

DAMMARANE-TYPE TRITERPENES FROM *CORDIA SPINESCENS*

NORIO NAKAMURA, SHIHO KOJIMA, YASMINA AURA LIM, MESELY R. MESELY, MASAO HATTORI,*
MAHABIR P. GUPTA† and MIREYA CORREA‡

Research Institute for Wakan-Yaku (Traditional Sino-Japanese Medicines), Toyama Medical and Pharmaceutical University, 2630 Sugitani, Toyama 930-01, Japan; † Pharmacognosy Research Center on Panamanian Flora, College of Pharmacy, University of Panama, Panama; ‡ Herbarium of the University of Panama and Smithsonian Tropical Research Institute, Panama

(Received 13 January 1997)

Key Word Index—*Cordia spinescens*; Boraginaceae; dammarane-type triterpene.

Abstract—Two new triterpenes, 3 α ,6 β ,25-trihydroxy-20(*S*),24(*S*)-epoxydammarane and 3 α -acetoxy-6 β ,25-dihydroxy-20(*S*),24(*S*)-epoxydammarane, were isolated from the methanol extract of the leaves of *Cordia spinescens*, together with cabraleadiol. © 1997 Elsevier Science Ltd

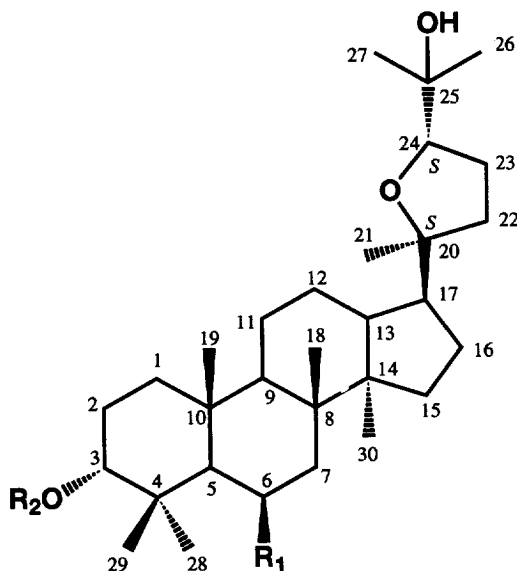
INTRODUCTION

Cordia spinescens L. is a shrub native to humid thickets and forests from western and southern Mexico to Venezuela and Peru. Infusions of the roots and leaves of this plant are used by Indians of northwestern Venezuela to relieve fever and headache [1], also the powdered stem bark is used externally for wound healing [2]. Several flavonoids, terpenoid benzoquinones and polyphenols have been isolated from the genus *Cordia* [3]. In this paper, we report on the isolation and structure elucidation of two new dammarane-type triterpenes, 3 α ,6 β ,25-trihydroxy-20(*S*),24(*S*)-epoxydammarane (1) and 3 α -acetoxy-6 β ,25-dihydroxy-20(*S*),24(*S*)-epoxydammarane (2) from *C. spinescens*.

RESULTS AND DISCUSSION

The MeOH extract of the leaves of *C. spinescens* was partitioned into hexane-, CH₂Cl₂-, EtOAc-, BuOH- and H₂O-soluble fractions. Repeated CC of the CH₂Cl₂-soluble fraction afforded three dammarane-type triterpenes (1–3). The ¹H and ¹³C NMR spectral data of 3 (Tables 1 and 2) were in good agreement with those reported for cabraleadiol [4]. The structures of 1 and 2 were determined as follows.

Compound 1 was assigned the molecular formula C₃₀H₅₂O₄ (EI-MS *m/z* 476 [M]⁺). The IR spectrum showed the presence of a hydroxyl group (3450 cm⁻¹). The ¹H NMR spectrum (Table 1) showed signals for eight tertiary methyls and three oxymethines at δ 3.37 (*br s*, H-3), 3.65 (*dd*, *J* = 10.2, 5.2 Hz, H-24) and 4.42



- 1 R₁=OH, R₂=H
2 R₁=OH, R₂=Ac
3 R₁=R₂=H

(*br s*, H-6). The ¹³C NMR spectrum (Table 2) analysed by the aid of DEPT experiment, indicated the presence of eight methyls, four sp³ quaternary carbons, and five oxygen-bearing carbons: C-6 (δ 69.4, *d*), C-25 (δ 70.2 *s*), C-3 (δ 77.6, *d*), C-24 (δ 86.3, *d*) and C-20 (δ 86.6 *s*). From the ¹H NMR spectrum, a hydroxyl group was assigned the α -configuration (*br s*) and located at C-3. The EI-mass spectrum of 1 showed a

* Author to whom correspondence should be addressed.

Table 1. ¹H NMR spectral data for compounds 1–3 (CDCl₃, 500 MHz)

H	1	2	3
H ₃ -18	1.32 (<i>s</i>)	1.33 (<i>s</i>)	0.97 (<i>s</i>)
H ₃ -19	1.23 (<i>s</i>)	1.24 (<i>s</i>)	0.86 (<i>s</i>)
H ₃ -21	1.16 (<i>s</i>)	1.17 (<i>s</i>)	1.15 (<i>s</i>)
H ₃ -26	1.12 (<i>s</i>)*	1.12 (<i>s</i>)*	1.11 (<i>s</i>)*
H ₃ -27	1.20 (<i>s</i>)*	1.20 (<i>s</i>)*	1.19 (<i>s</i>)*
H ₃ -28	1.03 (<i>s</i>)	0.94 (<i>s</i>)	0.94 (<i>s</i>)
H ₃ -29	1.23 (<i>s</i>)	1.29 (<i>s</i>)	0.84 (<i>s</i>)
H ₃ -30	0.86 (<i>s</i>)	0.90 (<i>s</i>)	0.89 (<i>s</i>)
CH ₃ CO	—	2.08 (<i>s</i>)	—
H-3	3.37 (<i>br s</i>)	4.59 (<i>br s</i>)	3.40 (<i>br s</i>)
H-6	4.42 (<i>br s</i>)	4.43 (<i>br s</i>)	—
H-24	3.65 (<i>dd</i> , <i>J</i> = 10.2, 5.2 Hz)	3.66 (<i>dd</i> , <i>J</i> = 10.2, 5.2 Hz)	3.64 (<i>dd</i> , <i>J</i> = 10.2, 5.2 Hz)

*Interchangeable values in each vertical column.

Table 2. ¹³C NMR spectral data for compounds 1–3 (CDCl₃, 125 MHz)

C	1	2	3
1	35.8 <i>t</i>	36.4 <i>t</i>	33.6 <i>t</i>
2	25.3 <i>t</i>	22.8 <i>t</i>	25.3 <i>t</i>
3	77.6 <i>d</i>	79.7 <i>d</i>	76.3 <i>d</i>
4	38.4 <i>s</i>	37.6 <i>s</i>	37.6 <i>s</i>
5	49.7 <i>d</i>	50.8 <i>d</i>	49.5 <i>d</i>
6	69.4 <i>d</i>	68.9 <i>d</i>	18.2 <i>t</i>
7	42.9 <i>t</i>	42.9 <i>t</i>	35.1 <i>t</i>
8	39.6 <i>s</i>	39.6 <i>s</i>	40.6 <i>s</i>
9	51.1 <i>d</i>	51.1 <i>d</i>	50.6 <i>d</i>
10	36.9 <i>s</i>	36.8 <i>s</i>	37.2 <i>s</i>
11	21.7 <i>t</i>	21.7 <i>t</i>	21.6 <i>t</i>
12	27.1 <i>t</i>	27.0 <i>t</i>	27.0 <i>t</i>
13	41.7 <i>d</i>	41.7 <i>d</i>	42.7 <i>d</i>
14	50.1 <i>s</i>	50.1 <i>s</i>	50.1 <i>s</i>
15	31.5 <i>t</i>	31.5 <i>t</i>	31.4 <i>t</i>
16	25.8 <i>t</i>	25.8 <i>t</i>	25.8 <i>t</i>
17	49.5 <i>d</i>	49.7 <i>d</i>	49.8 <i>d</i>
18	16.3 <i>q</i>	16.3 <i>q</i>	15.5 <i>q</i>
19	17.5 <i>q</i>	17.4 <i>q</i>	16.0 <i>q</i>
20	86.6 <i>s</i>	86.6 <i>s</i>	86.6 <i>s</i>
21	27.3 <i>q</i>	27.3 <i>q</i>	27.2 <i>q</i>
22	34.6 <i>t</i>	34.6 <i>t</i>	34.7 <i>t</i>
23	26.3 <i>t</i>	26.3 <i>t</i>	26.3 <i>t</i>
24	86.3 <i>d</i>	86.4 <i>d</i>	86.2 <i>d</i>
25	70.2 <i>s</i>	70.3 <i>s</i>	70.2 <i>s</i>
26	24.0 <i>q</i> *	24.0 <i>q</i> *	24.0 <i>q</i> *
27	27.8 <i>q</i> *	27.8 <i>q</i> *	27.9 <i>q</i> *
28	28.1 <i>q</i>	27.7 <i>q</i>	28.3 <i>q</i>
29	24.3 <i>q</i>	24.0 <i>q</i>	22.1 <i>q</i>
30	16.7 <i>q</i>	16.8 <i>q</i>	16.5 <i>q</i>
COOCH ₃		170.8 <i>s</i>	
COOCH ₃		21.4 <i>q</i>	

Multiplicity of carbon signals were verified by DEPT experiments.

*Interchangeable values in each vertical column.

characteristic fragment ion at *m/z* 143 (100%) suggesting the presence of a hydroxyisopropyl-methyl-tetrahydroxyfuran side chain [C₈H₁₅O₂]⁺ in the mol-

ecule [5, 6]. The above mentioned data were in accordance with those reported for epoxydammarane-type triterpenes [5–9]. From these observations, **1** was considered to be a hydroxy cabraleadiol [4]. The spin system, H-5 (δ 1.09)/H-6 (δ 4.42)/H-7a (δ 1.83) and H-7b (δ 1.48) seen in the ¹H ¹H COSY spectrum of **1**, and the relative downfield shifts of H₃-18, 19 and 29 (when compared with those of **3**) [4] suggested a β-hydroxyl substitution at C-6 (Table 1). This was further substantiated by NOESY experiment where spatial correlations were seen between H-6 (δ 4.42), H-5 (δ 1.09) and H₃-28 (δ 1.03).

As to the configuration at C-20 and C-24 of the side chain, both were assigned as *S* by comparing the chemical shifts of H-24 (δ 3.65, *dd*), H₃-26 (δ 1.12, *s*) and H₃-27 (δ 1.20, *s*) of **1** with those reported for 20*S* and 20*R* compounds [H₃-26 and H₃-27 of the latter compounds tend to have identical or quite similar (within 0.03 ppm) chemical shift values] [4, 8–11]. The chemical shift and coupling pattern of H-24/C-24 (δ 3.65, *dd*/δ_C 86.3) in **1** were comparable to those seen in **3** and in their C-24(*R*) epimers (δ 3.71 *t*/δ_C 83.2) such as ocotillone-II, ocotillo-II and eichlerianic acid [4] and suggested a 24*S* configuration in **1**. From the foregoing findings, compound **1** was concluded to be 3α,6β,25-trihydroxy-20(*S*),24(*S*)-epoxydammarane.

Most of the ¹H and ¹³C NMR spectra of **2** (Tables 1 and 2) were similar to those of **1** except for the presence of signals for an additional acetyl group (δ_H 2.08; δ_C 170.8 and 21.4). This was evident from the IR (1721 cm⁻¹) and the EI-MS (*m/z* 518 [M]⁺ and 459 [M—CH₃OO]⁺) spectra. The downfield shifts of H-3/C-3 in **2** (δ_H 4.59/δ_C 79.7) compared to those of **1**, suggested possible acylation of C₃—OH (Tables 1 and 2). This was unambiguously confirmed by HMQC, HMBC and NOESY experiments. Accordingly, the structure of **2** was determined to be 3α-acetoxy-6β,25-dihydroxy-20(*S*),24(*S*)-epoxydammarane.

EXPERIMENTAL

General. Mps: uncorr.; Optical rotations: 25°; IR: CHCl₃; NMR: given in δ values; EI-MS and HR-

MS: 70 eV; CC: silica gel (Kieselgel 60, 70–230 mesh, Merck), ODS (Cosmosil 140 C₁₈-OPN, Nacalai Tesque), and Florisil (60–100 and 100–200 mesh, Floridin); LiChroprep Si 60 (Size A and Size B, Merck); TLC: silica gel 60 F₂₅₄ (0.25 mm, Merck), and spots were visualized after spraying with anisaldehyde-H₂SO₄ followed by heating.

Plant material. The leaves of *Cordia spinescens* L. were collected from the Panama Canal area, Panama during July 1994 and identified by Mrs Carmen Galdames, Alex Espinoza and Eduardo Valdes. A voucher specimen is on deposit at the herbarium of the University of Panama.

Extraction and isolation of compounds. Dried leaves (1.5 kg) were extracted with MeOH (8 l × 6) at room temp., and the solvent was removed under red. pres. to give a residue (115 g). A part of the MeOH extract (89 g) was suspended in H₂O (500 ml) and shaken with solvents of increasing polarity to give hexane-soluble (24 g), CH₂Cl₂-soluble (10 g), EtOAc-soluble (6 g), BuOH-soluble (7 g) and H₂O-soluble (33 g) frs. The CH₂Cl₂-soluble fr. was subjected to CC on Florisil with increasing percentages of MeOH in CHCl₃ (0–100%) to give four frs (A–D). CC/silica gel of fr. A (C₆H₆–Me₂CO, 50:1 to 1:1) followed by MPLC/silica gel (C₆H₆–Me₂CO, 25:1) afforded **2** (15 mg) and **3** (6 mg). Repeated CC of fr. B on silica gel (C₆H₆–Me₂CO, 7:3) and ODS (70% aq. MeOH to MeOH) followed by MPLC/silica gel (C₆H₆–Me₂CO, 22:3) afforded **1** (14 mg).

3 α ,6 β ,25-Trihydroxy-20(S),24(S)-epoxydammarane (1). Yellowish oil, $[\alpha]_D -5.9^\circ$ (c 0.67, CHCl₃). IR $\nu_{\max}$ cm⁻¹: 3450; ¹H and ¹³C NMR: Tables 1 and 2; EI-MS m/z (rel. int.): 476 [M]⁺ (0.03), 440 (15), 399 (30), 381 (25), 143 [C₈H₁₅O₂]⁺ (100), 125 (72); HR-MS m/z 476.3911 [M]⁺, Calcd for C₃₀H₅₂O₄: 476.3865.

3 α -Acetoxy-6 β ,25-dihydroxy-20(S),24(S)-epoxydammarane (2). Yellowish oil, $[\alpha]_D -15.0^\circ$ (c 0.79, CHCl₃). IR $\nu_{\max}$ cm⁻¹: 3450, 1721; ¹H and ¹³C NMR: Tables 1 and 2; EI-MS m/z (rel. int.): 518 [M]⁺ (0.02), 503 [M–Me]⁺ (16), 500 [M–H₂O]⁺ (4), 459 [M–CH₃COO]⁺ (0.8), 399 (78), 381 (31), 143 [C₈H₁₅O₂]⁺ (100), 125 (58); HR-MS m/z 503.3777 [M–Me]⁺, calcd for C₃₁H₅₁O₅: 503.3736; 500.3880 [M–H₂O]⁺, calcd for C₃₂H₅₂O₄: 500.3866.

Cabraleadiol (3). Colourless needles, mp 168–174° (lit. [4] 171–173°). $[\alpha]_D +13.5^\circ$ (c 0.24, CHCl₃, (lit. [12] +13.6 (c 1, CHCl₃)). ¹H and ¹³C NMR: Tables 1 and 2; EI-MS m/z (rel. int.): 460 [M]⁺ (2.2), 445 (11), 427 (15), 401 (40), 383 (85), 191 (68), 143 [C₈H₁₅O₂]⁺ (100), 125 (85).

REFERENCES

1. Morton, J. F., in *Atlas of Medicinal Plants of Middle America*. Charles C. Thomas, Springfield, 1981, p. 719.
2. Joly, L. G., Guerra, S., Septimo, R., Solis, P. N., Correa, M. D., Gupta, M. P., Levy, S., Sandberg, F. and Perera, P., *Journal of Ethnopharmacology*, 1990, **28**, 191.
3. Marston, A., Zargorski, M. G. and Hostettmann, K., *Helvetica Chimica Acta*, 1988, **71**, 1210.
4. Hisham, A., Ajitha Bai, M. D., Fujimoto, Y., Hara, N. and Shimada, H., *Magnetic Resonance Chemistry*, 1996, **34**, 146.
5. Hasan, C. M., Islam, A., Ahmed, M., Ahmed, M. and Waterman, P. G., *Phytochemistry*, 1984, **23**, 2583.
6. Nagai, M., Tanaka, N., Tanaka, O. and Ichikawa, S., *Chemical and Pharmaceutical Bulletin*, 1973, **21**, 2061.
7. Tanaka, O. and Yahara, S., *Phytochemistry*, 1978, **17**, 1353.
8. Rao, M. M., Meshulam, H., Zelnik, R. and Lavie, D., *Tetrahedron*, 1975, **31**, 333.
9. Waterman, P. G. and Ampofo, S., *Phytochemistry*, 1985, **24**, 2925.
10. Shi, Q., Chen, K., Fujioka, T., Kashiwada, Y., Chang, J., Kozuka, M., Estes, J. R., McPhail, A. T., McPhail, D. R. and Lee, K., *Journal of Natural Products*, 1992, **55**, 1488.
11. Betancor, C., Freire, R., Hernandez, R., Suarez, E., Cortes, M., Prange, T. and Pascard, C., *Journal of the Chemical Society, Perkin Transactions I*, 1983, 1119.
12. Buckingham, J., ed., *Dictionary of Natural Products*, Vol. 2. Chapman & Hall, London, 1994, p. 2063.