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THREE DIBENZOYLMETHANE DERIVATIVES FROM LONCHOCARPUS SPECIES

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Key Word Index—Lonchocarpus latifolius; L. muehlbergianus; roots; Leguminosae; dibenzoylmenthanes.

Abstract—Three new dibenzoylmethane derivatives were isolated from petrol extracts of the roots of Lonchocarpus latifolius and L. muehlbergianus. Molecular structures were determined from spectroscopic data, especially NMR techniques. An alkyl group bonded to the central carbon (C-8) of a natural dibenzoylmethane, had not been found before. Spectral data analysis suggested a preponderance of the respective diketotautomers. © 1997 Elsevier Science Ltd

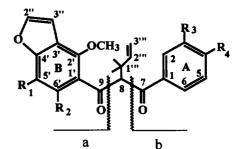
INTRODUCTION

Twenty four Brazilian species of *Lonchocarpus* are recognized from their morphological criteria. Inflorescence structure appeared as an outstanding character. According to this classification *L. muehlbergianus* belongs to the subgenus *Punctati* while *L. latifolius* is placed in the *Densiflori* section of the subgenus *Lonchocarpus*. Both species occur in southern Brazil.

Dibenzoylmethane are rarely found in nature [1]. Among 32 Lonchocarpus species previously investigated, only L. costaricensis [2] and L. subglaucescens [3] furnished such compounds, which were described as β -hydroxychalcone derivatives, because of their high degree of enolization. None of them, possessed a substituent on C-8, although dibenzoylmethane itself (4) has been reacted with allyl halide in the presence of lithium hydroxide monohydrate, furnishing the corresponding C-8 allyl derivative 5 in 92% yield [4]. Some dibenzoylmethane derivatives were synthesized in order to assay their mutagenic activities against Salmonella tiphymurium [5].

RESULTS AND DISCUSSION

The roots of each species were extracted with petrol, followed by chromatography on silica gel, allowing the isolation of several flavonoids. Compound 1 was isolated as a yellowish oil. The ¹H NMR spectrum



 $1 R_1 = R_2 = R_3 = R_4 = H$

 $2 R_1 = R_2 = H R_3 = R_4 = -O-CH_2-O-$

 $3 R_1 = R_2 = OCH_3 R_3 = R_4 = H$

4 R = H 5 R = allyl

(Table 1) showed absorptions corresponding to one free aromatic ring, an uncoupled methynic hydrogen, two *ortho*-coupled hydrogens, one furan ring, one 3,3-dimethylallyl group and one aromatic methoxyl group. The ¹³C NMR spectrum (Table 2) indicated the presence of eight C₀, eleven CH, one CH₂ and three CH₃, which were confirmed by DEPT 90 and

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Table 1. ¹H NMR spectra (300 MHz, CDCl₃) of compounds 1-3

	1	2	3
H-2	8.02 (2H, m)	7.50 (1H, d , $J = 2$)**	7.93–7.90 (2H, <i>m</i>)
H-3	7.45 (3H, m)		7.52–7.36 (3H, <i>m</i>)
H-4	7.53 (1H, m)		7.52-7.36 (3H, m)
H-5	7.45 (1H, m)	6.84 (1H, d , $J = 8)††$	7.52–7.36 (3H, <i>m</i>)
H-6	8.02~(2H, m)	7.66 (1H, dd , $J = 8$; 2)‡‡	7.93–7.90 (2H, m)
H-8	5.74 (1H, s)	5.64 (1H, s)	5.40 (1H, s)
H-5'	7.17 (1H, dd , $J = 9$; 1)*	7.19 (1H, d, J = 8; 1)*	_
H-6'	7.45(3H, m)	7.42 (1H, d, J = 8)	
H-2"	7.58 (1H, d , $J = 2$)†	7.60 (1H, d , $J = 2$)†	7.54 (1H, $d, J = 2$)†
H-3"	6.93 (1H, dd , $J = 2$; 1)‡	6.95 (1H, dd , $J = 2$; 1)‡	6.80 (1H, d , $J = 2$)†
H-2"	6.10 (1H, dd, J = 17; 11)§	6.10 (1H, dd, J = 17; 11)§	6.14 (1H, dd, J = 17; 10)§
H-3‴a	4.88 (1H, dd, J = 11; 1)	4.88 (1H, dd, J = 11; 1)	4.90 (1H, dd, J = 10; 1)
H-3‴b	4.92 (1H, dd, J = 17; 1)¶	4.92 (1H, dd, J = 17; 1)¶	4.95 (1H, dd, J = 17; 1)¶
—OCH₁	3.84 (3H, s)	3.94 (3H, s)	3.78 (3H, s)
0 011,	(, -,	, , ,	3.79(3H, s)
			4.02(3H, s)
CH ₃	1.21 (CH ₃)	1.21 (CH ₃)	1.26 (CH ₃)
,	1.23 (CH ₃)	1.23 (CH ₃)	1.28 (CH ₃)
_O_CH ₂ _O_	(3 <i>,</i>	6.04 (2H, s)	` "

Table 2. ¹³C NMR spectra (75 MHz, CDCl₃) of compounds 1–3. DEPT data (90 and 135°) in parenthesis

C DEPT	1*	2	3
1	138.7 (C ₀)	133.5 (C ₀)	135.0 (C ₀)
2	128.9 (CH)	107.7 (CH)	128.6 (CH)
3	128.4 (CH)	$148.0 (C_0)$	128.3 (CH)
4	132.7 (CH)	$151.5 (C_0)$	132.5 (CH)
5	128.4 (CH)	108.7 (CH)	128.3 (CH)
6	128.9 (CH)	125.2 (CH)	128.6 (CH)
7	195.4 (C=O)	193.3 (C==O)	194.5 (C ≔ O)
8	66.8 (CH)	66.6 (CH)	68.1 (CH)
9	197.6 (C=O)	197.6 (C ≕ O)	196.4 (C=O)
1'	$127.2 (C_0)$	$126.7 (C_0)$	$128.2 (C_0)$
2'	$152.3 (C_0)$	$152.2 (C_0)$	$138.6 (C_0)$
3′	$117.4 (C_0)$	$118.0 (C_0)$	$117.4 (C_0)$
4'	$158.8 (C_0)$	$158.7 (C_0)$	$148.9 (C_0)$
5'	106.6 (CH)	106.7 (CH)	$134.5 (C_0)$
6'	126.7 (CH)	126.8 (CH)	$144.7 (C_0)$
2"	144.7 (CH)	144.7 (CH)	144.5 (CH)
3"	105.5 (CH)	105.5 (CH)	105.2 (CH)
1‴	$41.7 (C_0)$	$41.5 (C_0)$	$41.6 (C_0)$
2""	146.1 (CH)	146.1 (CH)	146.0 (CH)
3‴a	111.7 (CH ₂)	111.5 (CH ₂)	111.4 (CH ₂)
3‴b	111.7 (CH ₂)	111.5 (CH ₂)	111.4 (CH ₂)
O-CH ₃	60.0 (CH ₃)	60.4 (CH ₃)	61.0 (CH ₃)
			61.1 (CH ₃)
			62.4 (CH ₃)
CH ₃	26.41 (CH ₃)	25.8 (CH ₃)	25.5 (CH ₃)
,	26.82 (CH ₃)	26.4 (CH ₃)	25.7 (CH ₃)
OCH ₂ O		101.8 (CH ₂)	

^{*} Assignments confirmed by HETCOR (Table 3) and COLOC (Table 4) spectra.

 $[\]begin{split} &*J_{5',6'};J_{5',3''};\dagger J_{2'',3''}; \, \sharp J_{3'',2'};\, J_{3'',5'},\\ &\S J_2''',3'''b;\, J_2''',3'''a;\, \|\, J_3'''a;2''';\, J_3'''a,b;\, \P\, J_3'''b,2''';\, J_3'''a,b.\\ &**J_{2,5};\, \dagger\dagger J_{5,6};\, \ddagger\sharp\, J_{6,5};\, J_{6,2}. \end{split}$

δ ¹ H (position)	HETCOR (attached carbon)	COSY (coupled hydrogen)
8.02 (H-2; H-6)	128.9 (C2-C6)	7.45 (H-3; H-5)
7.59 (H-2")	144.7 (C2")	6.87 (H-3")
7.45 (H-6')	126.7 (C6')	7.18 (H-5')
7.45 (H-3; H-5)	128.4 (C3; C5)	very near
7.53 (H-4)	132.7 (C4)	very near
7.18 (H-5')	106.6 (C5')	7.26–7.54 (H-6')
6.93 (H-3")	105.5 (C3")	7.59 (H-2")
		7.18 (H-5')
6.10 (H-2"')	146.1 (C2"')	4.87-4.99 (H-3"a; H-3"b)
5.74 (H-8)	66.8 (C8)	_
4.88 (H-3"'a)	111.6 (C3"')	6.10 (H-2"')
4.92 (H-3"b)	111.6 (C3"')	6.10 (H-2"')
3.84 (—OCH ₃)	60.0 (OCH ₃)	
1.21	26.4 (C—CH ₃)	
1.23	26.8 (CH ₃)	_

Table 3. Observed correlation in COSY (H-H) and HETCOR (vicinal C-H) spectra of compound 1

135°. Signals at δ 197.6 (C_0) and δ 195.4 (C_0) were assigned to dibenzoylmethane carbonyls [6, 7]. Two-dimensional spectra, such as COSY (Table 3), HETCOR (Table 3) and COLOC (Table 4), were also compatible with this assignment. The most prominent peaks in the mass spectrum could be rationalized as arising from the cleavage of α -carbonyl bonds (Table 5). These findings lead us to conclude that the 3,3-dimethylallyl group must be on C-8, in the middle of the molecule. The lack of a sharp singlet around δ 12–

Table 4. Observed long-range C_0 -H correlations (J^3) in the COLOC spectrum of compound 1 (See also fig. 2)

δ ¹³ C ₀	δ ¹ H (ppm)	Carbon assigned
197.57	7.45 (H-6′)	C-9
195.41	8.20 (H-2; H-6)	C-7
158.75	7.45 (H-6')	C-4'
152.34	3.84 (—OCH ₃)	C-2'
	7.45 (H-6')	
138.73	8.20 (H-2; H-6)	C-1
127.15	7.18 (H-5')	C-1'
117.43	7.59 (H-2")	C-3'
41.65	5.74 (H-8)	C-1‴
	4.92 (H-3"")	
	4.87 (H-3"')	
	1.23 (CH ₃)	
	1.21 (CH ₃)	

Table 5. Main fragments (a and b) in the mass spectra of compound 1-3

	m/z (rel. int.)	
Compound	a	b
1	175 (100)	105 (23)
2	175 (100)	149 (17)
3	235 (100)	105 (23)

18 (1H) in the ¹H NMR spectrum, together with the UV spectrum typical of a benzoyl-type chromophore $[\lambda_{\max}^{MeOH}(\log \epsilon) \text{ nm: } 239 \text{ (3.44)}]$ could also be taken as an evidence that the tautomeric equilibrium is highly shifted to the diketo form. In fact, much less steric hindrance is evident from the corresponding diketo molecular model.

In addition, compound 1 was recovered after being left in the presence of CH₂N₂ for 40 hr at room temperature; highly enolized dibenzoylmethanes are rapidly transformed into their methyl-enol derivatives under similar conditions [3, 8]. However, when 1 was refluxed under acidic conditions [HCl, dioxan, 40 hr), several products were detected by TLC. Preparative TLC of the crude reaction product furnished lanceolatin B (7) and pongamol (8), which could have been formed through the reaction pathway shown on Scheme 1.

Compounds 2 and 3 had ¹H NMR (Table 1) and ¹³C NMR (Table 2) spectra very similar to those of 1, except for a methylenedioxy group on A ring of 2 and two additional methoxyl groups on B ring of 3, confirmed by the most prominent peaks in their mass spectra (Table 5). In the NOE differential spectra of 1 and 2, irradiation of H-8 caused a pronounced enhancement of the H-2 and H-6 signals, but those corresponding to the hydrogens of B ring were little affected (Fig. 1). We suggest the preferential conformation where the A ring benzoyl moiety and the C-8/H-8 bond lie in the same plane, while the B ring benzoyl moiety (R_1) and 3,3-dimethylallyl group (R_2) project out of it, sustaining once more the idea that diketotautomers are the preponderant form of these compounds.

EXPERIMENTAL

Plant material. Roots of L. muehlbergianus (Hassler) were collected at the Ecological Park belong-

Scheme 1. Possible reaction pathway of compound 1 in an acidic medium under reflux.

ing to the Campinas State University (UNICAMP) in the summer of 1992. Those of *L. latifolius* (Wild.) DC were collected at the Instituto Agronômico de Campinas during the winter of 1994. Voucher samples are deposited in the University Herbarium.

Extraction and separation. Dry roots of L. latifolius (714 g) and L. muehlbergianus (932 g) were separately powdered and extracted for 60 hr in a Sohxlet apparatus with petrol. After solvent evapn, L. latifolius gave

 R_{2} R_{1} R_{1} R_{2} R_{1} R_{32} R_{1} R_{1} R_{2} R_{32} R_{1} R_{2} R_{33} R_{1} R_{2} R_{33} R_{1} R_{2} R_{33} R_{33} R_{13} R_{23} R_{33} R_{33} R_{33} R_{33} R_{34} R_{35} R_{45} R_{5}

Fig. 1. Preferential conformation of compounds 1-3 based on NOE differential spectra.

a viscous yellow oil (12.5 g) and *L. muehlbergianus*, a viscous brownish oil (9.4 g). Each extract was chromatographed on silica gel, eluting first with petrol. Eluent polarity was gradually increased by addition of CH₂Cl₂, EtOAc and MeOH to EtOAc-MeOH (1:1). By taking only part of the *L. latifolius* extract (6 g), elution with CH₂Cl₂ followed by prep. TLC in petrol-EtOAc (4:1) gave 1 (200 mg) and 2 (3.0 mg). From the *L. muehlbergianus* extract (9.9 g), elution with petrol-EtOAc (3:2) gave a complex fr. (85.5 mg) which was further submitted to prep. TLC in petrol-EtOAc (9:1), furnishing a less complex fr. (44.7 mg).

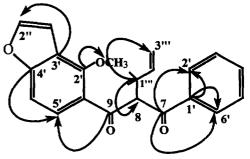


Fig. 2. Long-range C₀-H correlations in the COLOC spectrum of compound 1.

After two prep. TLC separations in petrol-EtOAc (190:1) compound 3 (11 mg) was isolated.

2'-Methoxy-[2",3",4',3']-furanodibenzoylmethane (1). Viscous yellowish oil. [α]₂₀ = +28.31 (c 0.84, CH₂Cl₂). UV λ _{max} nm (log ϵ) 239 (3.44). IR ν _{max} cm⁻¹ (film CH₂Cl₂): 3055, 2956, 2925, 1698, 1597, 1468, 1355, 1266, 1074, 738. HR-EIMS, m/z: Found 362.14418 [M]⁺ (C₂₃H₂₂O₄ requires 362.15180). GC-MS 70 eV, m/z (rel. int): 362 [M]⁺ (2), 334 (6), 319 (10), 263 (20), 175 (100), 160 (20), 105 (25), 77 (27).

3,4-Methylenedioxy-2'-methoxy-[2",3":4'4']-furano-dibenzoylmethane (2). Viscous yellowish oil. UV λ_{max} nm (log ε) 236 (3.47), 266 (3.05), 310 (2.89). IR ν_{max} cm⁻¹ (film CH₂Cl₂): 3055, 2987, 1723, 1700, 1507, 1157. EIMS 70 eV, m/z (rel. int): 406 [M]⁺ (0.4); 307 (5); 263 (10); 175 (100); 160 (17); 149 (17).

2',5',6'-Trimethoxy-[2'',3'':4',3']-furanodibenzoylmethane (3). Viscous yellowish oil. UV λ_{max} nm (log ε) 247 (3.87). IR ν_{max} cm $^{-1}$ (film CH₂Cl₂): 3386, 2936, 1709, 1594, 1478, 1423, 1347, 1265, 1133, 1108, 1067, 736, 701. EIMS 70 eV, m/z (rel. int.): 422 [M] $^+$ (7); 235 (100); 220 (11); 175 (4); 105 (11); 77 (10); 69 (5).

Transformation of 1. Compound 1 (60 mg) was refluxed in dioxane (3 ml) and conc. HCl (3 ml) under reflux for 40 hr. After usual work-up, the reaction mixt. (53.2 mg) was submitted to prep. TLC in hexane–EtOAc (17:3), furnishing the flavone 7 (15.5 mg) and the β -hydroxychalcone 8 (3.5 mg), which were identified as lanceolatin B [9] and pongamol [10], respectively, from their ¹H NMR.

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REFERENCES

- Demizu, S., Kajiyama, K., Hiraga, D., Kinoshida, K., Koyama, K., Takahashi, K., Tamura, D., Okada, K. and Kinoshida, T., Chemical and Pharmaceutical Bulletin, 1992, 40, 392.
- Waterman, G. P. and Mahmoud, E. N., Phytochemistry, 1985, 24, 571.
- 3. Magalhães, A. F., Tozzi, A. M. A. G., Salles, B. H. L. N. and Magalhães, E. G., *Phytochemistry*, 1996, **42**, 1459.
- Antonioletti, R., Bonadies, F., Orelli, L. R. and Scettri, A., Gazzeta Chimica Italiana, 1992, 122, 237.
- Choshi, T., Horimoto, S., Wang, C. Y., Nagase, H., Ichikawa, M., Sugino, E. and Hibino, S., Chemical and Pharmaceutical Bulletin, 1992, 40, 1047.
- Zeng, L., Fukai, T., Kanesi, T., Nomura, T., Zhang, R. Y. and Lou, Z. C., Heterocycles, 1994, 34, 85.
- Fukai, T., Nishizaka, J. and Nomura, T., Phytochemistry, 1994, 35, 515.
- 8. Kiuchi, F., Chen, Y. and Tsuda, Y., Chemical and Pharmaceutical Bulletin, 1990, 38, 1862.
- 9. Tanaka, T., Iinuma, M., Yuki, K., Fujii, Y. and Mizuno, M., *Phytochemistry*, 1992, 31, 534.
- Parnar, V. S., Ratthore, J. S., Jain, R., Henderson,
 D. A. and Malone, J. F., *Phytochemistry*, 1989,
 28, 591.