



1 β ,15 α -DIHYDROXYFRIEDELAN-3-ONE, A TRITERPENE FROM *SALACIA BEDDOMEI*

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Key Word Index—*Salacia beddomei*; Celastraceae; triterpenes; 1 β ,15 α -dihydroxyfriedelan-3-one.

Abstract—A new triterpene, 1 β ,15 α -dihydroxy-friedelan-3-one, has been isolated from the stem bark of *Salacia beddomei*. Its structure has been elucidated on the basis of NMR and MS techniques. Copyright © 1996 Elsevier Science Ltd

INTRODUCTION

Our previous reports on the constituents of the stem bark of *Salacia beddomei* (Gamble) deal with the isolation and characterization of several new lupane triterpenoids together with three known friedelanes, friedelan-3-one, 15 α -hydroxyfriedelan-3-one (**2**) and 15 α -hydroxyfriedelane-1,3-dione (**3**) [1, 2]. Purification of the fractions obtained during the above chromatographic fractionation yielded a new friedelane compound which has now been characterized as 1 β ,15 α -dihydroxyfriedelan-3-one (**1**) on the basis of spectral evidences.

RESULTS AND DISCUSSION

Compound **1** gave a positive Libermann–Burchard reaction for triterpenes and on TLC gave an orange red spot when sprayed with 2,4-dinitrophenylhydrazine followed by heating, indicating a reactive oxo group in the molecule. Its ^1H and ^{13}C NMR spectra showed characteristic signals of a friedelane compound similar to **2** and **3**, thereby suggesting a friedelane skeleton for **1**.

The EI-mass spectrum of **1** shows the molecular ion at m/z 458 corresponding to the formula $\text{C}_{30}\text{H}_{50}\text{O}_3$, in agreement with other spectroscopic data. The ^1H NMR spectrum shows seven tertiary methyl singlets and a secondary methyl doublet. The presence of two secondary hydroxyl groups in **1** is evident in the ^1H NMR spectrum from the presence of two carbinol methine protons at δ 3.70 (d , $J = 7.9$ Hz) and 4.85 ($br s$) which is supported by the signals of two hydroxylated carbons at δ 71.3 and 74.7 in the ^{13}C NMR spectrum. The additional features of the ^1H NMR spectrum are the presence of a pair of one proton double doublets at

δ 2.69 ($J = 4.6, 13.9$ Hz) and 2.40 ($J = 2.4, 13.9$ Hz), a one proton double doublet at δ 2.16 ($J = 7.9, 15.8$ Hz) and a one proton quartet at δ 2.32 ($J = 7.2$ Hz). The location of the functional groups in **1** were deduced from an analysis of the chemical shift data, decoupling studies and comparison with **2** and **3**.

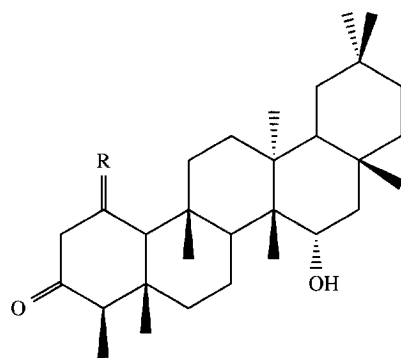
The carbinol methine proton shift at δ 3.70 and the hydroxylated carbon chemical shift at δ 74.8 have been directly assigned for a 15 α -hydroxyl substitution in **1** by analogy to the identical shifts observed for **2** and **3** [1, 3]. Moreover, the ^{13}C NMR shifts observed for the rings C, D and E are in good agreement with the corresponding shifts in **2** and **3** thus confirming a 15 α -hydroxyl substitution.

A C-3 keto substitution in **1** is evident from the one-proton quartet at δ 2.32 due to H-4 α and irradiation of this signal collapses the C-23 methyl doublet at δ 0.95 into a singlet as expected. The double doublets

Table 1. ^1H NMR data of compounds **1** and **2** (CDCl_3 , TMS, 270 MHz)

H	1	2
1	4.85 (<i>brs</i>)	1.94 (<i>m</i>), 1.68 (<i>m</i>)
2	2.69 (<i>dd</i> , $J = 4.6, 13.9$ Hz) 2.40 (<i>dd</i> , $J = 2.4, 13.9$ Hz)	2.40 (<i>m</i>) 2.20–2.28 (<i>m</i>)
4	2.32 (<i>q</i> , $J = 7.2$ Hz)	2.20–2.28 (<i>m</i>)
15	3.70 (<i>d</i> , $J = 7.9$ Hz)	3.73 (<i>d</i> , $J = 7.9$ Hz)
16	2.16 (<i>dd</i> , $J = 7.9, 15.8$ Hz) 1.25 (<i>m</i>)	2.15 (<i>dd</i> , $J = 7.7, 16.1$ Hz) 1.25 (<i>m</i>)
23	0.95 (<i>d</i> , $J = 7.2$ Hz)	0.87 (<i>d</i> , $J = 6.6$ Hz)
24	1.10 (<i>s</i>)	0.73 (<i>s</i>)
25	1.33 (<i>s</i>)	0.89 (<i>s</i>)
26	1.08 (<i>s</i>)	1.07 (<i>s</i>)
27	0.99 (<i>s</i>)	1.00 (<i>s</i>)
28	1.30 (<i>s</i>)	1.29 (<i>s</i>)
29	1.03 (<i>s</i>)	1.02 (<i>s</i>)
30	0.96 (<i>s</i>)	0.95 (<i>s</i>)

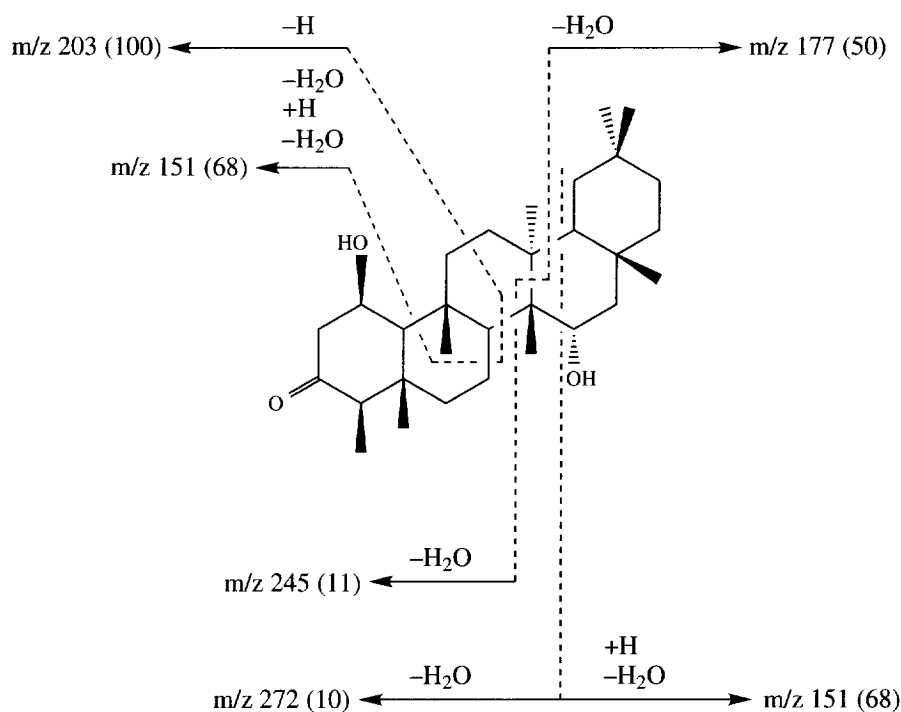
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1 R = β -OH, H

2 R = H₂

3 R = O



Scheme 1. EI mass spectral fragmentation of compound **1** (relative intensities are given in parentheses).

at δ 2.69 and 2.40 can be assigned to the diastereotopic C-2 methylene protons and their mutual connectivity has been established by decoupling studies. The multiplicity of each C-2 proton as double doublets can be rationalized in terms of a further splitting due to a third proton, i.e. at C-1.

Confirmation of this assignment as well as the location of the remaining hydroxyl group at C-1 comes from irradiation of the broad carbinol methine signal at δ 4.80 which affects the splitting pattern of both double doublets at δ 2.69 and 2.40 and simplifies them into AB doublets.

The orientation of the C-1 hydroxyl group as β and axial has been assigned from the following observation. In comparison with the tertiary methyl protons shifts of **2**, the shifts of the C-24 and C-25 tertiary methyl groups in **1** have been found to be deshielded considerably and appear at δ 1.10 and 1.33 i.e., 0.45–0.48 ppm higher than the corresponding shifts in **2** (Table 1). This deshielding effect can be explained in terms of a 1–3 syn-diaxial interaction of the 24 and 25 methyl groups with the axially oriented 1β hydroxyl group. Further evidence comes from the J values of the coupled doublets of C-2 protons. The J values of 4.6

Table 2. ^{13}C NMR data for compounds **1** and **2** (69.5 MHz, CDCl_3)

C	1	2
1	71.3	22.3
2	52.7	41.5
3	*	213.0
4	58.9	58.2
5	44.0†	42.0
6	43.8†	41.3
7	20.4	20.0
8	54.1	53.5
9	38.9	37.8
10	61.2	59.4
11	35.6	35.7
12	31.0	31.2
13	40.6	40.6
14	44.1	44.1
15	74.8	74.6
16	48.4	48.4
17	30.2	30.2
18	41.6	41.6
19	35.6	35.6
20	28.1	28.1
21	31.9	31.9
22	38.9	38.8
23	7.0	6.8
24	17.2	14.5
25	19.2	18.0
26	14.0	14.0
27	18.8	18.7
28	32.6	32.6
29	30.9	30.9
30	35.6	35.6

* Signal obscured due to poor signal to noise ratio.

† May be reversed within column.

fragmentation pattern (Scheme 1) are in agreement with the assigned structure thereby characterizing **1** as 1 β ,15 β -dihydroxyfriedelan-3-one.

EXPERIMENTAL

Collection, identification, and extraction of *Salacia beddomei* stem bark and the purification procedures used have been described previously [1, 2]. ^1H NMR: 270 MHz, CDCl_3 using TMS as int. standard; ^{13}C NMR: 69.5 MHz, CDCl_3 signal as reference, and the chemical shifts were assigned by comparison with compound **2**, [1]. EIMS: 70 eV.

Compound **1** was isolated from the CHCl_3 -EtOAc fractions of the EtOAc extract and eluted after betulin [2]. Evaporation of the fractions yielded a crude solid material (7 mg) contaminated with lipid impurities. Purification of the material by prep. TLC on silica gel using EtOAc-hexane yielded **1** as an amorphous material (ca 1 mg) which could not be crystallized.

1 β ,15 α -Dihydroxyfriedelan-3-one (**1**). ^1H NMR (270 MHz, CDCl_3): Table 1; ^{13}C NMR: Table 2; EIMS m/z (rel. int.): 458 $[\text{M}]^+$ (2), 440 $[\text{M} - \text{H}_2\text{O}]^+$ (12), 425 $[\text{M} - \text{H}_2\text{O} - \text{Me}]^+$ (10), 422 $[\text{M} - 2\text{H}_2\text{O}]^+$ (30), 407 $[\text{M} - 2\text{H}_2\text{O} - \text{Me}]^+$ (30), 272 (10), 245 (11), 231 (18), 218 (24), 203 (100), 189 (35), 177 (50), 151 (68), 107 (41).

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and 2.4 Hz can be explained in terms of axial-equatorial and equatorial-equatorial interaction of C-2 protons with an equatorially oriented 1 α proton. Moreover, the ^{13}C NMR data (Table 2) as well as the mass spectral