



CULARINE *N*-OXIDE ALKALOIDS FROM *CERATOCAPNOS HETEROCARPA*

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Key Word Index—*Ceratocarpus heterocarpa*; Fumariaceae; (+)-*cis*-sarcocapnine *N*-oxide; (+)-*cis*-cularine *N*-oxide; sarcocapnidine *N*-oxide.

Abstract—Two new alkaloids, (+)-*cis*-sarcocapnine *N*-oxide and (+)-*cis*-cularine-*N*-oxide, were isolated from *Ceratocarpus heterocarpa*. ¹H NMR spectroscopy clearly distinguishes the *cis*- and *trans*-series of cularine *N*-oxides. Copyright © 1996 Elsevier Science Ltd

INTRODUCTION

The genus *Ceratocarpus* has attracted phytochemical interest because it contains alkaloids related to the metabolism of the 1-benzylisoquinoline, crassifoline [1]. *Ceratocarpus claviculata* (syn. *Corydalis claviculata*) [2, 3] and *C. palaestinus* [4] have been found to contain mainly cularine alkaloids, while the third species, *C. heterocarpa*, is interesting from a biosynthetic point of view on account of the presence of cularine [5] and 1,2-berbine [6] alkaloids. From our studies on *C. heterocarpa*, we have reported the isolation and structural elucidation of both *trans*- and *cis*-cularidine *N*-oxides (**1** and **2**) that exhibit a different conformation at the dihydroxepine ring and a distinct chemical behavior [7]. In the present paper, we report the isolation of two new alkaloids, (+)-*cis*-cularine *N*-oxide (**3**) and (+)-*cis*-sarcocapnine *N*-oxide (**4**).

RESULTS AND DISCUSSION

High-resolution mass spectrometry provided the molecular formula C₂₀H₂₃NO₅ for the optically active compounds (+)-**3** and (+)-**4**. The non-phenolic nature of these alkaloids, with three methoxyl groups and five quarternary aromatic carbons bonded to oxygen in the ¹³C NMR spectra, suggest a cularine-type structure. The aromatic protons in the ¹H NMR spectra revealed a 3', 4'-oxygenation pattern at ring D for compound **3**, while positions 4', 5'- were substituted in compound **4**; consequently, the compounds must be derivatives of cularine and sarcocapnine, respectively. The *N*-oxide function was inferred from the presence of a peak at *m/z* 341 [M – 16]⁺ in the EI-mass spectrum and the low-field chemical shift exhibited in the ¹³C NMR

spectra by carbon atoms bonded to nitrogen (Table 1). Moreover, the large β-substituent effect [8] induced by the N–O oxygen over C-1 and C-3 suggested a *cis*-relationship between H-1 and the *N*-oxide function, while the high-field for the *N*-Me group indicated an axial position. The ¹H NMR of **3** and **4** exhibited the characteristic ABX-system for the protons at C-1 and C-α of cularines. The *trans*-diaxial relationship between H-1 and H-αβ, inferred from the large coupling constant (*J*_{1-αβ} ≈ 12 Hz) suggested a dihydroxepine ring in a twist-boat conformation.

The chemical shift for H-1 varied very little (Δδ ≈ + 0.1 p.p.m.) from the free base (cularine and sarcocapnine) to the *cis*-*N*-oxides **3** and **4**; this can be ascribed to a conformational change at the dihydroxepine ring. The heterocyclic oxygen departs from H-1 and approaches H-αβ, which is shifted to low-field. Thus, the nitrogen configuration in cularine *N*-oxides can easily be established from the chemical shift for H-1 and H-αβ, δ ≈ 5 and δ ≈ 3, respectively, in the *trans*-series, and δ ≈ 4.5 and δ ≈ 3.4 in the *cis*-series. Thus, we

Table 1. Relevant ¹H and ¹³C NMR data for cularine *N*-oxides

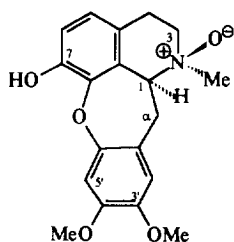
	<i>trans</i>	<i>cis</i>			
	1 ⁺	2 [*]	3 ⁺	4 ⁺	5 [‡]
C-1	69.6	73.8	72.8	73.1	
C-3	59.4	66.0	63.9	64.9	
C-α	32.3	28.8	30.0	28.8	
N-Me	55.3	49.1	51.3	49.0	
H-1	4.94	4.46	4.57	4.43	4.64
H-αα	4.40	4.20	4.02	4.10	3.93
H-αβ	2.99	3.36	3.35	3.40	3.52
N-Me	3.34	3.08	3.11	3.08	3.13

*Ref [7].

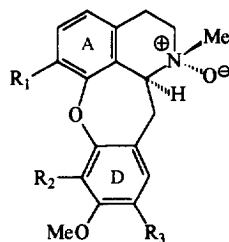
⁺See Experimental.

[‡]Ref. [9].

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- 2 R₁= OH, R₂=H, R₃=OMe
 3 R₁= OMe, R₂=H, R₃=OMe
 4 R₁= OMe, R₂=OMe, R₃=H
 5 R₁= OMe, R₂=OH, R₃=H

concluded that both **3** and **4** are the *cis*-*N*-oxides derived from (+)-cularine and (+)-sarcocapnine, respectively. In fact, (+)-**3** and (+)-**4** were obtained by oxidation of the corresponding free bases with *m*-CPBA. These results can be applied to (+)-sarcocapnidine *N*-oxide (**5**), isolated from *Sarcocapnos baetica* subsp. *integrifolia* [9], which was reported with no mention of its relative configuration. The ¹H NMR data obtained (Table 1) reveal that it belongs to the *cis*-series.

The parallelism between the aporphine and the cularine group of alkaloids has already been noted [3]. The greatest differences lie in the absence of naturally occurring 1,α-dehydrocularines; on the other hand, 6a,7-dehydroaporphines are of frequent occurrence [10]. Based on available knowledge, cularines can undergo quaternization at the nitrogen atom (either as *N*-oxide or as *N*-methyl cularinium salts), followed by β-elimination to the B-secocularine, as the preferential metabolic pathway. The chemical transformation of cularine *N*-oxides to the corresponding *N*-hydroxy-nor-secocularines has been reported [11] and proved to be particularly easy for the *trans*-*N*-oxides [7].

EXPERIMENTAL

General. Mps: uncorr. EIMS: direct inlet, 70 eV. FABMS: 2-hydroxyethyl disulphide as matrix. Silica gel 60 (70–230 mesh) was used for CC and silica GF₂₅₄ for TLC. ¹H and ¹³C NMR signals were measured at 200 and 50 MHz, respectively. Proton chemical shifts are referred to residual CHCl₃ (δ 7.24) and carbon chemical shifts to the solvent (¹³CDCl₃, δ 77). ¹H and ¹³C NMR signals were assigned from 2D COSY and DEPT expts.

Isolation. For a description of plant material and extraction conditions, see ref. [5]. The CHCl₃–MeOH-sol. part of the crude alkaloid extract was subjected to CC over silica gel. The fr. eluted with EtOAc–MeOH (1:4) was subsequently purified by CC and TLC to obtain the new compounds **3** (10 mg) and **4** (30 mg).

(+)-(1*S*,2*S*)-Cularine *N*-oxide (**3**). Amorphous powder. Mp 122–124°. [α]_D + 202° (MeOH; *c* 0.081). UV λ_{max} nm (log ε) MeOH: 230h (4.09), 284 (3.74). ¹H NMR (200 MHz, CDCl₃): δ 6.87 (2H, *s*, H-5, H-6),

6.80 (1H, *s*, H-2'), 6.79 (1H, *s*, H-5'), 4.57 (1H, *dd*, *J* = 3.2 and 12.0 Hz, H-1), 4.02 (1H, *dd*, *J* = 3.2 and 12.0 Hz, H-αα), 3.90 (3H, *s*, OMe), 3.84 (3H, *s*, OMe), 3.81 (3H, *s*, OMe), 3.35 (1H, *t*, *J* = 12.0 Hz, H-αβ), 3.9–3.6 (2H, *m*, H-3α, H-3β), 3.11 (3H, *s*, NMe), 3.2–2.9 (2H, *m*, H-4α, H-4β). ¹³C NMR (50 MHz, CDCl₃): δ 150.4, 150.3 (C-7, C-6'), 148.7, 146.7, 140.9 (C-8, C-3', C-4'), 124.0 (C-5), 123.9, 123.0 (C-8a, C-4a), 118.3 (C-1'), 112.9, 112.7 (C-2', C-6), 105.3 (C-5'), 72.8 (C-1), 63.9 (C-3), 56.4 (2 × OMe), 56.2 (OMe), 51.3 (NMe), 30.0 (C-α), 25.7 (C-4). EIMS *m/z* (rel. int.): 357 [M]⁺ (2), 341 [M – 16]⁺ (40), 326 [M – 16 – 15]⁺ (100), 298 [M – 59]⁺ (23). FABMS, *m/z*: 358 [M + H]⁺. HRMS *m/z*: 357.1581 ([M]⁺, calcd. for C₂₀H₂₃NO₅: 357.1576).

(–)-(1*S*,2*S*)-Sarcocapnine *N*-oxide (**4**). Amorphous powder. Mp 110–114°. [α]_D + 174° (MeOH; *c* 0.065). UV λ_{max} nm (log ε) MeOH: 230h (4.08), 284 (3.40). ¹H NMR (200 MHz, CDCl₃): δ 7.03 (1H, *d*, *J* = 8.5 Hz), 6.91 (1H, *d*, *J* = 8.5 Hz), 6.85 (1H, *d*, *J* = 8.5 Hz), 6.67 (1H, *d*, *J* = 8.5 Hz), 4.43 (1H, *dd*, *J* = 2.7 and 12.1 Hz, H-1), 4.10 (1H, *dd*, *J* = 2.7 and 12.1 Hz, H-αα), 3.91 (6H, *s*, 2 × OMe), 3.81 (3H, *s*, OMe), 3.40 (1H, *t*, *J* = 12.1 Hz, H-αβ), 3.9–3.7 (2H, *m*, H-3α, H-3β), 3.08 (3H, *s*, NMe), 3.2–2.9 (2H, *m*, H-4α, H-4β). ¹³C NMR (50 MHz, CDCl₃): δ 153.2 (C-7), 150.8, 150.7 (C-6', C-4'), 145.3, 141.1 (C-8, C-5'), 124.1, 123.9 (C-5, C-2'), 122.7, 122.4, 122.1 (C-4a, C-8a, C-1'), 113.6 (C-6), 109.4 (C-3'), 73.1 (C-1), 64.9 (C-3), 61.5, 56.5, 56.3 (3 × OMe), 49.0 (NMe), 28.8 (C-α), 25.9 (C-4). EIMS *m/z* (rel. int.): 357 [M]⁺ (6), 341 [M – 16]⁺ (74), 326 [M – 16 – 15]⁺ (53), 308 (45), 298 [M – 59]⁺ (100), 178 (48), 60 (44). FABMS, *m/z*: 358 [M + H]⁺. HRMS *m/z*: 357.1580 ([M]⁺, calcd. for C₂₀H₂₃NO₅: 357.1576).

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