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SYNTHESIS OF SYRINGOLIDE 1 KETALS AND ESTERS FOR RECEPTOR STUDIES

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Abstract—The syringolides, specified by avirulence gene D from *Pseudomonas syringae*, elicit a disease defense reaction only in soybean cultivars carrying the *Rpg4* disease resistance gene. In order to identify structural features important for elicitor activity and construct highly labelled elicitor-active probes, several syringolide esters and ketals were synthesized. Alteration of the C-3 hemiketal group of syringolide 1 abolished elicitor activity, but ester derivatives based on the C-4' hydroxy group all retained activity. [125] I]4'-(2-iodo-3,4,5-trimethoxy-phenylacetyl) syringolide 1 was prepared for receptor studies. Copyright © 1996 Elsevier Science Ltd

INTRODUCTION

Genetic studies in plant-pathogen interactions have established a gene-for-gene relationsip in which disease resistance is often governed by a single dominant gene for disease resistance in the host plant and a single dominant gene for avirulence in the pathogen [1, 2]. The elicitor-receptor hypothesis proposes that pathogen biotypes carrying an avirulence gene produce a specific elicitor that is, in turn, only recognized by plants carrying the matching resistance gene [3-5]. Although there have been several specific elicitors described from plant pathogens, nothing is known about the putative plant receptors.

Avirulence gene D (avrD) cloned from Pseudomonas syringae pv. tomato [6, 7] causes Gram-negative bacteria expressing the gene to elicit the hypersensitive defense response (HR) on soybean plants carrying the disease resistance gene, Rpg4. Bacteria expressing avrD produce syringolides, extracellularly secreted specific elicitors which initiate the HR only in soybean plants harbouring Rpg4 [8]. It has therefore been speculated that syringolide receptors exist in soybean plants harbouring Rpg4.

Since receptors generally are present in low abundance in cells, receptor studies require that the ligand be suitably labelled [9]. For example, receptors for insulin and steroid hormones were first demonstrated using radioiodinated ligands in animal cells [10, 11]. Agonists and antagonists of altered biological activity

are also useful to demonstrate the specificity of ligand binding [9, 11].

Considerable progress has been made in the detection of plant binding sites for pathogen produced elicitors. For example, Renelt *et al.* [12] showed ligand specific binding of a glycoprotein elicitor to parsley protoplasts and microsomal membranes and Cheong and Hahn [13] identified plasma membrane binding sites in soybean cells for a hepta- β -glucoside elicitor. Frey *et al.* [14] obtained data indicating that a 70 kDa membrane protein is involved in binding of the hepta- β -glucoside elicitor.

Unfortunately, little progress has occurred in assessing plant binding sites for specific elicitors that are active only in plants carrying a complementary disease resistance gene. As a prerequisite to these studies with the syringolide elicitors, we have synthesized several derivatives of syringolide 1 (1, Scheme 1) and determined their elicitor activities. We also report the preparation of a mono-iodinated syringolide derivative (4) which retains high elicitor activity and should accordingly be useful for the preparation of radioactive adducts for binding studies with soybean cell fractions.

RESULTS AND DISCUSSION

Syringolide derivatives based on the C-3 hemiketal and the C-4' alcohol were examined. Ketals **1a** (benzyl), **1b** (methyl), **1c** (ethyl), and **1d** (2-hydroxyethyl) were readily prepared by treatment of **1** with the corresponding alcohols in the presence of dry Dowex/HCl. All ketal derivatives thus constructed by reaction at the C-3 position of **1** were devoid of detectable elicitor activity in soybean leaves (Table 1). It was

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Scheme 1. Synthesis of syringolide 1 derivatives. Bn = benzyl; TMPA = 3,4,5-trimethoxyphenyl acetyl; MPP = 3-(4-methoxyphenyl) propionyl; Suc = succinyl; HCP = 3-(4-hydroxycyclohexyl) propionyl.

particularly interesting that 1d, in which the end of the (C-3)-O chain was hydroxyl, also had no elicitor activity. The results indicate that the C-3 hemiketal group is essential for syringolide elicitor activity and

lead to the prediction that ketal derivatives should not function as competitors of syringolide binding to a putative receptor in soybean cells.

Syringolide 1 required protection of its hemiketal as

Table 1. Elicitor activity as measured by hypersensitive responses in leaves of various soybean cultivars infiltrated with syringolide 1 or its derivatives

Cpd	R at 3- <i>O</i>	R at 4'-O	Acme (rpg4)	Flambeau (Rpg4)	Merit (rpg4)	Harosoy (Rpg4)
1	Н	Н	-	+	_	+
1a	Bn	Н	4.00	_	-	_
1b	Me	Н	_		_	_
1c	Et	Н			-	-
1d	$HO(CH_2)_2$	Н	_	-	_	_
2a	Bn	TMPA				-
2b	Bn	MPP		_		-
2c	Bn	Succinyl				-
2d	Me	Succinyl	-	van	-	
3a	Н	TMPA	1 40	+	-	+
3b	Н	MPP	***	+		+
3c	Н	Succinyl	_	+	-	+
3d	Н	НСР	_	+	_	+
4	Н	Iodo-TMPA		+	-	+

Bn = benzyl; TMPA = 3,4,5-trimethoxyphenyl acetyl; MPP = 3-(4-methoxyphenyl) propionyl; HCP = 3-(4-hydroxycyclohexyl) propionyl. Water solutions of compounds $(3.68 \times 10^{-5} \text{ M})$ were infiltrated into primary soybean leaves. Reactions were observed after 24 hr: +, positive reaction; -, no reaction.

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a ketal before esterification (Scheme 1). Direct esterification of C-4' alcohol was not possible since common esterification methods using N,N'-dicyclohexylcarbodiimide (DCC), 1-(3-dimethylaminopropyl)-3ethylcarbodiimide methiodide (EDCI), 4-(dimethylamino)pyridine (DMAP) or pyridine destroyed the syringolide ring system. It was also destroyed by esterification attempts using acids (HCl, H2SO4) or cobalt(II) chloride. However, syringolide ketals were stable to esterification using base catalysis. Thus syringolide 1 ketal esters 2a, 2b, 2c and 2d were synthesized, in which the C-4' alcohol of 1a or 1b was esterified with 3,4,5-trimethoxyphenylacetic acid, (3-(4methoxyphenyl) propionic acid, or succinic anhydride (Scheme 1). The (C-3)-O-methyl ketal of 2d was very resistant to deprotection by acid hydrolysis. However, the (C-3)-O-benzyl protecting groups in 2a, 2b and 2c could be easily removed by hydrogenation to produce the desired syringolide hemiketal ester analogues 3a, **3b**, and **3c**. Hemiketal esters of syringolide 1 elicited the HR only in soybean cultivars harbouring Rpg 4. whereas the ketal esters were inactive (Table 1). Hemiketal esters 3a, 3b and 3c were also active elicitors of glyceollin in soybean cotyledons, while ketals 1a and 1b were not active (Table 2). We conclude that the (C-4')-hydroxyl group is not necessary for elicitor activity.

The hemiketal ester derivatives, 3a and 3b, were elicitor active and contained highly activated aromatic rings which could be iodinated using iodine and silver trifluoroacetate [15]. Compound 3a was converted in 95% yield to a single product, 4, which retained elicitor activity (Table 1). The DEI mass spectrum of 4 showed $[M]^+$ at m/z 606 and the ¹H NMR spectrum showed a single aromatic proton at δ 6.69, confirming monoiodination. Thus 4 was identified as 4'-(2-iodo-3,4,5-trimethoxyphenylacetyl) syringolide 1. The iodination reaction of 3b was not useful since multiple iodinated compounds were produced in poor yield.

The high yield of the iodinated product 4 indicates that these iodination reagents are suitable for the production of [125]4. Radioiodination was accordingly carried out using the method of Fowler *et al.* [16] to generate [125]I₂. The availability of highly radioactive syringolide derivatives will facilitate the search for

Table 2. Soybean cotyledon assay of elicitor activity of syringolide 1 and its derivatives

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Cpd	R at 3- <i>O</i>	R at 4'- <i>O</i>	Elicitor activity (EC ₅₀ , μM)
1	Н	Н	27
1a	Bn	Н	>400
1b	Me	Н	>400
3a	Н	TMPA	94
3b	H	MPP	89
3c	Н	Suc	31
4	Н	Iodo-TMPA	89

 EC_{s0} is half maximum induction of phytoalexin accumulation in cultivar Harosoy cotyledons.

syringolide receptors in soybean leaves. Furthermore, syringolide succinate **3c** should be particularly useful in the preparation of affinity columns. The inactive C-3 and certain of the C-4' derivatives of reduced activity will also be extremely valuable as competitors with radiolabelled syringolide 1 in ongoing binding studies.

EXPERIMENTAL

NMR spectra were recorded at 300 MHz ¹H and 75 MHz 13 C in CDCl₃ and referenced to TMS = 0. TLC was carried out on silica gel F using phosphomolybdic acid visualization at 110°. HPLC samples were prepurified using silica Sep-Paks or vacuum liquid chromatography (VLC). Small samples were purified by HPLC on a Maxsil 5 (4.6 mm × 250 mm) and larger samples on a Dynamax (25.4 mm × 250 mm) silica column using refractive index detection. All solvents and benzyl alcohol were distilled before use. CH₂Cl₂ was dried by refluxing over CaH₂ before distillation. Anhydrous Dowex/HCl was prepared by washing HCltreated Dowex with dry MeOH followed by evacuation to 1 mm Hg for 16 hr. Positive ion mass spectra were observed by desorption chemical ionization (DCI) using NH₃ or CH₄ or by desorption electron ionization (DEI) or fast atom bombardment (FAB) using mnitrobenzyl alcohol (NBA). Polyethylene glycol (PEG) or perfluorokerosene (PFK) served as reference for high resolution measurements.

Isolation of syringolide 1 (1). Synthetic liquid M9 glucose (0.2%) medium [17] supplemented with thiamine $(4 \mu g/ml)$, ampicillin $(75 \mu g/ml)$ and 0.4 mM isopropyl-thiogalactoside was adjusted to pH 5.0 with conc HCl. One litre of medium in a 21 Erlenmeyer was seeded with E. coli DH5 α cells carrying expression plasmid pPADRI with the class II allele from Pseudomonas syringae phaseolicola G-50 [18]. The cultures were shaken at 28° for approximately 21 hr. The cell-free culture fluids were extracted three times with EtOAc (ca 1/4 volume). The combined organic phases were dried with MgSO₄ then filtered and taken to dryness at 40°. The crude extracts, further purified by VLC and HPLC on silica gel [8], afforded 1 in yields of about 7.5 mg per litre of culture. It was stored as a waxy solid.

Synthesis of syringolide 1 ketals 1a, 1b, 1c, and 1d. Syringolide 1 was stirred overnight with vacuum-dried Dowex 50W-X8/HCl and the appropriate alcohol in about 3-fold excess [8]. Dry CH₂Cl₂ was added as a solvent for 1a and 1d. The reaction mixtures were filtered, concentrated in vacuo and purified by HPLC.

 4.40 (1H, d, J = 10.4, H-1'a), 4.55 (1H, d, J = 10.4, H-1'b), 4.58 (2H, s, H-9), 7.3 (5H, m, H-11, H-12, H-13, H-14, and H-15); ¹³C NMR: δ 14.0 (C-8), 22.4 (C-7), 23.4 (C-5), 31.8 (C-6), 33.4 (C-4), 59.7 (C-2), 62.7 (C-9), 74.2 (C-5'), 74.7 (C-4'), 74.9 (C-1'), 91.3 (C-3'), 98.1 (C-2'), 111.5 (C-3), 126.7 (C-11 and C-15), 127.4 (C-13), 128.5 (C-12 and C-14), 137.8 (C-10), 171.8 (C-1); DCIMS (NH₃), m/z: 380 [M + NH₄] + (4), 272 (14), 256 (16), 255 [M - C₆H₅CH₂O] + (100), 211 (3), 151 (14), 108 (7), 99 (4), 91 (57); HRDCIMS (NH₃), 380.2075 (C₂₀H₃₀NO₆ = 380.2073, Δ = -0.5 ppm).

3-O-Methyl syringolide 1 (1b). Using syringolide 1 (15 mg) in 3 ml of dry MeOH [8], 1b was prepared in nearly quantitative yield. It had a retention time of 27.9 min. on Maxsil eluted with 25% EtOAc in hexane at 1.5 ml/min. ¹H NMR (CDCl₃, 300 MHz): δ 0.88 (3H, t, J = 8.0 Hz, H-8), 1.30 (4H, bm, H-6 and H-7),1.45 (1H, m, H-5a), 1.60 (1H, m, H-5b), 1.78 (1H, m, H-4a), 1.92 (1H, m, H-4b), 3.10 (1H, s, H-2), 3.24 (3H, s, H-9), 3.82 (1H, dd, J = 10.0, 2.7, H-5'a), 4.03 (1H, d, J = 10.0, H-5'b), 4.30 (1H, d, J = 2.7, H-4'), 4.31 (1H, s, H-3'), 4.42 (1H, d, J = 10.4, H-1'a), 4.65 (1H, d, J = 10.4, H-1'b); ¹³C NMR: δ 14.0 (C-8), 22.4 (C-7), 23.5 (C-5), 31.8 (C-6), 32.4 (C-4), 48.5 (C-9), 59.5 (C-2), 74.2 (C-5'), 74.7 (C-4'), 74.8 (C-1'), 91.1 (C-3'), 98.2 (C-2'), 111.3 (C-3), 171.8 (C-1); DCIMS (CH_4) , m/z: 287 $[M + H]^+$ (4), 255 $[M - CH_3O]^+$ (100), 215 (52), 211 (15), 155 (11), 151 (16), 55 (10); HRDCIMS (CH₄), 287.1483 (C₁₄H₂₃O₆ = 287.1495, $\Delta = 4.1 \text{ p.p.m.}$).

3-O-Ethyl syringolide 1 (1c). The ethyl ketal of syringolide I was obtained in nearly quantitative yield. It had a retention time of 20.5 min. on Maxsil eluted with 25% EtOAc in hexane at 1.5 ml/min. ¹H NMR $(CDCl_3, 300 \text{ MHz})$: $\delta 0.90 (3H, t, J = 8.0 \text{ Hz}, H-8)$, 1.17 (3H, t, J = 7.7, H-10), 1.30 (4H, bm, H-6 and H-7), 1.45 (1H, m, H-5a), 1.60 (1H, m, H-5b), 1.78 (1H, m, H-4a), 1.92 (1H, m, H-4b), 3.08 (1H, s, H-2). 3.47 (1H, dq, J = 16.6, 7.7, H-9a), 3.58 (1H, dq, J = 16.6, 7.7, H-9b), 3.84 (1H, dd, J = 10.4, 2.7, H-9b) 5'a), 4.04 (1H, dd, J = 10.4, 1.0, H-5'b), 4.30 (2H, m, H-3' and H-4'), 4.45 (1H, d, J = 10.3, H-1'a), 4.65 (1H, d, J = 10.3, H-1'b); ¹³C NMR: δ 14.0 (C-8), 15.3 (C-10), 22.4 (C-7), 23.4 (C-5), 31.8 (C-6), 33.2 (C4), 56.4 (C-9), 59.7 (C-2), 74.1 (C-5'), 74.5 (C-4'), 75.0 (C-1'), 91.1 (C-3'), 98.3 (C-2'), 111.1 (C-3), 171.8 (C-1), CIMS (NH₃), m/z: 318 [M + NH₄]⁺ (5), 272 (35), 256 (15), 255 $[M - CH_3CH_2O]^+$ (100), 229 (17), 151 (6), 136 (5), 55 (7); HRCIMS (NH₃), 318.1929 $(C_{15}H_{28}NO_6 = 318.1917, \Delta = 4.1 \text{ p.p.m.}).$

3-O-(2-Hydroxyethyl) syringolide 1 (1d). By reaction of ethylene glycol with 1 (30 mg) a nearly quantitative yield of 1d was obtained. Its retention time was 29 min on the Dynamax silica column (iso-PrOH–EtOAc–hexane, 1.5:43.5:55) at 6 ml/min. ¹H NMR (CDCl₃, 300 MHz) δ 0.90 (3H, t, J = 8.0, H-8), 1.30 (4H, bm, H-6 and H-7), 1.45 (1H, m, H-5a), 1.60 (1H, m, H-5b), 1.78 (1H, m, H-4a), 1.92 (1H, m, H-4b), 3.13 (1H, s, H-2), 3.57 (1H, m, H-9a), 3.63 (1H, m, H-9b), 3.73 (2H, m, H-10), 3.85 (1H, dd, J = 10.5, 2.5, H-5'a),

4.05 (1H, *d*, J = 10.5, H-5'b), 4.32 (1H, *d*, J = 2.5, H-4'), 4.43 (1H, *s*, H-3'), 4.45 (1H, *d*, J = 10.4, H-1'a), 4.68 (1H, *d*, J = 10.4, H-1'b); C¹³ NMR: δ 14.0 (C-8), 22.4 (C-7), 23.3 (C-5), 31.8 (C-6), 33.1 (C-4), 59.1 (C-10), 59.3 (C-2), 63.1 (C-9), 74.2 (C-5'), 74.7 (C-4'), 74.9 (C-1'), 91.1 (C-3'), 98.2 (C-2'), 112.0 (C-3), 171.5 (C-1); CIMS (NH₃), m/z: 334 [M + NH₄]⁺ (3), 272 (17), 256 (20), 255 [M – OCH₂CH₂OH]⁺ (100), 151 (12), 87 (69); HRCIMS (NH₃), 334.1879 (C₁₅H₂₈NO₇ = 334.1866, $\Delta = -3.9$ p.p.m.).

Synthesis of 3-O-benzyl-4'-(3,4,5-trimethoxyphenylacetyl) syringolide 1 (2a). To a solution of 1a (14 mg, 0.04 mmol) in 500 μ l dry CH₂Cl₂ were added 3,4,5trimethoxyphenylacetic acid (9 mg, 0.04 mmol), 1-(3dimethylaminopropyl)-3-ethylcarbodiimide methiodide (EDCI, 15 mg) and 4-dimethylaminopyridine (DMAP, 1.8 mg) and stirred at room temperature. Reaction progress was monitored by TLC (CHCl₃/MeOH 19:1). After 16 hr the solution was diluted with 5 ml of CHCl₃ and washed with 1% HCl. The CHCl3 phase was collected and dried with MgSO₄. The crude ketal ester, 2a, was prepurified on a silica Sep-Pak by elution with 10 ml of 20% EtOAc-hexane; the yield was 87%. It had a retention time of 12.5 min. on Maxsil eluted with 25% EtOAc in hexane at 1.5 ml/min. H NMR (CDCl₃, 300 MHz): δ 0.90 (3H, t, J = 8, H-8), 1.30 (4H, bm, H-6 and H-7), 1.45 (1H, m, H-5a), 1.63 (1H, m, H-5b), 1.90 (1H, m, H-4a), 2.06 (1H, m, H-4b), 3.18 (1H, s, H-2), 3.56 (2H, d, J = 0.8, H-7'), 3.83 (6H, s, C-3"-OMe and C-5"-OMe), 3.86 (3H, s, C-4"-OMe), 3.91 (1H, dd, J = 11.1, 2.8, H-5'a), 4.11 (1H, dd, J = 11.1,0.8, H-5'b), 4.24 (1H, d, J = 0.8, H-3'), 4.32 (1H, d, J = 10.4, H-1'a), 4.41 (1H, d, J = 10.4, H-1'b), 4.58 (2H, s, H-9), 5.18 (1H, d, J = 2.8, H-4'), 6.46 (2H, s,H-2"), 7.30 (5H, m, H-11, H-12, H-13, H-14 and H-15); ¹³C NMR: δ 14.0 (C-8), 22.4 (C-7), 23.4 (C-5), 31.8 (C-6), 33.3 (C-4), 41.2 (C-7'), 56.1 (C-3"-OCH3 and C-5"-OCH₃), 59.6 (C-2), 60.9 (C-4"-OMe), 62.8 (C-9), 71.4 (C-5'), 74.4 (C-1'), 77.2 (C-4'), 89.0 (C-3'), 98.4 (C-2'), 106.2 (C-2" and C-6"), 111.8 (C-3), 126.6 (C-11 and C-15), 127.5 (C-13), 128.5 (C-12 and C-14), 128.7 (C-1"), 137.6 (C-4"), 137.6 (C-10), 153.3 (C-3" and C-5"), 170.4 (C-1), 171.3 (C-6'); DEIMS, m/z: 570 $[M]^+$ (38), 463 $[M - C_6H_5CH_2O]^+$ (12), 226 (27), 181 (51), 165 (28), 107 (15), 91 (100), 55 (22); HRDEIMS, 570.2470 ($C_{31}H_{38}O_{10} = 570.2465$, $\Delta = 0.9 \, \text{p.p.m.}$).

Synthesis of 3-O-benzyl-4'-3-(4-methoxyphenyl-propionyl) syringolide 1 (**2b**). Ketal ester **2b** was synthesized from **1a** (18 mg) by the same method as **2a** except substituting 3-(4-methoxyphenyl) propionic acid for 3,4,5-trimethoxyphenylacetic acid. The yield was 78%. It had a retention time of 6 min. on Maxsil eluted with 25% EtOAc in hexane at 1.5 ml/min. ¹H NMR (CDCl₃, 300 MHz): δ 0.90 (3H, t, J = 8.0, H-8), 1.30 (4H, bm, H-6 and H-7), 1.45 (1H, m, H-5a), 1.63 (1H, m, H-5b), 1.90 (1H, m, H-4a), 2.06 (1H, m, H-4b), 2.60 (2H, m, H-7'), 2.86 (2H, t, t = 7.5, H-8'), 3.16 (1H, t s, H-2), 3.70 (3H, t s, C-4"-OMe), 3.88 (1H, t dd, t = 11.1, 3.0, H-5'a), 4.04 (1H, t d, t = 11.1, H-5'b), 4.17 (1H, t s, H-3'), 4.29 (1H, t d, t = 10.3, H-1'a), 4.35 (1H, t d,

J = 10.3, H-1'b), 4.58 (2H, s, H-9), 5.13 (1H, d, J =2.8, H-4'), 6.78 (2H, d, J = 8.6, H-3" and H-5"), 7.08 (2H, d, J = 8.6, H-2" and H-6"), 7.30 (5H, m, H-11,H-12, J-13, H-14 and H-15); 13 C NMR: δ 14.0 (C-8), 22.4 (C-7), 23.4 (C-5), 30.0 (C-8'), 31.8 (C-6), 33.3 (C-4), 35.9 (C-7'), 55.2 (C-4"-OMe), 59.6 (C-2), 62.9 (C-9), 71.8 (C-5'), 74.5 (C-1'), 77.2 (C-4'), 89.1 (C-3'), 98.4 (C-2'), 111.7 (C-3), 113.9 (C-3" and C-5"), 126.8 (C-11 and C-15), 127.5 (C-13), 128.5 (C-12 and C-14), 129.2 (C-2" and C-6"), 131.9 (C-1"), 137.7 (C-10), 158.3 (C-4"), 171.5 (C-1), 171.9 (C-6'); DCIMS (NH₃), m/z: 542 [M + NH₄]⁺ (6), 417 [M - $C_6H_4CH_7$] (68), 198 (17), 180 (30), 134 (13), 121 $[CH_3OC_6H_4CH_2]^+$ (100), 108 (15), 91 (62), 78 (16), 65 (10), HRDCIMS (NH₄), 542.2761 ($C_{30}H_{40}NO_8 =$ 542.2754, $\Delta = -1.3$ p.p.m.).

Synthesis of 3-O-benzyl-4'-succinyl syringolide 1 (2c). Succinic anhydride (5 mg) and DMAP (2 mg) were added to 1a (10 mg) in 600 μ l of CDCl₃. Followed by NMR, the esterification was complete after 72 hr. The crude mixture was diluted with 6 ml of CHCl, and washed with 6 ml of 2% HCl. The CHCl, phase was collected and dried with MgSO₄. After evaporation 12 mg (96%) of 2c was obtained and purified by HPLC. It had a retention time of 13 min. on Maxsil eluted with 25% EtOAc in hexane at 1.5 ml/ min. ¹H NMR (CDCl₃, 300 MHz): δ 0.90 (3H, t, J = 8H-8), 1.30 (4H, bm, H-6 and H-7), 1.40 (1H, m, H-5a). 1.60 (1H, m, H-5b), 1.90 (1H, bm, H-4a), 2.03 (1H, m. H-4b), 2.63 (4H, m, H-7' and H-8'), 3.17 (1H, s, H-2), 3.90 (1H, dd, J = 11.1, 2.9, H-5'a), 4.11 (1H, d, J =11.1, H-5'b), 4.27 (1H, s, H-3'), 4.37 (1H, d, J = 10.4, H-1'a), 4.51 (1H, d, J = 10.4, H-1'b), 4.58 (2H, s. H-9), 5.18 (1H, d, J = 2.8, H-4'), 7.3 (5H, m, H-11, H-12, H-13, H-14 and H-15); ¹³C NMR: 14.0 (C-8), 22.4 (C-7), 23.4 (C-5), 28.4 (C-8'), 28.8 (C-7'), 31.8 (C-6), 33.3 (C-4), 59.6 (C-2), 62.9 (C-9), 71.7 (C-5'), 74.6 (C-1'), 77.6 (C-4'), 88.9 (C-3'), 98.5 (C-2'), 112.0 (C-3), 126.8 (C-11 and C-15), 127.4 (C-13), 128.5 (C-12 and C-14), 137.7 (C-10), 171.1 (C-1), 171.6 (C-6'), 177.3 (C-9'); FABMS (NBA/NaCl), m/ z: 485 $[M + Na]^+$ (34), 445 (10), 413 (15), 355 [M - $C_6H_5CH_7O]^+$ (100), 255 (21), 219 (20), 193 (21), 123 (82); HRFABMS (NBA/NaCl/PEG) 485.1801 $(C_{24}H_{30}O_9Na = 485.1788, \Delta = -2.8 \text{ p.p.m.}).$

Synthesis of 3-O-methyl-4'-succinyl syringolide 1 (2d). This synthesis was analogous to that of 2c but started with 1b instead of 1a. The yield was 94%. It had a retention time of 8.5 min. on Maxsil eluted with 25% EtOAc in hexane at 1.5 ml/min. H NMR (CDCl₃, 300 MHz): δ 0.90 (3H, t, J = 8, H-8), 1.30 (4H, m, H-6 and H-7), 1.40 (1H, m, H-5a), 1.60 (1H, m, H-5b), 1.80 (1H, m, H-4a), 1.93 (1H, m, H-4b), 2.68 (4H, m, H-7' and H-8'), 3.12 (1H, s, H-2), 3.23 (3H, s, H-9), 3.90 (1H, dd, J = 11.1, 2.9, H-5'a), 4.11 (1H, d, J = 11.1,H-5'b), 4.37 (1H, s, H-3'), 4.40 (1H, d, J = 10.4, H-1'a), 4.60 (1H, d, J = 10.4, H-1'b), 5.18 (1H, d, J = 2.9, H-4'); ¹³C NMR: δ 14.0 (C-8), 22.4 (C-7), 23.2 (C-5), 28.6 (C-8'), 28.8 (C-7'), 31.8 (C-6), 32.3 (C-4), 46.8 (C-9), 59.5 (C-2), 71.7 (C-5'), 74.5 (C-1'), 77.2 (C-4'), 88.7 (C-3'), 98.5 (C-2'), 111.6 (C-3), 171.1 (C-1), 171.6 (C-6'), 177.3 (C-9'); DEIMS, m/z: 386 [M] $^{+}$ (2), 355 [M $^{-}$ MeO] $^{+}$ (44), 315 (71), 257 (7), 165 (29), 155 (18), 139 (14), 125 (10), 109 (35), 101 [COCH $_2$ CH $_2$ COOH] $^{+}$ (100), 81 (13), 73 (27), 55 (53); HRDEIMS, 386.1567 (C $_{18}$ H $_{26}$ O $_{9}$ = 386.1577, Δ = 2.5 p.p.m.).

Synthesis of 4'-(3,4,5-trimethoxyphenylacetyl) syringolide 1 (3a). Benzyl ketal ester 2a (6 mg in 10 ml of EtOAc) was hydrogenated by adding 20 mg of 10% Pd/C catalyst and stirring under H₂ atmosphere at room temp. for 3 hr. The resulting soln. of hemiketal ester 3a was then filtered through Whatman No. 2 paper and solvents were removed using a rotavap. The reaction was quantitative. It had a retention time of 21 min. on Maxsil eluted with 25% EtOAc in hexane at 1.5 ml/min. 1 H NMR (CDCl₃, 300 MHz): δ 0.90 (3H, t, J = 8, H-8, 1.30 (4H, bm, H-6 and H-7), 1.50 (2H, m, H-5), 1.93 (2H, t, J = 7.4, H-4), 3.09 (1H, s, H-2), 3.58 (2H, s, H-7'), 3.84 (3H, s, C-4"-OMe), 3.86 (6H, s, C-3"-OMe and C-5"-OMe), 3.90 (1H, dd, J = 11.2, 2.8, H-5'a), 4.12 (1H, d, J = 11.2, H-5'b), 4.51 (1H, s, H-3'), 4.36 (1H, d, J = 10.4, H-1'a), 4.54 (1H, d, J = 10.4, H-1'b), 5.16 (1H, d, J = 2.8, H-4'), 6.49 (2H, s, H-2" and H-6"); 13 C NMR: δ 14.0 (C-8), 22.5 (C-7), 23.1 (C-5), 31.5 (C-6), 38.7 (C-4), 41.4 (C-7'), 56.2 (C-3"-OMe and C-5"-OMe), 58.9 (C-2), 60.9 (C-4"-OMe), 71.8 (C-5'), 74.3 (C-1'), 77.2 (C-4'), 89.1 (C-3'), 98.0 (C-2'), 106.3 (C-2" and C-6"), 108.5 (C-3), 128.7 (C-1"), 137.6 (C-4"), 153.4 (C-3" and C-5"), 170.5 (C-6'), 171.8 (C-1); DEIMS, m/z: 480 [M]⁺ (8), 226 (100), 211 (31), 181 (64), 167 (20), 137 (10), 121 (15), 99 (19), 82 (11), 71 (25), 55 (50); HRDEIMS, 480.1990 (C₂₄H₃₂O₁₀ = 480.1995, Δ = 1.1 p.p.m.).

Synthesis of 4'-[3-(4-methoxyphenyl) propionyl syringolide] 1 (3b). From hydrogenation of 2b (12 mg) using 10% Pd/C catalyst (20 mg), 3b was obtained by the procedure described for 3a. The reaction, followed by silica TLC (CHCl₃-MeOH 19:1), was complete in 3 hr. and the yield of 3b was quantitative. It had a retention time of 26 min. on Maxsil eluted with 25% EtOAc in hexane at 1.5 ml/min. H NMR (CDCl₃) 300 MHz): δ 0.91 (3H, t, J = 8.3, H-8), 1.30 (4H, bm, H-6 and H-7), 1.50 (2H, m, H-5), 1.93 (2H, t, J = 7.6, H-4), 2.64 (2H, m, H-7'), 2.90 (2H, t, J = 7.2, H-8'), 3.08 (1H, s, H-2), 3.79 (3H, s, C-4"-OMe), 3.87 (1H, dd, J = 11.0, 2.9, H-5'a), 4.06 (1H, d, J = 11.0, H-5'b), 4.37 (1H, d, J = 10.4, H-1'a), 4.44 (1H, s, H-3'), 4.54 (1H, d, J = 10.4, H-1'b), 5.12 (1H, d, J = 2.9, H-4'), 6.85 (2H, d, J = 8.5, H-3" and H-5"), 7.11 (2H, d, J = 8.5, H-2" and H-6"); ¹³C NMR: δ 14.0 (C-8), 22.5 (C-7), 23.1 (C-5), 30.1 (C-8'), 31.5 (C-6), 36.0 (C-7'), 38.7 (C-4), 55.3 (C-4"-OCH₃), 58.9 (C-2), 71.9 (C-5'), 74.4 (C-1'), 77.1 (C-4'), 89.2 (C-3'), 97.9 (C-2'), 108.4 (C-3), 114.0 (C-3" and C-5"), 129.3 (C-2" and C-6"), 131.9 (C-1"), 158.3 (C-4"), 171.8 (C-1), 172.0 (C-6'); DEIMS, m/z: 434 $[M]^+$ (1), 180 (21), 134 (14), 121 $[CH_3OC_6H_4CH_2]^+$ (100), 91 (26), 77 (17), 71 (11), 55 (15); HRDEIMS 434.1928 ($C_{23}H_{30}O_9 =$ 434.1941, $\Delta = 2.9$ p.p.m.).

Synthesis of 4'-succinyl syringolide 1 (3c). Hydrogenation of 2c (10 mg), by the same method as 3a,

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gave 3c in quantitative yield. It had a retention time of 43 min. on Maxsil with 25% EtOAc in hexane flowing at 1.5 ml/min. 1 H NMR (CDCl₃, 300 MHz): δ 0.90 (3H, t, J = 8, H-8), 1.30 (4H, m, H-6 and H-7), 1.50(2H, m, H-5), 1.92 (2H, t, J = 8.1, H-4), 2.67 (4H, m,H-7' and H-8'), 3.10 (1H, s, H-2), 3.89 (1H, dd, J = 11.1, 3.1, H-5'a), 4.14 (1H, d, J = 11.1, H-5'b), 4.44 (1H, t, J = 10.5, H-1'a), 4.59 (1H, s, H-3'), 4.64 (1H, d, J = 10.5, H-1'b), 5.18 (1H, d, J = 3.1, H-4'); ¹³C NMR: δ 14.0 (C-8), 22.5 (C-7), 23.2 (C-5), 28.8 (C-7'), 28.9 (C-8'), 31.6 (C-6), 38.6 (C-4), 59.0 (C-2), 71.6 (C-5'), 74.6 (C-1'), 77.1 (C-4'), 89.0 (C-3'), 98.1 (C-2'), 108.5 (C-3), 171.3 (C-6'), 172.2 (C-1), 176.8 (C-9'); FABMS (NBA), m/z: 371 $[M-H]^+$ (62), 325 (51), 269 (51), 255 (100), 222 (49)' HRFABMS (NBA/PFK), 371.1344 ($C_{17}H_{23}O_9 = 371.1342$, $\Delta = -$

Synthesis of 4'-[3-(4-hydroxycyclohexyl)propionyl] syringolide 1 (3d). When 50 mg of Pd/C was stirred under H₂ atmosphere with **2b** (10 mg, 0.019 mmol) in 10 ml of EtOAc overnight, 3d was synthesized. After the soln was filled and evapd, 3d was isolated by HPLC (13.4 min) using 40% EtOAc in hexane on a Maxsil silica column at 1.5 ml/min. The yield was 3 mg (37%). H NMR (CDCl₃, 300 MHz): δ 0.91 (3H, t, J = 8.4, H-8), 1.30 (4H, m, H-6 and H-7), 1.40–1.80 (9H, m, H-1", H-2", H-3", H-5" and H-6"), 1.50 (2H, m, H-5), 1.60 (2H, m, H-8'), 1.93 (2H, t, J = 7.6, H-4), 2.38 (2H, t, J = 7.2, H-7'), 3.10 (1H, s, H-2), 3.90 (1H, dd, J = 11.1, 3.0, H-5'a), 4.00 (1H, m, H-4"), 4.10 (1H, d, J = 11.1, H-5'b), 4.45 (1H, d, J = 10.4, H-1'a), 4.60 (1H, s, H-3'), 4.69 (1H, d, J = 10.4, H-1'b), 5.14 (1H, d)d, J = 3.0, H-4'); ¹³C NMR: δ 14.0 (C-8), 22.5 (C-7), 23.2 (C-5), 26.5 (C-2" and C-6"), 31.1 (C-8'), 31.6 (C-6), 31.8 (C-1"), 32.2 (C-3" and C-5"), 35.9 (C-7'), 38.7 (C-4). 58.9 (C-2), 66.7 (C-4"), 72.0 (C-5'), 74.5 (C-1'), 77.2 (C-4'), 89.3 (C-3'), 98.0 (C-2'), 109.5 (C-3), 171.5 (C-1), 171.8 (C-6'); DCIMS (NH₃), m/z: 444 $[M + NH_4]^+$ (4), 409 (19), 391 (10), 274 (14), 255 $[M - COCH_2CH_2C_6H_{10}OH]^+$ (95), 237 (63), 225 (20), 211 (44), 190 (75), 155 [COCH₂CH₂C₆H₁₀OH] (77), 136 (61), 126 (16), 108 (25), 94 (100), 81 (52), 69 (22), 55 (75), HRDCIMS (NH₃), 444.2609 $(C_{22}H_{38}NO_8 = 444.2597, \Delta = -2.6 \text{ p.p.m.}).$

of 4'-(2-iodo-3,4,5-trimethoxyphenyl-Synthesis acetyl) syringolide 1 (4). Iodine (3.1 mg) and AgOCOCF, (4.3 mg) were added to **3a** (6.9 mg) in 0.4 ml dry CHCl₃. They were stirred 10 min at room temp, then diluted with 8 ml EtOAc and washed successively with 8 ml of 5% HCl and satd NaCl. The organic layer was dried over MgSO₄, filtered and evapd in vacuo to afford 4 in 95% yield. The compound eluted from a Maxsil silica column in 14.5 min using eluant 25% EtOAc in hexane and flow rate 1.5 ml/min. ¹H NMR (CDCl₃, 300 MHz): δ 0.90 (3H, t, J = 8, H-8), 1.30 (4H, bm, H-6 and H-7), 1.50 (2H, m, H-5), 1.93 (2H, t, J = 7.0, H-4), 3.09 (1H, s, H-2), 3.82 (2H, s, H-7'), 3.88 (6H, s, C-3"-OMe and C-5"-OMe), 3.84 (3H, s, C-4"-OMe), 3.90 (1H, dd, J = 11.2, 2.7, H-5'a),4.18 (1H, d, J = 11.2, H-5'b), 4.62 (1H, s, H-3'), 4.35

(1H, d, J = 10.4, H-1'a), 4.58 (1H, d, J = 10.4, H-1'b), 5.20 (1H, d, J = 2.7, H-4'), 6.69 (1H, s, H-6"); 13 C NMR, δ 14.0 (C-8), 22.5 (C-7), 23.1 (C-5), 31.5 (C-6), 38.7 (C-4), 46.3 (C-7'), 56.2 (C-3"-OMe), 59.0 (C-2), 60.8, (C-4"-OMe), 61.0 (C-5"-OMe), 71.8 (C-5'), 74.3 (C-1'), 77.2 (C-4'), 89.0 (C-3' and C-2"), 97.9 (C-2'), 108.5 (C-3), 110.1 (C-6"), 132.5 (C-1'), 141.8, (C-4"), 153.4 (C-5"), 153.7 (C-3"), 169.6 (C-6'), 171.8 (C-1); CIMS (NH₃), m/z: 606 [M] $^+$ (9), 353 (13), 352 (100), 334 (14), 307 (60), 226 (38), 211 (15), 181 (40), 167 (17), 165 (20), 149 (17), 121 (36), 99 (34), 55 (50); HRCIMS (NH₃), 606.0970 (C₂₄H₃₁O₁₀I = 606.0962, Δ = -1.3 p.p.m.).

Synthesis of [125 I]4. To 120 μ l of a 5% solution of KIO₃, 100 μ l of [125 I]Nal (1.1 mCi of radioactivity) and 500 μ l of CHCl₃ was added 40 μ l of 9N H₂SO₄. After 10 min the resulting CHCl₃ solution of [125 I]I₂ was dried by filtration through a 0.5 cm \times 0.5 cm column of Sephadex G-25. To 2 μ g of 3a and 2 μ g of AgO(CO)CF₃ (previously dissolved in 150 μ l of ether and blown dry in the presence of 3a) was added the dry CHCl₃ solution of [125 I]I₂. This was stirred at 40° for about 10 min. Free [125 I]I₂ was removed by washing with 100 μ l of 5% Na₂S₂O₃. The [125 I]4 was purified on a silica Sep-Pak using CHCl₃ as the eluting solvent, and after removal of CHCl₃ it was stored at -20° in a glass-lined lead container.

Determination of the soybean hypersensitive response. The elicitor activity of synthesized derivatives of syringolide 1 was qualitatively determined by the soybean leaf bioassay [17, 18]. The derivatives were dissolved at known concentrations in EtOH and diluted with water to concentration of 3.68×10^{-5} M for comparison of activity with syringolide 1. Water with the same conc of EtOH was used as a negative control. At 24 hr. after infiltration, the leaves were scored for the appearance of necrosis which occurred only in cultivars carrying the Rpg4 disease resistance gene.

Soybean cotyledon bioassay. The soybean cotyledon bioassay was performed as previously described [19]. This assay monitors phytoalexin production as an indicator of the hypersensitive defence response and permits quantitative determination of elicitor activity. Assay solutions were applied to wounded cotyledons of cultivar Harosoy, and the amounts of phytoalexin glyceollin were measured after 24 hr. at room temperature. Water was used as a negative control in all cotyledon assays.

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