

NONPROTEIN AMINO ACIDS FROM *CYCAS REVOLUTA*

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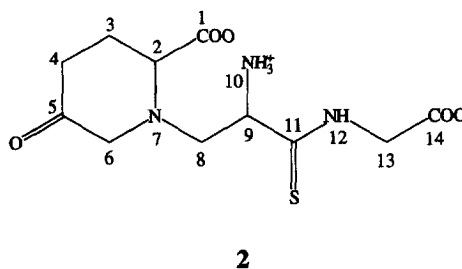
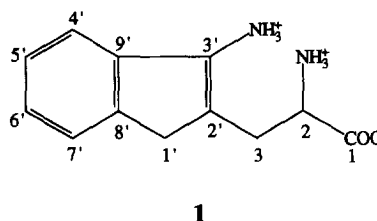
Abstract—Two nonprotein amino acids, cycasindene and cycastioamide, along with eight known nonprotein amino acids, were isolated from the seeds of *Cycas revoluta* Thunb. The structures of cycasindene and cycastioamide were elucidated as 3-[3'-amino-indenyl-2']-alanine (**1**) and *N*-[glyciny-1-11-thio]-5-one-pipecolic acid (**2**) by chemical and spectral methods. ©1997 Elsevier Science Ltd. All rights reserved

INTRODUCTION

Studies with primates and other animals implicated the nonprotein amino acid L-BMAA (β -*N*-methyl-amino-L-alanine) in the neurologic disorder, prevalent among the Chamorro population of Guam, known as amyotrophic lateral sclerosis–parkinsonism dementia complex (ALS-PDC) [1]. L-BMAA was first isolated from the seeds of the Guam cycad, *C. circinalis* L. [2] (which is now recognized as *C. micronesica* K. D. Hill [3]). Further studies on the Guam cycad toxin suggested that the L-BMAA-carbamate, a product of L-BMAA and CO₂ at physiological pH, activated glutamate receptors producing toxic levels of intracellular calcium in neuronal cells [4]. These findings led to an investigation of the nonprotein amino acids in seeds from other *Cycas* species used as food or medicine. The present paper describes the isolation and structural elucidation of two new nonprotein amino acids, cycasindene (**1**) and cycastioamide (**2**), from the seeds of *C. revoluta* Thunb.

RESULTS AND DISCUSSION

Compound **1** exhibited a yellow colour with ninhydrin reagent. The molecular formula of **1** was determined as C₁₂H₁₄N₂O₂ by high resolution FAB mass spectroscopy. 1D and 2D ¹H NMR experiments indicated nine protons in three separate spin systems: CH₂CH₂CHN and an *ortho*-substituted benzene ring. The proton splitting pattern of **1** was similar to indene, but not indole in comparison with reference NMR data



[5]. The ¹³C NMR spectrum of **1** suggested the presence of carbons belonging to a carboxyl group (δ 170.57), an α -amino acid group (δ 54.33), a benzene ring and a quaternary double bond (δ 104.32 and 124.73, respectively). The correlation between H-1' and C-2', -3', -8' in the HMBC spectrum confirmed that **1** contained an indene ring. The very weak coupling between one of the protons at C-3 and C-1' in the COSY spectrum indicated that the alanine side chain was attached to the 2'-position. This observation was in agreement with the HMBC experiment which showed ¹³C-¹H correlations between C-1' and H-2, as well as C-2 and H-1'. Based on the HMQC, HMBC

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Table 1. ^1H and ^{13}C NMR spectral data for compound **1** (D_2O - DCl , 500 MHz)*

C/H	^1H (J in Hz)	^{13}C	^1H - ^{13}C HMBC correlation
1		170.57	
2	3.78 <i>dd</i> (5.0, 10.5)	54.33	C-1, C-3, C-1'
3	3.03 <i>dd</i> (5.0, 16.4)	21.13	C-2, C-2', C-3'
	2.84 <i>dd</i> (10.5, 16.4)		C-2, C-2', C-3'
1'	4.29 <i>d</i> (15.6)	40.05	C-2, C-2', C-3', C-8'
	4.04 <i>d</i> (15.6)		C-2', C-3', C-8'
2'		104.32	
3'		124.73 [†]	
4'	7.27 <i>d</i> (8.0)	117.72	C-2', C-5', C-9'
5'	7.03 <i>t</i> (8.0)	122.31	C-4', C-9'
6'	6.93 <i>t</i> (8.0)	119.46	C-7', C-8'
7'	7.23 <i>d</i> (8.0)	113.34	C-6', C-8'
8'		125.02 [†]	
9'		136.11	

*Assignments confirmed by 2D experiments (COSY and HMQC).

[†]Interchangeable values.Table 2. ^1H and ^{13}C NMR spectral data for compound **2** (D_2O , 500 MHz)*

C/H	^1H (J in Hz)	^{13}C	^1H - ^{13}C HMBC correlation
1		178.74	
2	4.33 <i>m</i>	61.25	C-1, C-3
3	2.00 <i>m</i>	22.69	C-1, C-2, C-4
	2.43 <i>m</i>		C-1, C-2, C-4
4	2.54 <i>m</i>	29.79	C-2, C-3, C-5
5		179.91	
6	3.73 <i>d</i> (14.4)	42.54	C-2, C-5, C-8
	3.88 <i>d</i> (14.4)		C-2, C-5, C-8, C-9
7			
8	2.88 <i>dd</i> (15.0, 4.3)	30.94	C-6, C-9, C-11
	3.13 <i>dd</i> (15.0, 9.4)		C-6, C-11
9	4.33 <i>m</i>	51.89	C-8, C-11
10	6.98 <i>t</i> [†]		
11		168.01	
12	8.17 <i>s</i> [†]		
13	3.73 <i>dd</i> (17.2)	43.24	C-12, C-14
	3.88 <i>dd</i> (17.2)		C-12, C-14
14		175.81	

*Assignments confirmed by 2D experiments (COSY and HMQC).

[†]10% D_2O -90% H_2O .

and COSY spectra, **1** was identified as 3-[3'-amino-indenyl-2']-alanine, named here as cycasindene.

Compound **2** also developed a yellow colour with ninhydrin reagent, but the yellow colour turned to red in a few hours. High voltage electrophoresis results demonstrated that **2** was an acidic amino acid. Compound **2** decomposed during acid hydrolysis, suggesting that it was not a peptide. The elemental composition of **2** was deduced as $\text{C}_{11}\text{H}_{17}\text{N}_3\text{O}_5\text{S}$ from a high resolution CI mass spectrum. 1D and 2D ^1H NMR spectra in D_2O indicated that **2** had 12 protons belonging to four isolated coupling systems: CH_2 , CH_2 , CH_2CHN , $\text{CH}_2\text{CH}_2\text{CHN}$. The ^{13}C NMR spectrum showed that **2** had four quaternary carbons: two carboxyl, one carbonyl, and one amide-type. The HMBC experiment established that two protons with the same

chemical shift at δ 4.33 were correlated to two different carbons both of which had amino groups attached. The connections between carbons and protons were established from COSY, HMQC and HMBC spectra. Two moieties in **2**, the oxidized pipecolic acid ring and the side chain, were in agreement with MS fragments of m/z 142 and 161. A broad singlet for one proton at δ 8.17 was shown to be coupled to both protons at C-13 on the basis of PRESAT (1D Water Suppression) and COSY experiments in D_2O - H_2O (1:9). The latter spectrum verified that the proton at δ 8.17 belonged to an amide-type group and that the imino group (N-12) was connected to C-11 and C-13. The JUMPRET (Jump-and-Return Water Suppression, D_2O - H_2O) exhibited a triplet of three protons at δ 6.98 and a broad singlet for the proton at δ 8.17. The ratio of the

triplet was 1:1:1, indicative of quadrapole coupling with ^{14}N , thus implicating an ammonium ion (N-10) on the side chain. On the ^{15}N - ^1H HMQC spectrum, the triplet was correlated to the ammonium ion N at δ 25, and the singlet at δ 8.17 was correlated to what must be a thioamide N at δ 117 [6]. From the MS and NMR data, **2** was concluded to be *N*-[glyciny-alaninyl-11-thio]-5-one-pipecolic acid, named here as cycasthioamide. This is the first report of a thioamide nonprotein amino acid from plants.

EXPERIMENTAL

Plant material. Seeds of *C. revoluta* Thunb. were collected from Rockport, Texas, U.S.A., in December 1995. Seed and leaf vouchers are deposited in the Plant Resources Center, University of Texas at Austin, Voucher no. Mabry-Pan 210.

Extraction and isolation. Ground fresh endosperm (5.4 kg) was extracted with 80% EtOH at room temp and the extract was defatted with CHCl_3 . The concd extract was applied to an AG 50W-X4 cation exchange resin column. The column was eluted with an ammonium hydroxide gradient (0.2–1 M). Twenty frs were collected. Each fr. was further chromatographed on a cellulose column, eluted with *iso*-PrOH– H_2O gradient (4:1–1:1). **1** (13 mg) was obtained from frs 8 and 9 of the ion exchange CC, and final purification by cellulose CC with *iso*-PrOH– H_2O (4:1). **2** (12 mg) was isolated from fr. 5 and final purification by cellulose CC with *iso*-PrOH– H_2O (2:1).

Eight known nonprotein amino acids were isolated and identified as α -aminoadipic acid, β -alanine, α -, β - and γ -aminobutyric acid, citrulline, BMAA and hydroxyarginine by comparison with authentic non-protein amino acids using TLC, HVE (high voltage electrophoresis) and GC.

Cycasindene (1). Microcrystals. HRFABMS: m/z 217.0958 [$\text{M} - \text{H}$] $^+$, (calcd 217.0977) for $\text{C}_{12}\text{H}_{13}\text{N}_2\text{O}_2$. ^1H and ^{13}C NMR: Table 1.

Cycasthioamide (2). Amorphous powder. CIMS: m/z 302 [$\text{M} - \text{H}$] $^+$, 142, 161. HRCIMS: m/z 302.0805 [$\text{M} - \text{H}$] $^+$, (calcd 302.0810) for $\text{C}_{11}\text{H}_{16}\text{N}_3\text{O}_3\text{S}$. ^1H and ^{13}C NMR: Table 2.

Acid hydrolysis of 2. Solns of **2**, each containing 0.5 mg **2**, in 0.1 ml each of 1 M, 2 M and 6 M HCl were heated at 100° for 2 hr. After cooling, each soln was concd to dryness. Each sample was analysed for amino acids by TLC and HVE. **2** was decomposed during acid hydrolysis. Neither **2** nor any identifiable protein or nonprotein amino acids were detected in the hydrolysates.

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