

TRITERPENOID SAPONINS FROM *ANEMONE HUPEHENSIS*

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Key Word Index—*Anemone hupehensis*; Ranunculaceae; triterpenoid saponins; hupehensis saponins F,G.

Abstract—Two new triterpenoid saponins, named hupehensis saponin F and G, were isolated from the water soluble part of *Anemone hupehensis* Lemoine. By chemical and spectroscopic evidence, their structures were elucidated as 3-*O*- β -D-ribofuranosyl(1 $\rightarrow$ 3)- α -L-rhamnopyranosyl(1 $\rightarrow$ 2)- α -L-arabinopyranosyl hederagenin-28-*O*- α -L-rhamnopyranosyl(1 $\rightarrow$ 4)- β -D-glucopyranosyl(1 $\rightarrow$ 6)- β -D-glucopyranosyl(1 $\rightarrow$ 3)- α -L-rhamnopyranosyl(1 $\rightarrow$ 4)- β -glucopyranosyl(1 $\rightarrow$ 6)- β -D-glucopyranoside and 3-*O*- β -D-ribofuranosyl(1 $\rightarrow$ 3)- α -L-rhamnopyranosyl(1 $\rightarrow$ 2)- α -L-arabinopyranosyl hederagenin-28-*O*- β -glucopyranosyl(1 $\rightarrow$ 3)- α -L-rhamnopyranosyl(1 $\rightarrow$ 4)- β -D-glucopyranosyl(1 $\rightarrow$ 6)- β -D-glucopyranosyl(1 $\rightarrow$ 3)- α -L-rhamnopyranosyl(1 $\rightarrow$ 4)- β -D-glucopyranosyl(1 $\rightarrow$ 6)- β -D-glucopyranoside, respectively. Copyright © 1996 Published by Elsevier Science Ltd

INTRODUCTION

In Chinese folk medicine, *Anemone hupehensis* Lemoine, which is widely spread in China, is used for curing dysentery, infantile malnutrition, carbuncles, fungulosis, sores and scrofula [1]. There was also a report about anemone having insecticidal properties. Previously we have reported the triterpenoid saponins with less than seven sugar units isolated from this plant [2, 3]. In this paper, we report two new saponins, hupehensis saponin F and G, which contain more than nine sugar units.

RESULTS AND DISCUSSION

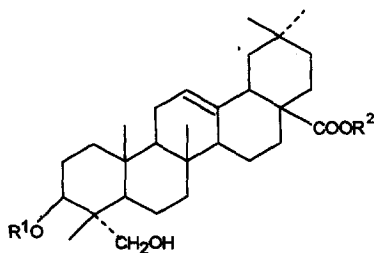
The crude saponin fractions were subjected to repeated CC on silica gel, affording saponins 1 and 2. The yields of the two saponins were 0.006 and 0.002% of the dry plant, respectively.

On acid hydrolysis, saponins 1 and 2 yielded hederagenin identified by comparison with an authentic sample. A comparison of the ^{13}C NMR signals due to the aglycone moieties with those of reported saponins revealed that they were all bisdesmosides of 3,28-di-*O*-glycosides [4].

Saponin 1 was hydrolysed with acid to yield D-glucose, L-rhamnose, L-arabinose and D-ribose as sugar components. The EI-mass spectrum of its acetate showed fragment ions at m/z 259 [terminal ribose

(Ac) $_3$] $^+$, 489 [ribose (Ac) $_3$, rhamnose (Ac) $_2$], 273 [terminal rhamnose (Ac) $_3$] $^+$, 561 [rhamnose (Ac) $_3$, glucose (Ac) $_3$] $^+$. The ^{13}C NMR spectrum of sugar moieties (Table 1) indicated the presence of nine monosaccharide units. On 6 M NH_4OH hydrolysis, prosapogenin 1a was obtained. By comparison with the authentic sample, prosapogenin 1a was identified as prosapogenin CP $_6$, which was previously isolated from the same plant. L-Rhamnose and D-glucose were detected by complete hydrolysis of the solution from which prosapogenin CP $_6$ was removed. The ^{13}C NMR signals due to a 28-*O*-saccharide were deciphered from the signals due to saponin 1 after removal of the signals due to prosapogenin CP $_6$. These signals indicated that there were six monosaccharide units. On comparison with the ^{13}C NMR spectrum of huzhangoside D, there were two groups of ^{13}C NMR signals the same as those due to the 28-*O*-saccharide of huzhangoside D [5]. That means that the 28-*O*-saccharide of saponin 1 contained two units of α -L-rhamnopyranosyl(1 $\rightarrow$ 4)- β -glucopyranosyl(1 $\rightarrow$ 6)- β -D-glucopyranoside. According to the glycosylation shift (δ 83.6) reported in the literature [6], this indicated that the α -L-rhamnopyranose was 3-*O*-glycosylated. Based on the above evidence, the structure of saponin 1 was established as 3-*O*- β -D-ribofuranosyl(1 $\rightarrow$ 3)- α -L-rhamnopyranosyl(1 $\rightarrow$ 2)- α -L-arabinopyranosyl hederagenin-28-*O*- α -L-rhamnopyranosyl(1 $\rightarrow$ 4)- β -D-glucopyranosyl(1 $\rightarrow$ 6)- β -D-glucopyranosyl(1 $\rightarrow$ 3)- α -L-rhamnopyranosyl(1 $\rightarrow$ 4)- β -glucopyranosyl(1 $\rightarrow$ 6)- β -D-glucopyranoside, which has been named as hupehensis saponin F.

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Saponin 1: $R^1 = \text{rib}(1 \rightarrow 3)\text{rha}(1 \rightarrow 2)\text{ara}$

$R^2 = \text{rha}(1 \rightarrow 4)\text{glc}(1 \rightarrow 6)\text{glc}(1 \rightarrow 3)\text{rha}(1 \rightarrow 4)\text{glc}(1 \rightarrow 6)\text{glc}$

Saponin 2: $R^1 = \text{rib}(1 \rightarrow 3)\text{rha}(1 \rightarrow 2)\text{ara}$

$R^2 = \text{glc}(1 \rightarrow 6)\text{rha}(1 \rightarrow 4)\text{glc}(1 \rightarrow 6)\text{glc}(1 \rightarrow 3)\text{rha}(1 \rightarrow 4)\text{glc}(1 \rightarrow 6)\text{glc}$

Saponin 2 was hydrolysed with acid to yield D-glucose, D-ribose, L-rhamnose and L-arabinose as sugar components. The EI-mass spectrum of its acetate showed fragment ions at m/z 331 [terminal glucose (Ac)₄]⁺, 561 [glucose (Ac)₄ rhamnose (Ac)₂]⁺, 259 [terminal ribose (Ac)₃]⁺ and 489 [ribose (Ac)₃ rhamnose (Ac)₂]⁺. The ¹³C NMR spectrum due to the sugar moieties indicated the presence of 10 monosaccharide units. On 6 M NH₄OH hydrolysis, the obtained prosapogenin 2a was identified as prosapogenin CP₆ by comparison with an authentic sample. It gave L-rhamnose and D-glucose by complete hydrolysis of the solution from which prosapogenin CP₆ was removed. The ¹³C NMR signals due to a 28-O-saccharide were obtained from the signals due to saponin 1 minus the signals due to prosapogenin CP₆. The signals obtained indicated that there were seven monosaccharide units. On comparison with the ¹³C NMR of saponin 1, saponin 2 contained one more β-D-glucopyranose than saponin 1. The glycosylation shift (δ 83.7) indicated that an α-L-rhamnopyranose was 3-O-glycosylated. Therefore, the structure of saponin 2 was elucidated as 3-O-β-D-ribofuranosyl(1→3)-α-L-rhamnopyranosyl(1→2)-α-L-arabinopyranosyl hederagenin-28-O-β-D-glucopyranosyl(1→3)-α-rhamnopyranosyl(1→4)-β-D-glucopyranosyl(1→6)-β-D-glucopyranoside(1→3)-α-L-rhamnopyranosyl(1→4)-β-D-glucopyranosyl(1→6)-β-D-glucopyranoside, which has been named hupehensis saponin G.

EXPERIMENTAL

¹H and ¹³C NMR spectra were recorded at 400 MHz in pyridine-*d*₅ using TMS as int. standard. EI-MS were measured at 40 eV accelerating voltage after acetylation. Optical rotations were measured with a J-20C automatic recording spectropolarimeter at 13° in MeOH.

Plant material. Roots and leaves of *Anemone hupehensis* Lemoine were collected in Kai-xan of Sichuan Province, China, and identified by Prof. S. C.

Xiao. A specimen is deposited in the Herbarium of the Chengdu Institute of Biology.

Extractions and isolation of saponins. Dry roots and leaves (3.5 kg) were extracted with 95% EtOH. After removal of solvent by evapn, the combined extracts were suspended in H₂O, extracted with EtOAc several times and then with *n*-BuOH. The residue (210 g) was subjected to sepn on a D101 resin column. After diluting with large amounts of H₂O, and 15% EtOH-H₂O to remove most of the coloured material, EtOH was then used to elute crude saponins (3 g). The crude saponin fr. was chromatographed on a silica gel column, eluting with CHCl₃-MeOH-H₂O to yield frs 1-6. Frs 1-4 had been isolated before. Frs 5 and 6 were purified by prep. TLC to give saponins 1 (218 mg) and 2 (90 mg) as powders. Compound 1, C₈₂H₁₃₄O₄₄, [α]_D¹³ 18.7° (MeOH; *c* 2.53). EI-MS m/z : 259 [terminal ribose (Ac)₃]⁺, 489 [ribose (Ac)₃ rhamnose (Ac)₂]⁺, 273 [terminal rhamnose (Ac)₃]⁺, 561 [rhamnose (Ac)₃ glucose (Ac)₃]⁺. Compound 1a, FAB-MS m/z 906 {[M]⁺ (C₄₆H₇₄O₁₆) + Na + H}. Compound 1a was identified as prosapogenin CP₆ by comparison with an authentic sample. Compound 2, C₈₈H₁₄₄O₄₉, [α]_D¹³ +8.0° (pyridine, *c* 3.1). EI-MS m/z : 331 [terminal glucose (Ac)₄]⁺. Compound 2a was also identified as prosapogenin CP₆.

Acid hydrolysis of the saponins and identification of the resulting monosaccharide. Each saponin (10 mg) was heated with 5% H₂SO₄ under reflux for 7 hr. The reaction mixture was diluted with H₂O and extracted with CHCl₃. Sapogenin was detected in the CHCl₃ layer by TLC. The aq. layer was neutralized with Ba(OH)₂ and concentrated, then subjected to PC analysis with authentic samples, developing solvent BuOH-HOAc-H₂O (4:1:5) (upper layer), detection reagent: aniline-phthalate.

Alkaline saponification of saponins. A soln of saponin (20 mg) in 6-M NH₄OH was heated at 100° for 3 hr. The reaction mixt. was extracted with *n*-BuOH to give prosapogenin, which was identified by comparison with the authentic sample on TLC. The aq. layer was

Table 1. ^{13}C NMR spectral data of the saponin sugar moieties of compounds **1** and **2** (in pyridine- d_5 , ppm)

	1	2		1	2		1	2
R1-ara-1	104.5	104.4	R2-glc-1	95.4	95.3	glc-1	104.3	104.7
2	75.1	74.9	2	73.6	73.5	2	75.8	75.7
3	73.6	73.5	3	77.7	77.5	3	76.1	76.0
4	68.6	68.5	4	70.5	70.4	4	78.1	78.0
5	65.8	65.6	5	76.8	76.7	5	76.8	76.7
rha-1	101.1	101.0	6	69.0	69.9	6	61.1	61.1
2	72.2	72.1	glc-1	104.3	104.4	rha-1	101.7	101.6
3	80.9	80.7	2	75.1	74.9	2	72.2	72.1
4	72.4	72.2	3	76.2	76.1	3	71.7	83.7
5	69.6	69.6	4	78.4	78.4	4	74.7	74.4
6	18.2	18.2	5	78.0	78.0	5	69.8	69.6
rib-1	104.3	104.2	6	61.1	61.0	6	18.0	18.0
2	72.4	72.2	rha-1	102.3	102.2	glc-1		104.7
3	70.0	69.9	2	71.8	71.7	2		76.0
4	68.2	68.1	3	84.0	83.6	3		78.0
5	64.9	64.8	4	72.5	72.4	4		70.9
			5	69.8	69.6	5		77.4
			6	18.2	18.2	6		61.0
			glc-1	105.7	105.5			
			2	75.1	74.9			
			3	77.4	77.4			
			4	71.0	70.9			
			5	76.8	76.7			
			6	69.3	69.2			

hydrolysed with 5% H_2SO_4 , after being neutralized with BaCO_3 and concd, then subjected to PC analysis to detect the sugar units by comparison with authentic samples.

Acetylation of sapogenin. To each sapogenin (5 mg) was added Ac_2O -pyridine (1:1) (0.5 ml) in a microtube. After standing at room temp. for 48 hr the soln was evapd to dryness and subjected to EI-MS analysis.

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REFERENCES

1. Jiangsu New Medical College (1979) *The Dictionary of Trading Chinese Medicines*.
2. Wang, M. K., Wu, F.-E. and Chen, Y.-Z. (1993) *Phytochemistry* **34**, 1395.
3. Wang, M. K., Wu, F.-E. and Chen, Y.-Z. (1994) *Acta Chim. Sinica* **52**, 609.
4. Kimata, H., Nakashima, T., Kokubun, S., Nakayama, K., Mitoma, Y., Kitahara, T., Yata, N. and Tanaka, O. (1983) *Chem. Pharm. Bull.* **31**, 1988.
5. Mizutani, K., Ohtani, K., Wei, J. X., Kasai, R. and Tanaka, O. (1984) *Planta Med.* **51**, 327.
6. Ahmad, V. U., Perveen, S. and Ranoshaheen (1989) *Planta Med.* **55**, 307.