

ROSMARINIC ACID FROM *LAVANDULA VERA* MM CELL CULTURE

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(Received in revised form 23 May 1996)

Key Word Index—*Lavandula vera*; Lamiaceae; plant cell culture; rosmarinic acid; caffeic acid.**Abstract**—Rosmarinic acid was isolated as the main phenolic component of *Lavandula vera* MM cell culture. It was identified by means of TLC, HPLC, ¹H NMR, ¹³C NMR and mass spectroscopy. Copyright © 1996 Elsevier Science Ltd

INTRODUCTION

In vitro cultured cells of *Lavandula vera*, which were being examined as a potential source of biotin [1, 2], were found out to synthesize and excrete a blue pigment into the cultural medium [3–5]. This was identified by Banthorpe *et al.* [6] as an enol ester of caffeic acid. We also obtained cell cultures of this species, using *L. vera* plants that produce phenolic compounds with antimicrobial and antioxidant activities [7]. This paper deals with the identification and quantitative determination of the main phenolics found in this cell culture.

RESULTS AND DISCUSSION

A methanolic extract from fresh cells of *L. vera* MM grown in LS medium [8] for nine days was used. The UV spectrum showed maxima between 280 and 340 nm, indicative of the presence of phenolic acids. Rosmarinic (RA), caffeic (CA), *p*-coumaric and ferulic acids were identified in the extracts by TLC. The

extracts contained a comparatively large amount of RA, smaller amounts of CA and traces of *p*-coumaric and ferulic acids (Table 1).

Banthorpe and co-workers [6, 9] reported that the main phenolic compound synthesized by *L. vera* callus culture was a blue pigment, which they identified as a complex of Fe(II) with the isomers of an enol ester formed by condensation of dopaldehyde with caffeic acid. Our *L. vera* MM strain, however, produced mainly RA. The latter was isolated from ethylacetate extracts of cell biomass as a yellowish powder (see Experimental) and its identity was confirmed by means of HPLC with a photodiode array detector, ¹H NMR, ¹³C NMR and mass spectroscopy [10–12]. This is the first report on the synthesis of RA by a *L. vera* cell culture, although RA is known to be synthesized by cultures of other species of the Lamiaceae [13–15].

EXPERIMENTAL

Culture method. Callus culture strain *L. vera* MM was originally derived from the leaves of *L. vera* plants in LS basal agar medium [8] containing 0.2 mg l⁻¹ 2,4-dichlorophenoxyacetic acid and 3% sucrose. It was subcultured on the same medium at 25–28° in the dark for more than 3 years at 2 week intervals. *L. vera* MM cell suspension cultures were grown using the same liquid medium [8] in Erlenmeyer flasks with 1/5 net vol. on the shaker (11.6 rad s⁻¹). It was subcultured at intervals of 1 week for 1 year. Cell biomass was harvested for chemical analyses 9 days after inoculation.

Extraction and isolation. Cells (20 g fr. wt) were extracted with MeOH (3 × 100 ml). The combined extracts were evapd *in vacuo* and the dry residue was extracted again with EtOAc (2 × 50 ml). The EtOAc extracts were evd to dryness. The residue was dissolved

Table 1. Content of phenolic acids in cell biomass of *Lavandula vera* MM

| Compound | Content* (mg g ⁻¹ dry wt) |
|-------------------------|---|
| Rosmarinic acid | 5.15 ± 0.73 |
| Caffeic acid | 0.37 ± 0.02 |
| <i>p</i> -Coumaric acid | Trace |
| Ferulic acid | Trace |

*Data represent the mean values of five different flasks and standard deviation.

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in a minimum vol. of MeOH and subjected to chromatography.

TLC and HPLC. MeOH and EtOAc extracts were fractionated by TLC (silica gel GF 254) using the following systems: (1) toluene–EtOAc–HOAc–MeOH–H₂O (20:60:10:20:3); (2) toluene–EtOAc–HOAc (4:5:2) and (3) CHCl₃–MeOH–H₂O (30:11:2).

RA was isolated from the EtOAc extract by prep. TLC on silica gel GF₂₅₄ using solvent system 1.

HPLC analyses were carried out on a Perkin Elmer liquid chromatograph with UV detection. The conditions were: analyt. column PEC₁₈ (10 µm, 250 × 4.6 mm i.d.); mobile phase: 2% HOAc (A) and 2% HOAc–MeCN (7:3) were used and a linear gradient 70–30% A over 40 min was applied; flow rate: 1 ml min⁻¹; pressure: 10 MPa; detection at 280 nm. Authentic samples of RA, CA and *p*-coumaric and ferulic acids were used as markers. The external standard method was used for quantitative determination of the analysed compounds.

Acknowledgment—The authors gratefully acknowledge the financial support of the National Research Foundation of Bulgaria.

REFERENCES

1. Watanabe, K., Yano S. and Jamada, J. (1983) *Phytochemistry* **21**, 513.
2. Watanabe, K. and Jamada, J. (1982) *Plant Cell Physiol.* **23**, 1453.
3. Watanabe, K., Sato, F., Furuta, M. and Yamada, Y. (1985) *Agric. Biol. Chem.* **49**, 533.
4. Nakajima, H., Sonomoto, K., Usni, N., Sato, F., Yamada, Y., Tanaka, A. and Fukui, S. (1985) *J. Biotechnol.* **2**, 107.
5. Nakajima, H., Sonomoto, K., Movikana, H., Satu, F., Ishimura, K., Jamada, Y. and Tanaka, A. (1986) *Appl. Microbiol. Biotechnol.* **24**, 266.
6. Banthorpe, D. V., Bilyard, H. J. and Watson, D. G. (1985) *Phytochemistry* **24**, 2677.
7. Ilieva, M., Kozhuharova, L., Pavlov, A. and Kovatcheva, E. (1994) Proceedings of the International Euro Food Tox IV Conference *Bioactive Substances in Food of Plant Origin*, II. Centre for Agrotechnology and Veterinary Sciences, Polish Academy of Sciences, Olsztyn, Poland.
8. Linsmayer, E. F. and Skoog, F. (1965) *Physiol. Plant.* **18**, 100.
9. Banthorpe, D. V., Bilyard, H. J. and Brown, G. D. (1989) *Phytochemistry* **28**, 2109.
10. Newbert, L. A., Breneman, W. R. and Carmack, M. (1975) *J. Org. Chem.* **40**, 1804.
11. Kelly, C., Harruff, R. C. and Carmack, M. (1976) *J. Org. Chem.* **41**, 449.
12. Fukui, H., Yazaki, K. and Tabata, M. (1984) *Phytochemistry* **23**, 2398.
13. De-Ekmankul, W. and Ellis, B. E. (1988) in *Biotechnology in Agriculture and Forestry*, Vol. 4, *Medical and Aromatic Plants* (Bajaji, J. P., ed), p. 310. Springer, Berlin.
14. Zenk, M. N., El-Shigi, H. and Ulbrich, B. (1977) *Naturwissenschaften* **64**, 585.
15. Hyppolyte, L., Marin, B., Baccen, J. C. and Jonard, R. (1992) *Plant Cell Rep.* **11**, 109.