

### PII: S0031-9422(97)00312-9

## TERPENOIDS AND FLAVONOIDS FROM SALVIA CYANESCENS

GAMZE GÖKDIL, GÜLAÇTI TOPCU,\*† UFUK SÖNMEZ‡ and AYHAN ULUBELEN†‡

Faculty of Pharmacy, University of Ankara, Tandoğan, Ankara, Turkey; † TUBITAK, Marmara Research Center, Department of Chemistry, P.O. 21, 41470, Gebze, Kocaeli, Turkey; ‡ Faculty of Pharmacy, University of Istanbul, 34452, Istanbul, Turkey

(Received 2 January 1997)

Key Word Index—Salvia cyanescens; Lamiaceae; sesquiterpenes; diterpenes; triterpenes; steroids; flavonoids.

Abstract—The following compounds have been identified in the aerial parts of Salvia cyanescens: a new diterpenoid 12-isopentenyl-3-oxosalvipisone, two sesquiterpenes caryophyllene oxide, spathulenol, five diterpenes manoyl oxide,  $11\beta$ -hydroxymanoyl oxide, ferruginol, aethiopinone and salvipisone, three triterpenes  $\alpha$ -amyrin,  $\alpha$ -amyrin 3-acetate and lupeol 3-acetate as well as sitosterol and four flavonoids salvigenin, 6-hydroxyapigenin-7,4'-dimethyl ether, 6-hydroxykaempferol-3,6-dimethyl ether and kaempferol-3,7-dimethyl ether. © 1997 Elsevier Science Ltd

#### INTRODUCTION

As a part of our investigations of Turkish Salvia species we have studied the aerial parts of S. cyanescens Boiss et Ball. In addition to a new diterpenoid 12isopentenyl-3-oxo-salvipisone we have isolated two sesquiterpenes caryophyllene oxide [1], spathulenol [2], five diterpenes manoyl oxide [3],  $11\beta$ -hydroxymanoyl oxide [4], ferruginol [5], aethiopinone [6], salvipisone [7], three triterpenes  $\alpha$ -amyrin,  $\alpha$ -amyrin 3-acetate and lupeol 3-acetate together with sitosterol and four flavonoids salvigenin, 6-hydroxy-apigenin-7,4'-dimethyl ether, 6-hydroxy-kaempferol-3,6-dimethyl ether, kaempferol-3,7-dimethyl ether. The structures of the known compounds were determined by comparing their spectral data to those of literature values and by TLC comparison with authentic samples. The structure of the new compound was established by spectral data.

# RESULTS AND DISCUSSION

The HREI mass spectrum of the new compound (1) gave the molecular formula  $C_{25}H_{30}O_4$  (m/z 394.2136, calc 394.2144). The <sup>1</sup>H NMR spectrum of 1 showed typical signals of salvipisone at  $\delta$  7.96 (1H, d, J=8 Hz, H-7), 7.50 (1H, d, J=8 Hz, H-6), 4.75 (2H, br s, CH<sub>2</sub>-18), 3.37 (1H, sept. J=7 Hz, H-15), 3.16 (2H, m, CH<sub>2</sub>-1), 2.43 (3H, s, Me-20), 1.78 (3H, s, Me-19), 1.28 (6H, d, J=7 Hz, Me-16 and Me-17), and

additional signals for the isopentenyl moiety at  $\delta$  5.30 (1H, br t, J = 6.5 Hz, H-2'), 4.50 (2H, br d, J = 6.5Hz,  $H_2$ -1'), 1.80 (3H, br s), 1.72 (3H, br s) (Me-4' and Me-5'). The relationships between the protons of C-1 and C-2, as well as between H-6 and H-7, between H-1' and H-2' were deduced by spin decoupling experiments. The signal at  $\delta$  3.16 (2H, m) was typical for the C-1 protons of salvipisone which was also observed in the <sup>1</sup>H NMR spectrum of 3-oxosalvipisone [8]. Since the C-1 protons were not present in the 'H NMR spectrum of 1-oxosalvipisone [9], the location of the oxo group can not be considered at this position. If the oxo group was placed at C-2, two sets of isolated methylene doublet signals should be observed clearly, therefore the only plausible place for the oxo group was C-3. The <sup>13</sup>C NMR spectrum of 1 showed the pquinoid carbonyl signals at  $\delta$  184.5 and 183.2. The ketone at C-3 was indicated by the signal at  $\delta$  201.7. The IR and <sup>1</sup>H NMR spectrum did not show the presence of a hydroxyl group, the acetylation of 1 was unsuccessful indicating that the fourth oxygen in the molecule should be an ether function which followed from the resonance at  $\delta$  65.8 s in the <sup>13</sup>C NMR spectrum. Mass spectral fragment ions at m/z 325 [M-C<sub>5</sub>H<sub>9</sub>]<sup>+</sup> and 309 [M-C<sub>5</sub>H<sub>9</sub>O]<sup>+</sup> indicated the presence of an isopentenyl moiety in the molecule. The spectral data showed that compound 1 was 12-isopentenyl-3oxosalvipisone.

### EXPERIMENTAL

Plant material. The aerial parts of S. cyanescens were collected from central Turkey (Konya, Ermenek)

<sup>\*</sup> Author to whom correspondence should be addressed.

800 Short Reports

at an elevation 1650 m in late June 1995. The plant was identified by Dr M. Vural (Ankara), a voucher specimen is deposited in the Herbarium of the Faculty of Pharmacy, University of Ankara AEF 19485.

Extraction and isolation. Powdered plant material (1.8 kg) was extracted with Me<sub>2</sub>CO in a Soxhlet to yield 35 g of a residue. The Me<sub>2</sub>CO extract was sepd by flash chromatography using solvent mixts hexane-CH<sub>2</sub>Cl<sub>2</sub>, CH<sub>2</sub>Cl<sub>2</sub>-Me<sub>2</sub>CO and MeOH. Combined frs were further sepd on smaller flash chromatography columns and some frs were sepd on a chromatotron and purified by prep. TLC when necessary. The compounds were isolated in the following order, caryophyllene oxide (20 mg), spathulenol (15 mg), 1 (18 mg), ferruginol (12 mg), aethiopinone (10 mg), salvipisone (10 mg), manoyl oxide (20 mg), lupeyl 3acetate (20 mg), 11-hydroxymanoyl oxide (15 mg), α-amyrin 3-acetate (16 mg), salvigenin (17 mg), 6hydroxyapigenin-7,4'-dimethyl ether (5 mg), α-amyrin (12 mg), sitosterol (25 mg), 6-hydroxykaempferol-3,6dimethyl ether (8 mg), kaempferol-3,7-dimethyl ether (6 mg).

12-Isopentenyl-3-oxosalvipisone (1). UV  $\lambda_{max}^{MeOH}$  nm

(log  $\varepsilon$ ): 445 (3.5), 350 (3.2), 285 (3.4), 270 (3.4), 210 (4.2). IR  $v_{\text{max}}^{\text{CHCl}_3}$  cm<sup>-1</sup>: 3050, 2950, 2860, 1710, 1670, 1670, 1645, 1595, 1560, 1460, 1380, 1250, 1150. <sup>1</sup>H NMR (CDCl<sub>3</sub>) given in the text. <sup>13</sup>C NMR (CDCl<sub>3</sub>): C-1 38.1 t, C-2 30.2 t, C-3 201.7 s, C-4 145.5 s, C-5 133.4 s, C-6 136.3 d, C-7 125.4 d, C-8 131.5 s, C-9 126.4 s, C-10 145.5 s, C-11 184.5 s, C-12 153.2 s, C-13 126.4 s, C-14 183.2 s, C-15 25.7 d, C-16 19.8 q, C-17 20.2 q, C-18 110.2 t, C-19 22.4 q, C-20 22.4 q, C-16 65.8 t, C-2′ 123.7 d, C-3′ 142.9 s, C-4′ 24.5 q, C-5′ 19.2 q, HREIMS m/z (rel. int.): 394.2136 [M]<sup>+</sup> (4), 325 [M-69]<sup>+</sup> (25), 309 [M-85]<sup>+</sup> (40), 244 [M-C<sub>6</sub>H<sub>8</sub>O-C<sub>4</sub>H<sub>6</sub>]<sup>+</sup> (100), 229 (16), 185 (20), 141 (30), 94 (24), 83 (20), 69 (78).

Acknowledgements—This study was partly supported by the Research fund Ö-216.

#### REFERENCES

- Bisset, N. G., Diaz, M. A., Ehret, C., Quisson, G., Palmade, M., Patil, F., Pesnelle, P. and Streith, J., Phytochemistry, 1966, 5, 865.
- 2. Bowyer, R. C. and Jefferies, P. R., Chemical Industry (London), 1963, 1245.
- Anthosen, T. and Bergland, G. Acta Chemica Scandinavica, 1970, 24, 1860.
- 4. De Pascal Teresa, J., San Feliciano, A. and Miguel del Corral, Y. M., Farmaceutica Nueva, 1976, 41, 343; Chemical Abstracts, 1977, 86, 29949 c.
- 5. Cambie, R. C., Madden, R. J. and Parnell, J. C., Australian Journal of Chemistry, 1971, 24, 217.
- Boya, T. M. and Valverde, S., Phytochemistry, 1981, 20, 1367.
- 7. Rodriguez, B., Fernandez-Gadea, F. and Savona, G., *Phytochemistry*, 1984, 23, 1805.
- 8. Topcu, G., Tan, N., Ulubelen, A., Sun, D. and Watson, W. H., *Phytochemistry*, 1995, **40**, 501.
- 9. Ulubelen, A., Topcu, G. and Tan, N., *Phytochemistry*, 1992, 31, 3637.