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# A L-GALACTOSE-CONTAINING CARRAGEENAN FROM CYSTOCARPIC GIGARTINA SKOTTSBERGII

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**Key Word Index**—*Gigartina skottsbergii*; Rhodophyta; carrageenans; L-galactose; methylation; <sup>13</sup>C NMR.

Abstract—Alkaline treatment of a  $\kappa$ /1-carrageenan from cystocarpic Gigartina skottsbergii and further fractionation with potassium chloride, led to the isolation of a minor product soluble in 2 M KCl with unusual structural features. It contains small amounts of L-galactose and is built up from  $\kappa$ /1- and  $\lambda$ -blocks. © 1997 Elsevier Science Ltd. All rights reserved

#### INTRODUCTION

The structures of the major carrageenans extracted from cystocarpic Gigartina skottsbergii have been thoroughly studied [1–3]. In a previous paper, we reported some structural features of an L-galactose-containing product, soluble in 2 M KCl, obtained by alkaline treatment of a partially cyclised  $\mu/\nu$ -carrageenan and further fractionation with potassium chloride [4]. Some unusual galactans with similar characteristics have been isolated from tetrasporophytes [5, 6] and from cystocarpic plants of other algae belonging to the Gigartinaceae [7]. We report herein, the characterization and structural analysis of another 2 M KCl-soluble fraction, isolated when the same procedure described above was carried out on a gelling carrageenan of the same origin.

### RESULTS

 $\kappa$ /1-Carrageenan 1C<sub>1</sub> [3] was submitted to an alkaline treatment and the modified product (1C<sub>1</sub>T) was further fractionated with potassium chloride to yield a minor fraction soluble in 2 M KCl (SF). Yield, analysis and optical rotation of SF are shown in Table 1; it contains only galactose and 3,6-anhydrogalactose (molar ratio 1.27:1.00) and the ratio D:L-galactose is 1.00:0.14. Chromatography of SF on DEAE Sephadex A-50 (Cl<sup>-</sup>) resulted in irreversible binding of the sample to the resin; SF was not recovered even

SF was converted into the triethylammonium salt [8] and methylated using the Hakomori procedure [8, 9], in the conditions usually employed for carrageenans of the  $\kappa$  family [5]. Table 2 indicates the composition of the permethylated derivative. SF is comprised mainly of 3-linked galactose 4-sulphate and 2-sulphate units in a molar ratio of 1.6:1.0, and 4-linked 3,6-anhydrogalactose and 3,6-anhydrogalactose 2-sulphate residues in a molar ratio of 1.0:1.0; non-substituted  $\beta$ - and  $\alpha$ -galactose units also contribute but in minor proportion to the structure of SF. Significant amounts of 2,3,4,6-tetra-O-methylgalactose (~4.5%) in permethylated SF would indicate the presence of single stubs of galactose, while 3-O-methylgalactose arises from 4-linked galactose 2,6disulphate units not cyclised during the sequence of alkaline treatment methylation. Minor quantities of other mono-O-methylgalactose residues are consistent with the presence of disulphated units in the original

The 75 MHz  $^{13}$ C NMR spectrum of SF showed, in the anomeric region, major signals at  $\delta$  103.0 (with a shoulder at 102.9), 95.8 and 92.6, and smaller peaks at  $\delta$  104.0 and 92.1, which were assigned to  $\kappa$ -, 1- and  $\lambda$ -diads; resonances of the other carbon atoms of these diads are also present in the spectrum (Table 3) [10–12]. The molar ratio of 3,6-anhydrogalactose: 3,6-anhydrogalactose 2-sulphate, calculated from the area of the corresponding anomeric signals, is 1.6:1.0.

# DISCUSSION

The matrix galactans of the Rhodophyta consist of linear chains of alternating 3-linked  $\beta$ -galactose (A

after treatment of the anion-exchanger with boiling 4 M NaCl or 0.5 M NaOH in 3.5 M NaCl.

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Table 1	Vield	analysis and	Lontical	Irotation	of SE*
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	Yield†			Carbohydrate composition (mol%)				[α] <sub>D</sub> (°)	
Fraction	(%)	D-Gal	ւ-Gal	D-3,6-AnGal	D-3,6-AnGal 2S	L-3,6-AnGal	(mol %)		
SF	0.9 (3.0)	49.1	6.9	26.8§	17.2§	_	85.0	+24.0	
Fs¶	0.2 (1.6)	48.8	24.5		8.9	17.8	41.1	-13.0	
$T_3$	8.6 (10.2)	68.6	29.4	<b>———</b>	<del>2</del> .0		71.6	+11.2	

- \* Fractions Fs [4] and T<sub>3</sub> [5] were included for comparison.
- † Yield from 100 g of whole carrageenan 1C (in parentheses, % of total recovered from 1C) [1].
- # Moles of sulphate per 100 mol sugar.
- § Determined from areas of corresponding anomeric signals in <sup>13</sup>C NMR spectrum.
- ¶ Fs also contains minor quantities of Rha (1.2%), Xyl (2.4%), Man (4.3%), Glc (5.3%) and 3-O-methylgalactose (3.0%).

units) and 4-linked  $\alpha$ -galactose (B units). The A units always occur in the D-form, while the B units can occur in either D- (carrageenans) or L- (agars) form. This difference defines two families of polysaccharides which are bridged by a group of polysaccharides containing variable amounts of D- and L-galactose B-units interspersed in the same molecule [13].

Carrageenophytes of the Gigartinaceae and Phyllophoraceae, biosynthesise different structures according to the stage of the life-cycle [13, 14]. For G. skottsbergii and Iridaea undulosa, it has been proved that carrageenans of the  $\kappa$ -family are produced by cystocarpic plants, whereas tetrasporic plants yield  $\lambda$ -carrageenans [1–3, 5]. This clear-cut scheme has been altered recently by reports of the biosynthesis of minor quantities of L-galactose-containing galactans by cystocarpic [4, 7] and tetrasporic carrageenophytes [5, 6]. In addition, these molecules could be hybrids built up by structures of the  $\kappa$ - and  $\lambda$ -types [4–6].

SF was obtained after an elaborated fractionation procedure which comprised a sharp potassium chloride precipitation (between 0.30–0.31 M KCl) of a  $\kappa/1$ fraction (1C<sub>1</sub>) from the raw carrageenan [1], its alkaline treatment and a further potassium chloride precipitation of the gelling derivatives. SF remained soluble in 2 M KCl and this solubility behaviour seems to be a general characteristic of these galactans [4-6]. Such soluble polysaccharides obtained from cystocarpic plants were always isolated after alkaline treatment of the starting material [4, 7]. This procedure is similar to that of ref. [15], used for the fractionation of a mixture of  $\mu/\nu$ - and  $\lambda$ -carrageenans obtained from an unsorted sample of Chondrus crispus. This approach indicates complexation between  $\mu/\nu$ - and  $\lambda$ -structures, which is broken by cyclisation of the 4-linked α-galactose 6-sulphate or 2,6-disulphate units. The interaction between different carrageenan structures has been reported previously [5], showing its influence on the potassium chloride solubilities of the individual products. Moreover, the solubility behaviour of SF would indicate that this product is a block copolymer with  $\kappa/1$ - and  $\lambda$ -domains as the complexation requires, not only complementary interaction sites but also their arrangement in ordered polyvalent arrays [16]. The unsuccessful anionexchange chromatography of SF, as well as the electrophoresis of similar galactans obtained from *Chondrus crispus*, *G. canaliculata*, *G. leptorynchos* and *Mastocarpus stellata* [7] are consistent with the presence of a copolymer with  $\kappa/1$ - and  $\lambda$ -blocks. The existence of these blocks would also explain why a product with mainly a  $\kappa/1$ -structure (59%) does not have gelling properties.

It may be unusual to attribute a  $\lambda$ -structure to a product obtained after an alkaline treatment but, because the starting material (1C<sub>1</sub>) was a  $\kappa$ /1-carrageenan [1-3], the conditions of this treatment were those reported previously for polysaccharides of the  $\kappa$ -family [17]. It is known that the cyclisation reaction is 20–60 times faster for carrageenans of the  $\kappa$ -family than for those of the  $\lambda$ -family [18]. Taking into account the amount of  $\lambda$ -structure in the precursor of SF (given by the percentage of 4,6-di-O-methylgalactose in the methylated derivative), it may be estimated from the half-life [18] that  $\sim 2.5\%$  of the  $\alpha$ galactose 2,6-disulphate units linked to  $\beta$ -galactose 2sulphate residues were cyclised during the alkaline treatment and, therefore, SF should still contain  $\sim 12\%$  of the former residues. This result is in agreement with the signals found at  $\delta$  104.0 and 92.1 in the <sup>13</sup>C NMR spectrum of SF and with the detection of  $\sim 3\%$  of 3-O-methylgalactose in its methylated derivative, because cyclisation proceeds further during methylation. A new alkaline treatment of SF for 24 hr ( $\sim$ eight half-lives for the cyclisation of  $\alpha$ -galactose 2,6-disulphate residues in  $\lambda$ -carrageenans) [18] led to an increase of  $\sim 3\%$  in the 3,6-anhydrogalactose content, in agreement with the above discussion and with the resistance shown by the  $\alpha$ -units to cyclisation.

The small amount of L-galactose is probably interspersed in the carrageenan structure because, if present as traces of a contaminant galactan, it would probably had been lost during the isolation procedure of SF.

In conclusion, cystocarpic G. skottsbergii biosynthesises not only the major  $\kappa/1$ - and  $\mu/\nu$ -carrageenans previously reported [1–3] but also a small amount of a polymer that would be a hybrid containing  $\kappa$ - (41%), 1- (18%) and  $\lambda$ - (29%) structures and single stubs of galactose ( $\sim$ 4.5%). Products with similar charac-

Table 2. Composition of sugars produced by permethylation and hydrolysis of SF\*

	3,6-AnGal	2	20.3 20.7	←41.0→	←25.9→	←2.0→
			Ħ.	Ħ.	1	
		4	←2.7→	1.9	9.6	tr
		3	<b>←</b> 2	3.5	†	tī
indicated		2	1.7	3.6	2.1	1.7
Mol % of sugars having methyl groups at the positions indicated		9	3.7	<b>←4.8</b> →		8.0
oups at the		2,3 6	1.9 3.7	±4.	I	4.3
g methyl gr	ose	2,4	-	ļ	***************************************	25.3
ars having	Galactose	3,4 2,4	tr§	-	2.4	
₹ns Jo % Ic		4,6	4.0→	5 14.6	16.2	10.7
Ĕ		2,6 4,6	÷3.	23.5	18.7	16.3
		3,4,6	2.1	tī.	1	1.6
		2,4,6	3.5	1.8	5.5	9.5
		2,3,6	3.3	2.2	3.0	9.9
		2,3,4,6	6.1	3.2	19.5	14.0
	Fraction	i	SF†,‡	SF	Fs¶,**	T3¶††

\*Composition of permethylated derivatives of Fs and T<sub>3</sub> given for comparison.

† Subjected to reductive hydrolysis and further acetylation.

† Minor amounts of 2,3,6-tri-O-methylglucose (1.9%) and traces of glucose detected.

§Percentages lower than 1% are considered as trace (tr).

| Hydrolysed with 2 M TFA and derivatised to aldononitrile acetates. | Hydrolysed with 45% HCO<sub>2</sub>H and derivatised to aldononitrile acetates.

\*\* Permethylated Fs also contains 2,3,4-tri-O-methylxylose (8.0%) and minor quantities of 2,4-di-O-methyl- and 2,3-di-O-methyl-xylose and 2,3,6-tri-O-methylglucose. †† In T, 2,3,4-tri-O-methylxylose (2.5%) detected. 1012 M. Ciancia et al.

Table	3 13C	NMR	cional	assignment	of SE
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Diad	C-1	C-2	C-3	C-4	C-5	C-6
3-Linked β-D-Gal 4-sulphate	103.0	70.1	79.4	74.8	75.5	61.9
4-Linked α-D-3,6-AnGal	95.8	70.3	79.8	78.9	77.4	70.1
3-Linked β-D-Gal 4-sulphate	102.9*	70.1	77.4	72.8	75.4	61.9
4-Linked α-D-3,6-AnGal 2-sulphate	92.6	75.5	78.5	78.9	77.6	70.3
3-Linked β-D-Gal 2-sulphate	104.0	78.1	76.4	64.8	74.7	61.9
4-Linked α-D-Gal 2,6-disulphate	92.1	75.5	70.1	81.0	69.3	68.8

<sup>\*</sup> Shoulder on signal at  $\delta$  103.0.

teristics have been reported in the  $\mu/\nu$ -fraction of the same seaweed [4], in gamethophytes of *C. crispus*, *G. canaliculata*, *G. leptorynchos* and *M. stellata* [7] and in tetrasporophytes of *G. skottsbergii* [6] and *I. undulosa* [5], suggesting that they are usual minor components of the carrageenans produced by seaweeds belonging to the Gigartinaceae.

#### **EXPERIMENTAL**

Material. Cystocarpic G. skottsbergii was collected in Bahía Camarones (Provincia de Chubut, Argentina) and sorted in the Instituto Nacional Patagónico (CONICET) (Puerto Madryn, Chubut).

General. Sulphate was analysed by the method of ref. [19]. 3,6-Anhydrogalactose was determined, before and after alkaline treatment of the sample, by the resorcinol method [20]. Optical rotations were measured using a 1.1% soln of the sample in  $H_2O$ . The monosaccharide composition was determined by reductive hydrolysis of the sample [8] and derivatization for GC. GC of the alditol acetates [21] was performed on a fused-silica capillary column (30 m × 0.25 mm i.d.), WCOT coated with SP-2330 (film thickness 0.20  $\mu m$ ). The oven temp. was 220° and the inj. and FID temps were 235°. N<sub>2</sub> was used as carrier at 1 ml min<sup>-1</sup> with split ratio of 80:1. Determination of D:L-galactose was carried out by GC of the acetylated 1-deoxy-1-(2'-hydroxypropylamino) alditols [22]. <sup>13</sup>C NMR were recorded at 75 MHz with proton decoupling at 80°, with ext. TMS as ref.; a soln of the sample (30 mg) in 1:0.2 H<sub>2</sub>O-D<sub>2</sub>O (0.4 ml) and a 5 mm-tube were used. Specific parameters include a pulse angle of 90°, an acquisition time of 0.27 sec, no pulse delay and a spectral width of 15 kHz; the number of scans was 720 000.

Alkaline treatment of  $1C_1$  and fractionation of the modified product. Alkaline treatment of the sample (4 g) was carried out in 0.5 M NaOH-0.5 M NaCl (3 l) for 90 min at  $80^{\circ}$  [17]; the soln was neutralised with HOAc with constant agitation. Fractionation of  $1C_1T$  was carried out as indicated for whole carrageenan [1]. In order to achieve further cyclisation of  $\alpha$ -galactose 2,6-disulphate residues in  $\lambda$ -carrageenan, alkaline treatment was carried out in M NaOH for 24 hr at  $80^{\circ}$ C [18].

Methylation analysis of SF. The sample (5 mg) was converted into the corresponding triethylammonium salt and methylated by the Hakomori procedure [9] as described in refs [5, 8]. One portion of permethylated SF was subjected to reductive hydrolysis [8] and the partially methylated alditols were acetylated [21]; another portion of the permethylated sample was hydrolysed with 2 M TFA for 2 hr at 121° and the partially methylated galactoses derivatised to the aldononitrile acetates [23]. GC of the alditol acetates and aldononitrile acetates was carried out as described in ref. [4]. GC-EIMS of the alditol acetates [24, 25] and aldononitrile acetates [23] was performed at 70 eV; MS were recorded over the range 40-500 mu. Chromatography was carried out using the SP-2330 column, programmed to run with an initial 1 min hold at 130° and then at 6° min<sup>-1</sup> to 230°. The He flow rate was 1 ml min<sup>-1</sup>, the inj. temp. 240° and the split ratio 100:1.

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## REFERENCES

- Matulewicz, M. C., Ciancia, M., Noseda, M. D. and Cerezo, A. S., Phytochemistry, 1989, 28, 2937.
- 2. Matulewicz, M. C., Ciancia, M., Noseda, M. D. and Cerezo, A. S. *Phytochemistry*, 1990, **29**, 3407.
- Ciancia, M., Matulewicz, M. C., Finch, P. and Cerezo, A. S., Carbohydrate Research, 1993, 238, 241
- Ciancia, M., Matulewicz, M. C. and Cerezo, A. S., *Phytochemistry*, 1993, 34, 1541.
- 5. Stortz, C. A. and Cerezo, A. S., Carbohydrate Research, 1993, 242, 217.
- Noseda, M. D., Polissacarídeos sulfatados da fase tetrasporofitica de Gigartina skottsbergii (Rhodophyta, Gigartinales) Thesis, Federal University of Paraná, Brazil, 1994.
- 7. Craigie, J. S. and Rivero-Carro, H. in XIVth International Seaweed Symposium, Abstracts. Brest-St. Malo, 1992, p. 71.

- 8. Stevenson, T. T. and Furneaux, R. H., Carbohydrate Research, 1991, 210, 277.
- 9. Hakomori, S., Journal of Biochemistry (Tokyo), 1964, 55, 205.
- 10. Stortz, C. A. and Cerezo, A. S., *Carbohydrate Polymers*, 1992, **18**, 237.
- Stortz, C. A., Bacon, B. E., Cherniak, R. and Cerezo, A. S., Carbohydrate Research, 1994, 261, 317.
- 12. Falshaw, R. and Furneaux, R. H., Carbohydrate Research, 1994, 252, 171.
- 13. Painter, T. J, *The Polysaccharides*, Vol. 2, G. O. Aspinall. Academic Press, London, 1983, p. 195.
- McCandless, E. L., West, J. A. and Guiry, M. D., Biochemistry Systematics and Ecology, 1982, 10, 275.
- Anderson, N. S., Dolan, T. C. S., Lawson, C. J., Penman, A. and Rees, D. A., Carbohydrate Research, 1968, 7, 468.
- Spillmann, D., Glycoconjugate Journal, 1994, 11, 169.

- Ciancia, M., Matulewicz, M. C. and Cerezo, A. S., Carbohydrate Polymers (in press).
- 18. Ciancia, M., Noseda, M. D., Matulewicz, M. C. and Cerezo, A. S., *Carbohydrate Polymers*, 1993, **20**, 95.
- 19. Dodgson, K. S. and Price, R. G., *Biochemistry Journal*, 1962, **84**, 106.
- Yaphe, W., Analytical Chemistry, 1960, 32, 1327.
- 21. Sloneker, J. H., Methods In Carbohydrate Chemistry, 1972, 6, 20.
- 22. Cases, M. R., Cerezo, A. S. and Stortz, C. A., Carbohydrate Research, 1995, 269, 333.
- 23. Stortz, C. A., Matulewicz, M. C. and Cerezo, A. S., Carbohydrate Research, 1982, 111, 31.
- Lindberg, B., Methods in Enzymology, 1972, 28, 178.
- Lönngren, J. and Svensson, S., Advances in Carbohydrate Chemistry and Biochemistry, 1974, 29, 49.