



IRIDOID GLYCOSIDES FROM *EUCNIDE BARTONIOIDES*

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Key Word Index—*Eucnide bartonioides*; Loasaceae; iridoid glycosides; morroniside; kingside; sweroside; secologanol; 8-*epi*-loganin; loganin; 5-hydroxyloganin.

Abstract—*Eucnide bartonioides* yielded morroniside as the main iridoid constituent. In addition, six minor iridoid glucosides were isolated namely the known glucosides: kingside, sweroside, secologanol, 8-*epi*-loganin and loganin as well as a novel iridoid glucoside, which by spectroscopic methods was assigned to be 5-hydroxyloganin. The compound 8-*epi*-loganin has so far been reported only from Scrophulariaceae and related families and it is rather unexpected in Loasaceae. © 1997 Elsevier Science Ltd. All rights reserved

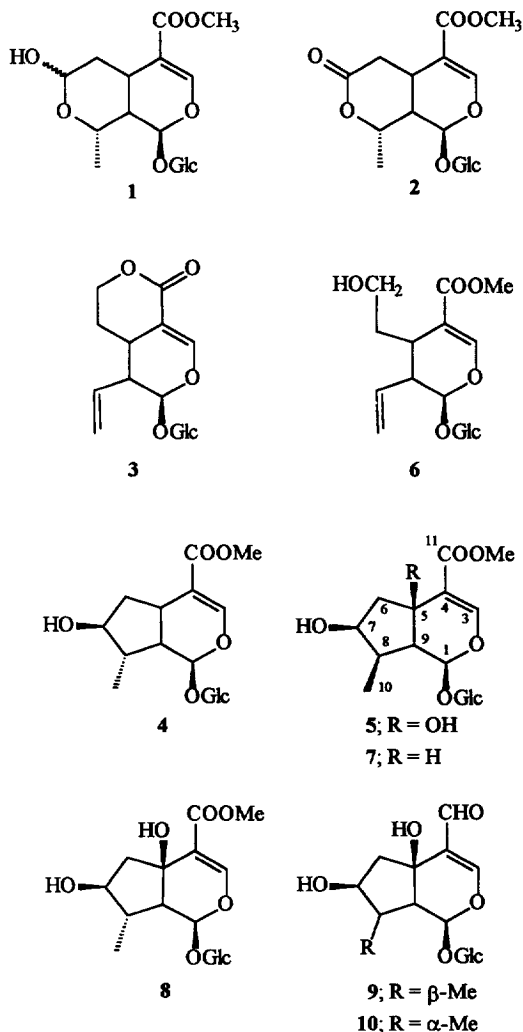
INTRODUCTION

The taxonomic position of Loasaceae has for a long time been a matter of controversy. Thus, Thorne [1] has placed this family in its own order near Myrtales, Cronquist [2] preferred the more traditional position in Violales while Dahlgren believed in an affinity to his Cornales-Dipsacales complex as well as to Gentianales [3], mainly due to the embryological characters together with the presence of iridoids in the family [4]. Previous chemical work on these compounds has been done mainly on *Mentzelia* [5–9] although iridoids have been reported from other genera too [10–13]. In continuation of our research on iridoid glycosides from Loasaceae, we have now examined *Eucnide bartonioides* Zucc. To our knowledge, no previous phytochemical study of this genus has been reported.

RESULTS AND DISCUSSION

The water-soluble part of an ethanolic extract of *E. bartonioides* was fractionated by reverse phase chromatography to give as the major compound morroniside (1), seen in the ^1H NMR spectrum as a mixture of the anomeric forms [14]. Minor constituents were kingside (2), sweroside (3), 8-*epi*-loganin (4), an unknown (5), secologanol (6) and loganin (7).

The ^{13}C NMR spectrum of 5 (Table 1) contained 17 signals of which six could at once be assigned to a β -glucopyranosyl moiety. The remaining 11 signals fitted well with a loganin-like structure, including the secondary hydroxyl group but with an additional tertiary hydroxyl substituent (δ 72.9) which therefore had



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Table 1. ^{13}C NMR data for iridoid glucosides in CD_3OD

C	5	8*	9†	10‡
1	96.0	95.68	97.12	96.5
3	152.4	153.43	162.70	164.0
4	115.4	115.26	126.78	126.4
5	72.9	71.47	72.12	70.4
6	49.0	48.05	48.88	46.8
7	74.4	77.93	73.38	77.7
8	41.4	43.57	41.23	43.0
9	55.0	51.56	54.72	51.1
10	13.1	13.84	13.14	13.9
11	168.2	168.07	192.95	192.5
1'	99.9	99.68	100.12	98.8
2'	73.5	74.41	74.37	74.2
3'	77.6	77.54	78.43	77.3
4'	71.6	71.73	71.54	71.5
5'	78.5	78.41	77.55	78.4
6'	62.7	62.84	62.69	62.7
OMe	51.6	51.62	—	—

* Data from ref. [16].

† Data from ref. [17].

‡ Data from ref. [18].

to be situated at C-5 or C-8. The ^1H NMR spectrum of **5** showed a singlet for the 11-carbomethoxy group at δ 3.72. Another singlet at δ 7.40 was assigned to H-3 and proved that the carbon atom at C-5 must be completely substituted. The doublet nature of the C-10 methyl group at δ 1.10 further showed that the second hydroxy group was not at C-8. The coupling pattern allowed assignment of all the signals from H-6 through H-9 and H-1 (see Experimental), proving the position of the secondary hydroxyl group to be at C-7. This was in agreement with the overall structure **5**. Assuming the usual configuration at C-1, C-5 and C-9, only the stereochemistry at C-7 and C-8 remained to be established. One compound with the same gross structure is known, namely auroside (**8**) isolated from *Phlomis aurea* [15]; however, no data were reported. Fortunately, **8** has been prepared synthetically [16], and NMR data were thus available (Table 1) to show the non-identity of **5** and **8**. Other compounds with a similar structure are known, namely the 8-epimeric pair tecomoside/*epi*-tecomoside (**9/10**), both with a C-11 aldehyde function [17, 18]. Comparison of the ^{13}C NMR spectra of **5** and **8** with those of the pair **9/10** (particularly the shift values for C-7, C-8 and C-9) showed that **5** and **8** had to be epimeric at C-8, too. It is significant that the C-9 resonances in **5** and **9** with a β -methyl group at C-8 are more low field (δ 55) than those of **8** and **10** with the α -methyl group (δ 51) [17]. Furthermore, comparisons of the coupling constants for the protons of the cyclopentane ring in compounds **5/8** and **9/10** confirm the relative configuration to be 8β -methyl in compound **5** (see Table 2). Particularly, the large size of $J_{8,9}$ (12 Hz) show that H-8 and H-9 are in a transdiaxial position in **5** and **8**.

Table 2. Coupling constants for the protons of the cyclopentane ring in compounds **5** and **8–10**

Compound	$J_{1\beta,9\beta}$	$J_{6\alpha,7\alpha}$	$J_{6\beta,7\alpha}$	$J_{7\alpha,8\alpha}$	$J_{8\alpha,9\beta}$	$J_{8\beta,9\beta}$
5	2.0	5.6	2.7	5.1	12.0	—
8*	1.6	6.7	5.6	§	—	10.3
9†	1.7	5.8	2.7	5.8	12.0	—
10‡	1.1	5.6	7.3	§	—	10.8

* Data from ref. [16].

† Data from ref. [17].

‡ Data from ref. [18].

§ No data given.

Within Loasaceae, *Eucnide* has been connected with *Mentzelia* and *Schismocarpus* in the subfamily Mentzelioidae [17]. The carbocyclic iridoids found in *E. bartonioides* are similar to schismoside isolated from *Schismocarpus matudai* [12], but different from the compounds isolated from the genus *Mentzelia*. Apparently, this genus is the only one in the family which contains iridoids lacking C-10. However, both types of iridoids can be presumed to be biosynthetically derived from iridodial with the 8β -stereochemistry [12, 20]. The finding of 8-*epi*-loganin in Loasaceae is unexpected since compounds with the 8α -methyl stereochemistry have so far been restricted to the Scrophulariales/Lamiales complex. We do not consider this compound to have taxonomic significance, since the remaining compounds (including the secoiridoids) all belong biosynthetically to the 8β -series which is characteristic for Cornales/Dipsacales and Gentianales.

EXPERIMENTAL

General procedures. Mps. uncorr.; ^1H and ^{13}C NMR: CD_3OD at 250 and 63 MHz, respectively, (the solvent peak was used as int. standard). Prep. TLC: 20×40 cm plates coated with 1 mm layers of Silica Gel PF₂₅₄ (Merck); bands were detected in UV light (254 nm); Reverse phase MPLC: Merck Lobar C-18 columns size B and C. H_2O –MeOH mixts were used as eluents and peaks were detected by UV at 210 nm.

Plant material. *Eucnide bartonioides* was grown by The Botanical Garden of Copenhagen at the field station at Tåstrup. (The voucher IOK 25-96 was deposited at The Botanical Museum, The University of Copenhagen and identified by Dr Bertel Hansen). It was either used fresh or frozen in polyethylene bags and kept at -23° .

Isolation of iridoids. Fresh whole plants (142 g) were extracted with EtOH (2×0.4 l), filtered, taken to dryness and partitioned in H_2O –Et₂O. The aq. layer was concd, redissolved in MeOH and treated with activated charcoal. Evapn of the filtrate gave a white foam (4 g), which was subjected to MPLC (C-column) eluting with H_2O –MeOH mixts (10:1 to 2:1). This gave (in order of elution) kingiside (**2**; 12 mg), morroniside (**1**; 509 mg, 0.35%), sweroside (**3**; 10 mg), a

mixture of **4** and **5** (20 mg), secologanol (**6**, 20 mg) and loganin (**7**; 10 mg). The above mixt. was sepd. by prep. TLC (CHCl₃-MeOH; 3:1) to give 8-*epi*-loganin (**4**) (5 mg) and 5-hydroxyloganin (**5**; 10 mg). The known compounds were identified by comparison of the NMR spectra with those of authentic samples.

5-Hydroxyloganin (5). $[\alpha]_D^{21} - 88^\circ$ (MeOH; *c* 0.6); UV $\lambda_{\max}^{\text{MeOH}}$ nm: 232; MS *m/z* (rel. int.): 424 $[\text{M} + \text{NH}_4]^+$ (0.04), 406 $[\text{M} + \text{NH}_4 - \text{H}_2\text{O}]^+$ (0.07), 191 $[\text{M} - \text{Glu} - 2\text{H}_2\text{O}]^+$ (100); ¹H NMR (CD₃OD): δ 7.40 (*s*, H-3), 5.63 (1H, *d*, *J* = 2.0 Hz, H-1), 4.57 (1H, *d*, *J* = 8.0 Hz, H-1'), 3.92 (1H, *ddd*, *J* = 5.6, 5.1 and 2.7 Hz, H-7), 3.89 (1H, *dd*, *J* = 12.0 and 1.8 Hz, H_a-6'), 3.72 (3H, *s*, COOMe), 3.67 (1H, *dd*, *J* = 12.0 and 5.2 Hz, H_b-6'), *ca* 3.3 (3H, H-3', H-4' and H-5'), 3.18 (1H, *dd*, *J* = 9.0 and 8.0 Hz, H-2'), 2.47 (1H, *dd*, *J* = 15.0 and 5.6 Hz, H_a-6), 2.33 (1H, *dd*, *J* = 12.0 and 2.0 Hz, H-9), 2.16 (1H, *dd*, *J* = 15.0 and 2.7 Hz, H_b-6), 1.67 (1H, *ddq*, *J* = 12.0, 5.1 and 7.0 Hz, H-8), 1.10 (3H, *d*, *J* = 7.0 Hz, 10-CH₃); ¹³CNMR: Table 1.

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