

## PII: S0031-9422(96)00682-6

# HYDROHALIMIC ACIDS FROM HALIMIUM VISCOSUM

DINA I. M. D. DE MENDONÇA, JESUS M. L. RODILLA,\* ANNA M. LITHGOW† and ISIDRO S. MARCOS‡

Departamento de Química, Universidade da Beira Interior, 6200 Covilha, Portugal; †Servicio General de Resonancia Magnética Nuclear, Fac. de Ciencias Químicas, Universidad de Salamanca, 37008 Salamanca, Spain; †Departmento de Química Orgánica, Universidad de Salamanca, 37008 Salamanca, Spain

(Received in revised form 29 August 1996)

**Key Word Index**—*Halimium viscosum*; Cistaceae; diterpenoid acids; rearranged *ent*-labdanes; *ent*-halimanes.

**Abstract**—Several diterpenoid acids, with an *ent*-halimane skeleton, were isolated from the aerial parts of *Halimium viscosum* as methyl esters. Besides the well known hydrohalimic acid, acetoxyhydrohalimic acid, cinnamoyloxyhydrohalimic acid and 2-hydroxyhydrohalimic acid, three new ones have been identified as methyl 15-*Z*-cinnamoyloxy-1(10)-*ent*-halimen-18-oate, methyl 15-hydroxy-2-oxo-1(10)-*ent*-halimen-18-oate and methyl 15-methoxy-1(10)-*ent*-halimen-18-oate. Their structures have been determined by chemical and spectroscopic methods, using 2D correlation experiments (¹H−¹³C and ¹³C−¹³C). Copyright © 1997 Elsevier Science Ltd

#### INTRODUCTION

A variety of diterpenoid structures and skeleta, many of them described for the first time, have been isolated during the last years from *Halimium viscosum* chemotypes in the Iberian peninsula and this has encouraged the search for another chemotype.

Villarino's H. viscosum contained only ent-halimane acids with a  $\Delta^{1(10)}$  double bond, a carboxyl at C-4 as in the case of the major component halimic acid (1) [1, 2], and with polyfunctionalization [3] of the side chain or even its degradation [4]. Fregeneda's chemotype, however, contained only ent-halimane acids with a saturated side chain, e.g. hydrohalimic acid (2) [5, 6]. The neutral fraction of Fregeneda's H. viscosum contained a much greater variety of diterpenoid skeleta than the neutral part from Villarino's H. viscosum [7, 8]; in fact, 3-oxygenated labdanes [9, 10] such as 7,13*E*-labdadien-3 $\beta$ ,15-diol (3), the major component, were isolated from it as well as other labdane alcohols. Moreover, four different carbon backbones were isolated from the extract: tormesane as in tormesol (4) [11, 12], tormesolane as in tormesolanone (5) [13], fregenedane as in fregenedadiol (6) [14-16] and isofregenedane as in isofregenedadiol (7) [17, 18]. Valparaíso's chemotype contained only labdane acids with a carboxyl group at C-17 such as zamoranic acid (8) [19, 20], with either a saturated or unsaturated side chain [20] and the neutral part afforded labdanes

In this work we have studied the composition of the acid part of *H. viscosum* (Celorico da Beira, Portugal) and isolated several diterpenoid acids, with a saturated side chain belonging to the *ent*-halimane class. Some of them are new and, for the well known acids, the spectroscopic assignment is completed with the <sup>13</sup>C NMR data.

### RESULTS AND DISCUSSION

A hexane extract of H. viscosum (collected at Celorico da Beira, Portugal) was dewaxed with methanol and separated into neutral and acid parts. A small aliquot of the acid part was subjected to column chromatography on silica gel, and an acetoxy acid (12) (IR 2936, 1739, 1691 and 1241 cm<sup>-1</sup>) and a hydroxy acid (13) (IR 3360 and 1691 cm<sup>-1</sup>) were isolated. The <sup>1</sup>H NMR spectra of 1 and 2 were very similar: CH<sub>2</sub>-CH=C ( $\delta$  5.27, br s), four methyl groups, two of them doublets ( $\delta$  0.84 and 0.75; J = 5.0 and 7.0 Hz) and two singlets ( $\delta$  1.08 and 0.83), one of the latter ( $\delta$ 1.08, s) being geminal to a carboxyl group. The main difference was observed in the CH<sub>2</sub>-CH<sub>2</sub>-OAc group in 12 ( $\delta$  4.08, m) and the CH<sub>2</sub>-CH<sub>2</sub>-OH group in 13 ( $\delta$  ppm, 3.60, m). The <sup>13</sup>C NMR data for both compounds, except for the acetoxyl group signal in 12, showed the same multiplicity for the 20 carbon atoms: i.e. four methyls, eight methylenes, one of them

without an oxygenated function on ring A (e.g. 9) [21] and tricyclic diterpenoid alcohols with valparane and valparolane skeleta such as valparene (10) [7, 22–24] and valparolone (11) [25], respectively.

<sup>\*</sup>Author to whom correspondence should be addressed.

directly bonded to an oxygen function ( $\delta$  63.2 and 60.9), four methines, one of them sp<sup>2</sup> hybridized ( $\delta$  119.5 and 119.3), and four fully substituted carbon atoms, one olefinic and one carboxyl.

10

When 12 and 13 were esterified with diazomethane, 14 and 15 were obtained, respectively, for which the spectroscopic data (Table 1) indicated the same similarities between 14 and 15 as for 12 and 13. When 15 was acetylated 14 was obtained.

Two-dimensional heteronuclear experiments, XHCORR and COLOC, and a 2D homonuclear experiment, INADEQUATE, on 15 (Table 2) and

comparison with an authentic sample ( $\alpha_d + 67.6$  specific rotation) allowed the identification of **13** as hydrohalimic acid and **12** as acetoxyhydrohalimic acid [5, 6]. Both compounds were described by us several years ago as methyl esters, isolated from *H. viscosum* (La Fregeneda) without <sup>13</sup>C NMR data [5].

11

The remainder of the acid part was esterifed with diazomethane and carefully chromatographed (column chromatography, silica gel<sub>2</sub>) affording seven methyl esters (14–20). (The numbering is not in order of elution). Methyl esters 16–19 showed in their <sup>1</sup>H NMR spectra the same characteristics of the bian-

Table 1. <sup>1</sup>H NMR data for compounds 14–22 (250 MHz, CDCl<sub>1</sub>)

			Table	l. 'H NMR data	for compounds 14	Table 1. 'H NMR data for compounds 14-22 (250 MHz, CDCI;)			
Н	72	15	16	17	18	19	20	21	22
_	5.27, 1H, <i>t</i> (4.3)	5.16, 1H, <i>t</i> (4.3)	5.30, 1H, t (4.3)	5.31, 1H, m	5.32, 1H, br s	5.79, 1H, s	5.22, 1H, br s	5.79, IH, s	5.87, 1H, <i>d</i> (5.9)
2							5.41, 1H, dt (4.4, 1.9)		4.69, 1H, <i>t</i> (4.9)
3						2.66, 1H, d		2.67, 1H, d	
						(15.9) 2.24, 1H, d		(16.0) 2.25. 1H. d	
						(15.9)		(16.0)	
S	2.63, 1H, dd	2.53, 1H, dd	2.65, 1H, dd	2.66, 1H, m	2.67, 1H, m	2.97, 1H, dd	2.71, 1H, dd	3.00, 1H, dd	2.20, 1H, dd
	(12.0, 3.8)		(12.0, 3.4)			(12.8, 4.9)	(12.2, 3.6)	(12.5, 4.6)	(12.7, 4.9)
15	4.05, 2H, m		3.39, 2H, t	4.14, 2H, m	4.23, 2H, m	3.63, 2H, m	4.07, 2H, m	4.06, 2H, m	4.04, 2H, dt
,			(6.8)						(6.8, 2.0)
16	0.87, 3H, d		0.89, 3H, d	0.88, 3H, d	0.95, 3H, d	0.88, 3 <b>H</b> , <i>d</i>	0.90, 3H, d	0.90, 3H, d	0.84, 3H, d
	(6.3)		(7.1)	(6.5)	(6.2)	(6.1)	(6.3)	(6.3)	(5.9)
17	0.75, 3H, d	0.73, 3H, d	0.78, 3H, d	0.79, 3H, d	0.79, 3H, d	0.78, 3H, d	0.81, 3H, d	0.78, 3H, d	0.77, 3H, d
	(7.0)		(6.9)	(7.0)	(7.0)	(7.3)	(7.0)	(7.1)	(7.8)
19	1.07, 3H, s		1.10, 3H, s	1.11, 3H, s	1.10, 3H, s	1.20, 3H, s	1.13, 3H, s	1.21, 3H, s	1.21, 3H, s
20	0.84, 3H, s		0.87, 3H, s	0.87, 3H, s	0.88, 3H, s	0.95, 3H, s	0.87, 3H, s	0.95, 3H, s	0.88, 3H, s
$CO_2CH_3$	3.61, 3H, s		3.64, 3H, s	3.64, 3H, s	3.64, 3H, s	3.60, 3H, s	3.64, 3H, s	3.61, 3H, s	
2,				5.95, 1H, d	6.43, 1H, d				
				(12.6)	(16.0)				
3,				6.94, 1H, d	7.67, 1H, d				
				(12.6)	(16.0)				
5′,9′				7.57, 2H, m	7.51, 2H, m				
,8′,2′,9′				7.33, 3H, m	7.37, 3H, m				
15-0 <sub>2</sub> CCH <sub>3</sub>	2.00, 3H, s						2.02, 3H, s	2.02, 3H, s	2.02, 3H, s
$2-0$ 2CC $\overline{H}_3$			116 66 6				2.04, 3H, s		
OH OH		3.03, 1H, br s	5.55, 5H, S						
	:								

Coupling constant (J in Hz) are given in parentheses.

Position	C	Н	COLOC	INADEQUATE
1	119.0	5.16	2, 5, 9	2, 10
2	22.4	1.90		1
3	30.3	1.30-1.65	2, 18	4
4	44.5			3, 5, 18, 19
5	38.0	2.53	4, 10	4, 6, 10
6	22.6	1.10		5, 7
7	28.1	1.97	6, 17	6
8	38.1	1.40		7, 9, 17
9	42.4			8, 10, 11, 20
10	141.2			1, 5, 9
11	35.9	1.15-1.75	9, 12, 20	9, 12
12	30.6	0.92 - 1.00		11
13	29.7	1.30-1.60		14, 16
14	39.3	1.20-1.42		13, 15
15	60.5	3.47		14
16	19.5	0.83	12, 14	13
17	15.3	0.73	7, 9	8
18	178.4			4
19	19.7	1.00	3, 4, 5	4
20	22.2	0.81	8, 9, 10, 11	9
COOMe	51.4	3.50	18	

Table 2. NMR data for compound 15 and correlations observed in 2D experiment

nular system described for **14** and **15**. The main difference resides in the function at C-15 (Table 3).

Compound **16** has a methoxyl group bonded to the methylene at C-15 ( $\delta$  3,39, t,J = 6.8 Hz, 71.23), **17** has a Z-cinnamoyloxy group –O–CO–CH—CH–C<sub>6</sub>H<sub>5</sub> ( $\delta$  5.95, 6.94, d, J = 12.6 Hz, Table 1) while **18** has a Z-cinnamoyloxy group –O–CO–CH—CH–C<sub>6</sub>H<sub>5</sub> ( $\delta$  6.43, 7.67, d, J = 16.0 Hz).

A small amount of 19 was separated from the most polar fraction of the Me esters together with a mixture from which 20 and 21 were isolated. Compound 20 is a methyl ester with similar characteristics to the previously isolated compounds with two acetoxyl groups:  $CH_2$ - $CH_2$ -OAc and AcO-CH-CH-CH ( $\delta$  4.07, m, and 5.41, dt,  $J_1$  = 4.4 Hz and  $J_2$  = 1.9 Hz). This secondary acetoxyl group can be positioned at C-2 in an *ent*-halimane skeleton. The multiplicity of the H-2 (dt) indicated an  $\alpha$ -stereochemistry for the allylic group, which is confirmed later (see below).

Compound 19 has a –CO–CH—C– group with the carbonyl at C-2 conjugated with the annular double bond ( $\delta$  5.79, s). Acetylation of 19 gave 21 (UV  $\lambda_{max} = 241.0$  nm). The 2-oxo structure for 19 was confirmed by treatment of compound 14 with Na<sub>2</sub>CrO<sub>4</sub> to give 21 in good yield. Reduction of 21 with NaBH<sub>4</sub> gave 21, and a mixture which after acetylation afforded 20. However, reduction of 21 with aluminium isopropoxide afforded lactone 22 (16 mg).

### **EXPERIMENTAL**

Spectral analysis. NMR: 250 or 400 MHz for  $^{1}$ H and 62.9 or 100.1 MHz for  $^{13}$ C. Chemical shifts are given in  $\delta$  (ppm) and are referenced to the residual

CHCl<sub>3</sub>, 7.26 ppm for the <sup>1</sup>H and 77.0 ppm for <sup>13</sup>C, respectively. GC-MS: VG Trio 1000, 70 eV.

Extraction and isolation. Aerial parts of H. viscosum (131 g) collected in Celorico da Beira (Guarda, Portugal) were dried and extracted with n-hexane in a Soxhlet apparatus for 24 hr. The extract (10 g) was dewaxed with MeOH (0.8 g) and then extracted with 4% NaOH (6.4 g). The neutral fr. weighed 2.8 g. An aliquot (500 mg) of the acid part was subjected to CC on silica gel, affording 12 (10 mg) and 13 (200 mg). The remaining acid part (5.9 g) was esterified with  $CH_2N_2$  and then the Me esters were subjected to CC on silica gel with n-hexane–EtOAc mixts giving three frs (1–III).

CC of fr. I on silica gel/10% AgNO<sub>3</sub> gave **16** (18 mg), **18** (9 mg) and **14** (25 mg). CC of fr. II on silica gel gave **15** (2.05 g). CC of fr. III on silica gel gave **19** (17 mg). Acetylation of a part of fr. III followed by CC on silica gel afforded **20** (18 mg) and **21** (39 mg).

Acetoxyhydrohalimic acid (12). Oil. IR  $v_{\rm max}^{\rm film}$  cm<sup>-1</sup>: 2936, 2869, 1739, 1691, 1463, 1375, 1241; <sup>1</sup>H NMR: δ 5.32 (1H, dt, J = 5.0, 2.5 Hz, H-1), 4.08 (2H, m, H-15), 2.65 (1H, dd, J = 12.5, 2.5 Hz, H-5), 2.04 (3H, s, O<sub>2</sub>CCH<sub>3</sub>), 1.14 (3H, s, Me-19), 0.89 (3H, d, J = 4.8 Hz, Me-16), 0.88 (3H, s, Me-20), 0.80 (3H, d, J = 7.0 Hz, Me-17); <sup>13</sup>C NMR: Table 3.

*Hydrohalimic acid* (13). Oil. IR  $v_{\text{max}}^{\text{film}}$  cm<sup>-1</sup>: 3360, 2910, 1691, 1456, 1369, 1281; <sup>1</sup>H NMR: δ 6.93 (1H, br, CO<sub>2</sub>H), 5.27 (1H, br, H-1), 3.60 (2H, m, H-15), 2.61 (1H, d, J = 9.8 Hz, H-5), 1.08 (3H, s, Me-19), 0.84 (3H, d, d = 5.0 Hz, Me-16) 0.83 (3H, d Me-20), 0.75 (3H, d, d = 6.7 Hz, Me-17); <sup>13</sup>C NMR: Table 3.

Methyl 15-acetoxy-1(10)-ent-halimen-18-oate (14). Oil. IR  $v_{max}^{film}$  cm<sup>-1</sup>: 3057, 1736, 1461, 1378, 1243, 1166;

Table 3. <sup>13</sup>C NMR data of compounds 12-22 (62.9 MHz, CDCl<sub>3</sub>)

C	12	13	14	15	16	17	18	19	20	21	22
1	119.5	119.3	119.3	119.0	119.2	119.4	119.4	124.8	119.9	124.8	122.8
2	22,7	22.7	22.7	22.4	22.7	22.8	22.7	197.2	69.1	197.1	72.2
3	30.2	30.1	30.6	30.3	30.8	30.8	30.8	43.0	33.5	43.1	37.4
4	44.6	44.6	44.8	44.5	44.8	44.9	44.9	46.2	44.7	46.2	44.7
5	38.1	38.0	38.2	38.0	38.2	38.3	38.3	40.5	38.2	40.5	41.2
6	23.0	23.0	22.8	22.6	22.8	22.9	22.9	23.4	23.7	23.4	22.3
7	28.4	28.4	28.3	28.1	28.3	28.4	28.4	28.2	28.8	28.2	28.1
8	38.6	38.5	38.3	38.1	38.3	38.4	38.4	41.0	39.4	41.0	38.8
9	42.9	42.7	42.7	42.4	42.7	42.8	42.8	45.2	43.4	45.2	42.6
10	141.4	147.5	141.4	141.2	141.5	141.5	141.5	169.9	147.0	169.7	149.6
11	35.4	35.8	35.4	36.0	36.6	35.4	35.6	36.4	35.3	35.2	35.3
12	30.8	30.5	30.8	30.6	31.0	30.9	30.9	30.9	30.8	31.0	30.7
13	30.5	29.8	30.5	29.8	30.4	30.5	30.6	30.2	30.6	30.5	30.2
14	36.6	39.1	36.5	39.3	36.6	36.5	36.6	39.4	36.2	36.6	35.5
15	63.2	60.9	63.1	60.5	71.2	63.1	63.3	60.9	63.1	62.9	63.0
16	19.6	19.7	19.6	19.5	19.9	19.6	19.8	19.6	19.6	19.5	19.1
17	15.6	15.5	15.5	15.3	15.6	15.6	15.6	15.5	15.5	15.5	15.3
18	184.7	184.1	178.5	178.4	178.6	178.6	178.6	176.8	177.3	176.7	181.9
19	20.2	20.3	19.9	19.7	19.9	19.9	19.9	21.2	21.9	21.1	19.5
20	22.4	22.3	22.5	22.2	22.6	22.6	22.6	21.9	23.6	21.9	21.8
COOCH <sub>3</sub>			51.6	51.4	51.7	51.7	51.7	52.4	52.0	52.4	
1'						166.3	167.1				
2'						142.9	144.5				
3'						120.0	118.3				
4'						135.0	134.5				
5'						128.0	128.0				
6'						129.6	128.8				
7′						128.9	130.2				
8'						129.6	128.8				
9′						128.0	128.0				
OCH <sub>3</sub>					58.6	120.0	120.0				
15-OOCMe	171.3		171.1		20.0				171,2	171.1	171.2
15-00CM			21.0						21.0	21.0	21.0
2-OOCMe	3 41.0		41.0						170.8	21.0	21.0
2-00CMe 2-00CCH <sub>3</sub>									21.5		
2-00CCR3									21.3		

<sup>1</sup>H and <sup>13</sup>C NMR: Tables 1 and 3; GC-MS 70 eV, *m/z* (rel. int.): 378 [M]<sup>+</sup> (1), 236 (16), 235 (97), 176 (26), 175 (100), 147 (12), 133 (15), 119 (28), 107 (13), 105 (38), 95 (12), 93 (16), 91 (20), 81 (15), 69 (17), 55 (43).

Methyl 15-hydroxy-1(10)-ent-halimen-18-oate (15). Oil;  $[\alpha]_D^{22} = +67.65$  (CHCl<sub>3</sub>, c 2.365);  $1R v_{max}^{film}$  cm<sup>-1</sup>: 3364, 3044, 1731, 1380, 1164, 1054;  $^1H$  and  $^{13}C$  NMR: Tables 1–3; GC-MS 70 eV, m/z (rel, int.): 336 [M]<sup>+</sup> (<1), 236 (11), 235 (75), 175 (100), 133 (10), 119 (19), 105 (25), 93 (10), 91 (16), 69 (19), 55 (52).

Methyl 15-methoxy-1(10)-ent-halimen-18-oate (**16**). Oil. IR  $\nu_{\text{max}}^{\text{film}}$  cm<sup>-1</sup>: 2930, 1725, 1462, 1375, 1248, 1187; <sup>1</sup>H and <sup>1</sup>C NMR: Tables 1 and 3; GC-MS 70 eV, m/z (rel. int.): 350 [M]<sup>+</sup> ( $\ll$ 1), 236 (15), 235 (100), 176 (16), 175 (95), 119 (14), 105 (16).

Methyl 15-Z-cinnamoyloxy-1(10)-ent-halimen-18-oate (17). Oil. IR  $v_{\text{max}}^{\text{film}}$  cm<sup>-1</sup>: 2922, 1731, 1712, 1631, 1470, 1389, 1254, 1153, 830, 763, 689; <sup>1</sup>H and <sup>13</sup>C NMR: Tables 1 and 3; GC-MS 70 eV, m/z (rel. int.): 466 [M]<sup>+</sup> (2), 263 (17), 235 (100), 207 (18), 175 (61), 147 (14), 131 (40), 103 (17).

Methyl 15-cinnamoyloxy-1(10)-ent-halimen-18-oate

(18). Oil. IR  $v_{\text{max}}^{\text{film}}$  cm<sup>-1</sup>: 3064, 1726, 1705, 1664, 1380, 1166, 981, 765, 709; <sup>1</sup>H and <sup>13</sup>C NMR: Tables 1 and 3; GC-MS 70 eV, m/z (rel. int.): 466 [M]<sup>+</sup> (2), 236 (16), 235 (100), 207 (28), 175 (51), 131 (43), 103 (19), 77 (10).

Methyl 15-hydroxy-2-oxo-1(10)-ent-halimen-18-oate (19). Oil. IR  $v_{\text{max}}^{\text{film}}$  cm<sup>-1</sup>: 3384, 3064, 1728, 1664, 1611, 1380, 1157, 823; <sup>1</sup>H and <sup>13</sup>C NMR: Tables 1 and 3; GC-Ms 70 eV, m/z (rel. int.): 350 [M]<sup>+</sup> (5), 189 (26), 161 (17), 147 (11), 122 (10), 121 (100), 55 (17).

Methyl 2,15-diacetoxy-1(10)-ent-halimen-18-oate (20). Oil. IR  $v_{\rm max}^{\rm film}$  cm<sup>-1</sup>: 3060, 1757, 1717, 1650, 1380, 1245, 1156; <sup>1</sup>H and <sup>13</sup>C NMR: Tables 1 and 3; GC-Ms 70 eV, m/z (rel. int.): 436 [M]<sup>+</sup> ( $\ll$ 1), 174 (15), 173 (100), 159 (15), 145 (35), 131 (31), 119 (19), 105 (62), 55 (21).

Methyl 15-acetoxy-2-oxo-1(10)-ent-halimen-18-oate (21). Oil. IR  $\nu_{\text{max}}^{\text{film}}$  cm<sup>-1</sup>: 3057, 1734, 1675, 1610, 1380, 1240, 1160; UV  $\lambda_{\text{max}} = 241.0$  nm (ε = 13 000); <sup>1</sup>H and <sup>13</sup>C NMR: Tables 1 and 3; GC-MS 70 eV, m/z (rel. int.): 392 [M]<sup>+</sup> (4), 189 (37), 175 (10), 161 (22), 147 (12), 135 (10), 121 (100), 105 (10), 91 (10), 55 (19).

$$R_{1} \qquad R_{2} \qquad R_{3}$$

$$R_{1} \qquad R_{2} \qquad R_{3}$$

$$R_{2} \qquad H \qquad H \qquad OAc$$

$$R_{3} \qquad H \qquad H \qquad OH$$

$$R_{4} \qquad Me \qquad H \qquad OAc$$

$$R_{5} \qquad Me \qquad H \qquad OH$$

$$R_{6} \qquad H \qquad OH$$

$$R_{7} \qquad Me \qquad H \qquad OCOCH = CHC_{6}H_{5}$$

$$R_{8} \qquad Me \qquad H \qquad OCOCH = CHC_{6}H_{5}$$

$$R_{9} \qquad Me \qquad OAc \qquad OAc$$

$$R_{1} \qquad Me \qquad H \qquad OCOCH = CHC_{6}H_{5}$$

$$R_{2} \qquad Me \qquad OAc \qquad OAc$$

$$R_{3} \qquad CH_{2}OAc$$

$$R_{4} \qquad CH_{2}OAc$$

$$R_{5} \qquad CH_{2}OAc$$

$$R_{7} \qquad CH_{2}OAc$$

$$R_{7} \qquad CH_{2}OAc$$

$$R_{8} \qquad CH_{2}OAc$$

Acetylation of fraction III. A portion of fr. III (62 mg) was treated with 1 ml Ac<sub>2</sub>O and pyridine. The acetylated product (71 mg) was recovered and subjected to CC on silica gel, affording **20** and **21** (39 mg).

Allylic oxidation of 14 with Na<sub>2</sub>CrO<sub>4</sub>. To 14 (117 mg) in  $C_6H_6$  (0.5 ml) were added dry NaOAc (85 mg), Ac<sub>2</sub>O (0.7 ml) and HOAc (0.4 ml) and heated to 40°. When the reaction was complete, H<sub>2</sub>O was added and after 1 hr the mixt. was extracted with Et<sub>2</sub>O. The ethereal part was washed with KHCO<sub>3</sub> and H<sub>2</sub>O to give 145 mg crude product. Silica gel CC, eluting with n-hexane–EtOAc (4:1), yielded 21 (87 mg, 74% yield).

Reduction of 21 with NaBH<sub>4</sub>. To a soln of 21 (87 mg) in MeOH (10 ml), NaBH<sub>4</sub> (3.5 mg) was added. The mixt. was kept at room temp. for 7 hr, then H<sub>2</sub>O and a few drops of 2M HCl were added. The mixt. was extracted with Et<sub>2</sub>O and washed with H<sub>2</sub>O. Evapn of the solvent gave a crude product (78 mg). CC on silica gel afforded 22 (3 mg, n-hexane–EtOAc, 9:1) and a mixt. (37.0 mg), which was acetylated following the usual procedure. The acetylated product (46.0 mg) was recovered and subjected to CC on silica gel, affording 20 (13.0 mg, 28%) and 21 (22.0 mg, 48%).

Reduction of 21 with aluminum isopropoxide. To a soln of 21 (67 mg) in iso-PrOH (3 ml) Al isopropoxide (140 mg) was added. The mixt. was kept at  $56^{\circ}$  for 7 hr, was then extracted with Et<sub>2</sub>O after adding H<sub>2</sub>O. The Et<sub>2</sub>O phase was washed with 0.5 M HCl, 10% NaOH and H<sub>2</sub>O, dried and evapd to give 57 mg of a mixt. which, on silica gel CC, yielded 22 (16 mg).

15-*Acetoxy*-1(10)-ent-*halimen*-18,2 $\beta$ -*olide* (**22**). Oil. IR  $\nu_{\text{max}}^{\text{film}}$  cm<sup>-1</sup>: 2949, 1795, 1732, 1631, 1456, 1368, 1241; <sup>1</sup>H and <sup>13</sup>C NMR: Tables 1 and 3; GC-MS 70

eV, m/z (rel. int.): 392 [M]<sup>+</sup> ( $\ll$ 1), 334 (16), 292 (2), 207 (17), 191 (56), 189 (15), 175 (12), 173 (22), 164 (14), 163 (67), 161 (18), 152 (11), 151 (14), 149 (13), 147 (13), 137 (14), 135 (48), 133 (12), 123 (22), 122 (21), 121 (24), 119 (15), 109 (41), 108 (79), 107 (37), 95 (78), 93 (41), 91 (30), 83 (33), 81 (78), 79 (52), 77 (25), 69 (58), 67 (42), 55 (100), 45 (23).

### REFERENCES

- Urones, J. G., de Pascual, J., Marcos, I. S., Martin, D. D. and Garrido, N. M., *Phytochemistry*, 1987, 26, 1077.
- Urones, J. G., Marcos, I. S., Basabe, P., Sexmero, M. J., Carrillo, H. and Melchor, M. J., *Phytochemistry*, 1994, 37, 1359.
- 3. Urones, J. G., Marcos, I. S., Sexmero, M. J., Basabe, P. and Lithgow, A., *Phytochemistry*, 1990, **29**, 1247.
- 4. Urones, J. G., Marcos, I. S., Sexmero, M. J., Basabe, P. and Lithgow, A. M., *Phytochemistry*, 1990, **29**, 3597.
- de Pascual, J., Urones, J. G., Carrillo, H. and Carrillo, M. A., Anales Quimia, 1979, 75, 140.
- de Pascual, J., Urones, J. G., Basabe, P., Carrillo, H. and Muñoz, A. G., *Phytochemistry*, 1985, 24, 791.
- Urones, J. G., Basabe, P., Marcos, I. S., Alonso, C., Oliva, I. M., Garrido, N. M., Martin, D. D. and Lithgow, A. M., *Phytochemistry*, 1993, 34, 747.

- 8. Urones, J. G., Marcos, I. S., Oliva, I. M., Garrido, N. M., Hagget, J. and Humphreys, V. M., *Phytochemistry*, 1995, **38**, 663.
- 9. Urones, J. G., Marcos, I. S., Basabe, P. and Garrido, N. M., *Phytochemistry*, 1988, 27, 501.
- Urones, J. G., Marcos, I. S., Garrido, N. M. and Basabe, P., Phytochemistry, 1990, 29, 2927.
- Urones, J. G., Marcos, I. S., Garrido, N. M., de Pascual, J. and San Feliciano, A., *Phytochemistry*, 1989, 28, 183.
- Urones, J. G., Marcos, I. S. and Garrido, N. M., *Phytochemistry*, 1990, 29, 3243.
- Urones, J. G., Marcos, I. S. and Garrido, N. M., *Phytochemistry*, 1990, 29, 2585.
- 14. Urones, J. G., Marcos, I. S. and Garrido, N. M. and Moro, R. F., *Phytochemistry*, 1990, **29**, 3042.
- Urones, J. G., Marcos, I. S., Basabe, P., Garrido, N. M., Martin, D. D., Jorge, A., Moro, R. F. and Lithgow, A. M., *Tetrahedron*, 1993, 49, 6079.
- Urones, J. G., Marcos, I. S., Basabe, P., Garrido, N. M., Jorge, A., Moro, R. F. and Lithgow, A. M., Natural Product Letters, 1993, 3, 173.
- Urones, J. G., Jorge, A., Marcos, I. S., Basabe,
   P., Martin, D. D., Garrido, N. M., Lithgow, A.
   M., Fonseca, O. F. and Rodilla, J. M. L., Tetrahedron Letters, 1996, 37, 1659.
- 18. Marcos, I. S., Jorge, A., Diez, D., Basabe, P.,

- Lithgow, A. M., Sexmero, M. J., Garrido, N. M. and Urones, J. G., *Phytochemistry*, 1996, 41, 1155.
- de Pascual, J., Urones, J. G., Marcos, I. S., Martin, D. D. and Monje, V. A., *Phytochemistry*, 1986, 25, 711.
- Urones, J. G., Marcos, I. S., Martin, D. D., Brito, F. M. S. and Rodilla, J. M., *Phytochemistry*, 1987, 26, 3037.
- Urones, J. G., Marcos, I. S., Basabe, P., Diez, D., Garrido, N. M., Alonso, C., Oliva, I. M., Lithgow, A. M., Moro, R. F., Sexmero, M. J. and López, C., *Phytochemistry*, 1994, 35, 713.
- Urones, J. G., Marcos, I. S., Basabe, P., Alonso, C. A., Diez, D., Garrido, N. M., Oliva, I. M., Rodilla, J. M., Slawin, A. M. Z. and Willians, D. J., Tetrahedron Letters, 1990, 31, 4501.
- Urones, J. G., Marcos, I. S., Basabe, P., Alonso, C. A., Martin, D. D., Garrido, N. M. and Oliva, I. M., *Tetrahedron Letters*, 1990, 31, 5665.
- Urones, J. G., Marcos, I. S., Basabe, P., Alonso, C., Oliva, I. M., Garrido, N. M., Martin, D. D. and Lithgow, A. M., *Tetrahedron Letters*, 1992, 33, 5269.
- Urones, J. G., Basabe, P., Marcos, I. S., Alonso, C., Oliva, I. M., Garrido, N. M., Martin, D. D. and Lithgow, A. M., *Tetrahedron*, 1993, 49, 4051.