

PII: S0031-9422(97)00210-0

AN EPITETRATHIODIOXOPIPERAZINE RELATED TO EMESTRIN FROM EMERICELLA FOVEOLATA

MIKIKO OOIKE, KOOHEI NOZAWA and KEN-ICHI KAWAI*

Faculty of Pharmaceutical Sciences, Hoshi University, Ebara 2-4-41, Shinagawa-ku, Tokyo 142, Japan

(Received 18 December 1996)

Key Word Index—Emericella foveolata; E. striata; fungi; epipolythiodioxopiperazine; secoemestrin C; emestrin; emestrin B; aurantioemestrin; dethiosecoemestrin.

Abstract—An epitetrathiodioxopiperazine, secoemestrin C, related to the macrocyclic antifungal epidithiodioxopiperazine, emestrin, originally isolated from *Emericella striata* and *E. quadrilineata*, was isolated along with the epitrithiodioxopiperazine, emestrin B, from the culture medium of the early state of *E. foveolata*. This fungus also produced emestrin in the mycelium and dethiosecoemestrin and violaceic acid in the culture broth after two weeks cultivation. The structure of secoemestrin C was established by spectroscopic and chemical investigation. Secoemestrin C may be the key intermediate from emestrin and/or emestrin B to dethiosecoemestrin. © 1997 Elsevier Science Ltd. All rights reserved

INTRODUCTION

The novel antifungal macrocylic epipolythiodioxopiperazines, emestrin (1) [1] and emestrin B (2) [2], have been isolated from mycelial extracts of Emericella striata. Emestrin (1) was also isolated from E. quadrilineata, as mycotoxin EQ-1 [3], E. nidulans var. acristata and E. parvathecia. Positive coloration (dark brown) with aqueous silver nitrate [4] was used for detecting epipolythiodioxopiperazines on TLC. From the extract of the culture filtrate of E. striata, E. quadrilineata and E. nidulans var. acristata, dethiosecoemestrin (3) [5], aurantioemestrin (4) [6] and violaceic acid (5) [5], which was originally isolated from E. violacea [7], were isolated. It was confirmed that 4 was degradated to 3 and finally to 5 [6]. In the course of a search for a key intermediate for 1 to 4, the new compound (6) positive to silver nitrate reagent was detected on TLC along with 3 and 5 from culture filtrates of E. foveolata, whereas 1 was isolated from the mycelium. The amount of 6 increased during the first week and then decreased slowly. This compound, designated secoemestrin C, was isolated along with 5 and 2 from the culture filtrate of the early state (6 days) on cultivating this fungus. The paper deals mainly the structural elucidation of 6.

RESULTS AND DISCUSSION

The [M]⁺ of secoemestrin C (6) was not detected on EI mass spectrometry, but its molecular formula

was confirmed as $C_{27}H_{22}N_2O_{10}S_4$ by high resolution positive FAB mass spectrometry at m/z 663 [M+H]⁺ and negative FAB mass spectrometry at m/z 661 [M-H]⁻. The highest peak at m/z 534, which corresponded with $C_{27}H_{22}N_2O_{10}$, in the EI mass spectrum of 6, was assigned as the partial structure lacking four sulphur atoms.

The ¹H NMR signals at δ 8.14, 8.07 and 7.08, and 7.60, 7.56, and 7.13 for 6 in CDCl₃ (Table 1) confirmed the presence of two 1,2,4-trisubstituted benzene moieties. In addition, signals for one methoxyl (δ 3.94) and one aldehyde (δ 9.78) group were observed. These signals and the strong peak at m/z 288 (C₁₅H₁₂O₆) in the EI mass spectrum showed the presence of a violaceic acid (5) moiety in 6, as in dethisecoemestrin (3) [5] and aurantioemestrin (4) [6]. Six ¹H NMR signals at δ 6.86, 6.32, 5.48, 5.33, 4.90 and 4.89 were similar to those observed in the dihydrooxepine and neighbouring five-membered rings of emestrin (1) and emestrin B (2). These coupling patterns were confirmed as the same as in 1 and 2 by ¹H-¹H decoupling experiments. The remaining ¹H NMR signals were observed at δ 3.04 (3H) and 5.24 (1H). The former was assigned to an N-methyl group, whereas the latter should be the proton attached to the carbon bearing the carbonyl carbon and the amide nitrogen, as in dithiosivatin (7) (δ 5.33) [8], which was isolated from Aspergillus silvaticus. The above results suggested that the structure of secoemestrin C is an epitetrathiodioxopiperazine derivative as depicted in 6.

Since 6 was slowly degraded in CDCl₃ solution, the ¹³C NMR, ¹H-detected heteronuclear multiple-quantum coherence via direct coupling (HMQC), and

^{*} Author to whom correspondence should be addressed.

124 M. Ooike *et al.*

heteronuclear multiple bond connectivity by 2D multiple quantum NMR (HMB C) spectra were taken using Me₂CO-d₆ as solvent. The relative structure of secoemestrin C (6) was confirmed by means of HMBC correlations (Fig. 1) and decoupling experiments.

Comparison of the CD curve of 6 [230 (negative) and 326 (negative) nm] with those of emestrin (1) [233 (negative), 266 (positive), and 338 (negative) nm] and emethallicin B (8) [236 (negative), 272 (positive) and 300 (negative) nm] [9], isolated from *E. hetero-thallica*,

Table 1. ¹ H and ¹³ C NMR chemical shifts of secoemestrin C (6)) and related compounds in CD	Cl_{3}
---------------------------------------------------------------------------------------	-------------------------------	----------

Carbon no.	6 ^(a)		6	1 ^(b)		3		4	
	$\delta_{ m C}$	$\delta_{\scriptscriptstyle \mathrm{H}}$	$\delta_{ extsf{H}}$	$\delta_{ m C}$	$\delta_{ ext{H}}$	δ_{C}	δ_{H}	$\delta_{ m C}$	$\delta_{\scriptscriptstyle \mathrm{H}}$
1	166.7			164.3		155.9		154.1	
2-NMe	32.3	2.98	3.04	27.3	3.26	27.3	3.29	34.7	3.67
3	68.6	5.43	5.24	75.6		157.5		187.1	
4	164.3			160.6		149.2		148.2	
5a	60.2	5.35	5.48	60.0	5.67	63.8	5.53	64.3	5.61
6	72.7	5.28	5.33	72.9	4.67	69.4	5.39	69.5	5.45
7	108.5	4.93	4.89	107.2	4.91	107.8	4.97	107.9	5.02
8	140.0	6.43	6.32	137.2	6.41	140.2	6.39	140.1	6.42
10	142.3	7.03	6.86	141.7	7.06	141.2	7.02	141.4	7.06
10a	113.2			112.4		118.9		119.3	
11	78.6	4.95	4.90	74.8	5.47	122.2	6.86	122.2	6.87
11-OH		5.38			6.25				
11a	81.1			81.0		131.1		131.4	
1'	123.8			122.2		122.2		122.3	
2'	122.9	7.82	8.14	124.0	7.38	122.2	$7.74^{(d)}$	122.8	7.83 ^(d)
3′	145.4			143.4 ^(c)		144.3		144.0	
4'	156.2			148.9		155.2		155.2	
4'-OMe	56.7	3.93	3.94	55.8	3.94	56.2	3.90	56.2	3.92
5′	113.2	7.27	7.08	115.4	7.21	112.2	7.02 ^(e)	112.2	$7.05^{(e)}$
6′	129.0	8.03	8.07	124.9	7.58	128.6	7.94 ^(f)	128.7	$7.99^{(f)}$
7′	165.8			164.4		165.2		165.2	
1"	146.5			145.3 ^(c)		145.1		145.4	
2"	154.6			153.3		153.3		153.1	
2"-OH		9.18			9.74		10.73		
3"	117.7	7.17	7.13	112.3	6.88	117.2	7.14 ^(e)	116.9	7.16 ^(e)
4"	128.5	7.61	7.56	122.7	7.17	127.6	$7.57^{(f)}$	127.6	$7.58^{(f)}$
5"	130.8			127.1		130.1		130.1	
6"	118.5	7.34	7.60	120.4	7.77	118.7	7.41 ^(d)	118.1	7.43 ^(d)
7"	190.9	9.78	9.78	74.8	4.97	190.5	9.79	190.5	9.80
7"-OH					5.99				

⁽a) Measured in (CD₃)₂CO.

indicated that these three compounds had the same configuration about the epipolythiodioxo-piperazine ring [10]. The positive Cotton effect at ca 270 nm in 6 would disappear because of the large negative Cotton effect at 263 nm (attributed to the benzene moiety). Thus, the structure of secoemestrin C, including the absolute stereochemistry, was confirmed as 6.

Secoemestrin C is the first example of an epitetrathiodioxopiperazine derivative isolated from an emestrin (1)-producing fungus. The dioxopiperazinethione derivatives, aurantioemestrin (4) and silvathione [8], have been isolated previously from *E. striata* and *A. silvaticus*, respectively. We mentioned that these compounds might be the intermediate from epipolythiodioxopiperazine derivatives, like 1, to trioxopiperazines, like dethiosecoemestrin (3) [6]. Degradation from 4 to 5 via 3 was recognized in ethanol solutions of 4. It is noteworthy that 6 was isolated not from the mycelium but from the culture broth, along with emestrin B (2); the amount of 6 was maximal after 6 days cultivation of *E. foveolata*. Thus, secoemestrin C (6) may be the key intermediate from emestrin (1) and/or emestrin B(2) to aurantioemestrin (4).

EXPERIMENTAL

General. Mps: uncorr. FAB MS: *m*-nitrobenzyl alcohol as matrix. ¹H (500 MHz) and ¹³C (125 MHz) NMR: TMS as int. standard. LPLC: glass column (300 × 10 i.d. mm) with silica gel CQ-3 (Wako). HPLC: YMC-Pack SIL-06 (300 × 10 i.d. mm). TLC: Kieselgel 60 F₂₅₄ plates (Merck); detection by spraying 5% AgNO₃ soln and then heating.

Isolation of secoemestrin C (6). Emericella foveolata Horie, strain IFO 30559 (= IFM4547), was cultivated on Czapek medium (30 l) supplemented with 0.2% yeast extract using 180 Roux flasks at 27° for 6 days. After removal of mycelia by filtration, the culture filtrate was extracted with CH₂Cl₂ at pH 3 and the organic layer was dried (Na₂SO₄) and then evapd in vacuo. The obtained extract (1.02 g) was chromatographed on silica gel with CHCl₃-Me₂CO (6:1), followed by repeated purification by LPLC with

⁽b) Measured in (CD₃)₂SO.

⁽c) Assignments may be reversed.

⁽d,e,f) Assignments reversed.

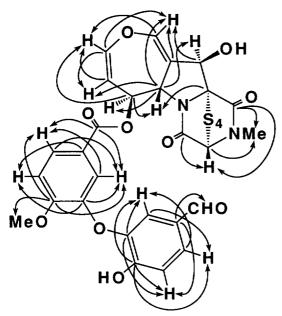


Fig. 1. Correlations in HMBC spectrum os secoemestrin C (6) Arrows indicate correlations from carbon (C_A) to proton (H_B) : $C_A \rightarrow H_B$.

CHCl₃-Me₂CO (6:1) and/or cyclohexane-Me₂CO (3:1), to obtain secoemestrin C (6) (60 mg) and emestrin B (2 mg).

Secoemestrin (6). Amorphous, mp 164–166°. [α]₂²⁰ –108.5° (Me₂CO, c 0.8). AgNO₃ test: positive (dark brown). EIMS (probe), m/z (rel. int): 534.1275 [M-S₄]⁺ (534.1275 for C₂₇H₂₂N₂O₁₀, 1.1), 505 (0.5), 288.0634 [C₁₅H₁₂O₆]⁺ (288.0634 for C₁₅H₁₂O₆, 48). Positive FAB MS m/z (rel. int.): 685 [M + Na]⁺ (0.1), 663.0240 [M + H]⁺ (663.0233 for C₂₇H₂₃N₂O₁₀S₄, 1.3), 534 [M - S₄]⁺ (4), 505 (2), 375 (4). Negative FAB-MS m/z (rel. int.): 815 [M + NBA]⁻ (0.9), 661 [M - H]⁻ (6), 597 [M - S₂ - H]⁻ (16), 565 [M - S₃ - H]⁻ (12), 533 [M - S₄ - H]⁻ (2). UV λ^{MOOH}_{max} nm (log ε): 222 (4.79), 265 (4.35), 289 sh (4.21), 344 (3.27). IR ν^{KBF}_{max} cm⁻¹: 3400 (OH), 1715 (-COO-), 1700 (-CO-), 1680, 1650

(-CON-). CD ($c 5.1 \times 10^{-5}$, MeOH) $\Delta \epsilon^{20}$ (nm): -38.2 (230), -24.0 (248), +1.7 (303), -3.2 (326) ¹H NMR: Table 1. ¹³C NMR: Table 1.

Acknowledgements—We thank Dr M. Kubo of Tsumura and Co. for FAB mass measurements. We are also grateful to Mrs T. Ogata, Mrs Y. Kawada, and Mr H. Komiya of Hoshi University for elemental analysis and NMR and mass measurements. This work was supported in part by a Grant-in-Aid for Scientific Research (no. 07672296) from the Ministry of Education Science, Sports and Culture, Japan.

REFERENCES

- 1. Seya, H., Nozawa, K., Nakajima, S., Kawai, K. and Udagawa, S., *Journal of the Chemical Society*, *Perkin Transactions* 1, 1986, 109.
- Nozawa, K., Udagawa, S., Nakajima, S. and Kawai, K., Chemical and Pharmaceutical Bulletin, 1987, 35, 3460.
- Maebayashi, Y., Horie, Y. and Yamazaki, M., Proceedings of the Japanese Association of Mycotoxicology, 1984, 20, 28.
- 4. Rahman, R. et al., Journal of Chromatography, 1970, 53, 592.
- Seya, H., Nozawa, K., Udagawa, S., Nakajima, S. and Kawai K., Chemical and Pharmaceutical Bulletin, 1986, 34, 2411.
- Kawai, K., Nozawa, K., Seya, H., Kawahara, N., Udagawa, S. and Nakajima, S., Heterocycles, 1987, 26, 475.
- 7. Yamazaki, M. et al., Chemical and Pharmaceutical Bulletin, 1982, 30, 509.
- Kawahara, N., Nozawa, K., Nakajima, S. and Kawai, K., Journal of the Chemical Society, Perkin Transactions 1, 1987, 2099.
- Kawahara, N., Nozawa, K., Yamazaki, M., Nakajima, S. and Kawai, K., Chemical and Pharmaceutical Bulletin, 1990, 38, 73.
- 11. Nagarajan, R. and Woody, R. W., Journal of the American Chemical Society, 1973, 95, 7212.