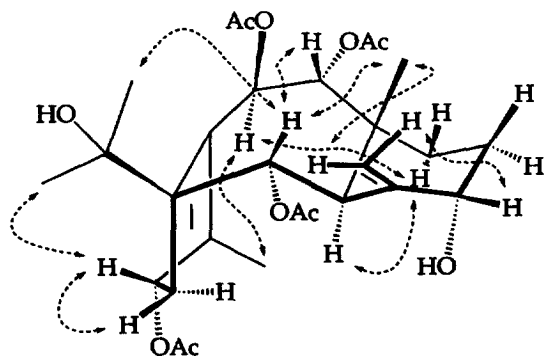


Fig. 1. NOE experiments on teixidol (1).

Fr. I (60 g) was subjected to flash CC on an  $8 \times 25$  cm column of Merck 230–400 silica gel with a  $\text{CH}_2\text{Cl}_2$ –MeOH (1:0–1:1) gradient as eluent; 9 frs were collected. Fr. I-7 (8.6 g) was diluted with  $\text{CH}_3\text{CN}$  and left for 72 hr. The yellow amorphous ppt (500 mg) was filtered out and crystallized from MeCN–MeOH to give 356 mg 10-deacetylbaecatine III. Fr. I-2 (6.3 g) was sepd by reversed-phase HPLC (Zorbax ODS,  $250 \times 20$  mm; flow rate  $6.0 \text{ ml min}^{-1}$ ; UV detection at 230 nm; 7:3–0:100  $\text{H}_2\text{O}$ – $\text{CH}_3\text{CN}$  gradient) to give frs I-2a (706 mg,  $R_f$  23 min) and I-2b (762 mg,  $R_f$  32 min). preparative TLC of I-2a (Merck GF-254 silica gel;  $\text{CH}_2\text{Cl}_2$ –MeOH, 19:1) gave 280 mg deaminoacetylta-xine A. Prep. TLC of I-2b (Merck GF-254 silica gel;  $\text{CH}_2\text{Cl}_2$ – $i$ -PrOH, 49:1) gave 15 mg baecatine III, 20 mg taxuspine F, 31 mg decinnamoyl-1-hydroxy-taxinine J and 22 mg 1. Reversed-phase HPLC of fr. I-9 (5.8 g) (Zorbax ODS  $250 \times 20$  mm; flow rate  $6.0 \text{ ml min}^{-1}$ ; UV detection at 230 nm; 1:1–0:100  $\text{H}_2\text{O}$ –MeCN gradient) gave fr. I-9a (153 mg,  $R_f$  28 min.), which by prep. TLC (Merck GF-254 silica gel;  $\text{CH}_2\text{Cl}_2$ – $n$ -BuOH, 19:1) afforded 5-cinnamoyl-10-acetylta-xine I (75 mg).

**Teixidol (1).** Amorphous solid; mp  $159^\circ$  [ $\alpha_D^{20}$   $-15.91$  ( $c$  0.77,  $\text{CHCl}_3$ )]. Positive EIMS  $m/z$ : 418 [ $\text{M} - 2\text{xAcO}$ ] $^+$ . IR  $\nu_{\text{max}}^{\text{CHCl}_3}$   $\text{cm}^{-1}$ : 3400, 3600, 2931, 1734. UV  $\lambda_{\text{max}}^{\text{CHCl}_3}$  nm: 274.  $^1\text{H}$ – $^1\text{H}$  COSY correlations ( $\text{CDCl}_3$ , H/H): 2/3, 9/10, 13/14, 13/14', 14/14'. NOESY correlations ( $\text{CDCl}_3$ , H/H): 2/9, 2/16, 2/17, 2/19, 3/7, 3/14', 5/6, 5/20, 7/10, 9/19, 10/18, 13/14, 13/16, 13/17, 13/18, 14/14', 19/20', 20/20'.

## REFERENCES

1. Chauvière, G., Guénard, D., Picot, F., Sénilh, V. and Potier, P. (1981) *C. R. Acad. Sci. Paris, Ser. II* **293**, 501.
2. Sénilh, V., Blechert, S., Colin, M., Guénard, D., Picot, P. and Varenne, P. (1984) *J. Nat. Prod.* **47**, 131.
3. Kobayashi, J., Inubushi, A., Hosoyama, H., Yoshida, N., Sasaki, T. and Shigemori, H. (1995) *Tetrahedron* **51**, 5971.
4. Barboni, L., Gariboldi, P., Appendino, G., Enriu, R., Gabetta, B. and Bombardelli, E. (1995) *Liebigs Ann. Chem.* 345.
5. Appendino, G., Gariboldi, P., Pisetta, A., Bombardelli, E. and Gabetta, B. (1992) *Phytochemistry* **41**, 4253.
6. Appendino, G., Cravotto, G., Enriu, R., Jakupovic, J., Gariboldi, P., Gabetta, B. and Bombardelli, E. (1994) *Phytochemistry* **36**, 407.
7. Chen, R. and Kingston, D. G. I. (1994) *J. Nat. Prod.* **57**, 1017.
8. Appendino, G., Özen, H. Ç., Gariboldi, P., Gabetta, B. and Bombardelli, E. (1993) *Fitoterapia* **64**, 47.
9. Appendino, G., Barboni, L., Gariboldi, P., Bombardelli, E., Gabetta, B. and Vitevo, D. (1993) *J. Chem. Soc., Chem. Commun.* 1587.
10. Woods, M. C., Chiang, H.-C., Nakadaira, Y. and Nakanishi, K. (1968) *J. Am. Chem. Soc.* **90**, 522.