



OPTIMISED ISOLATION PROCEDURE FOR BIOLOGICALLY ACTIVE COMPOUNDS NIMBOLIDE AND 28-DEOXONIMBOLIDE FROM *AZADIRACHTA INDICA* LEAVES

MANGALAM S. NAIR,* SINDHU GOPAL and DEEPA ISSAC

Regional Research Laboratory, Trivandrum-695 019, India

(Received 2 April 1997)

Key Word Index—*Azadirachta indica*; meliaceae; leaves; α -linolenic acid; nimbolide; 28-deoxonimbolide; limonoids.

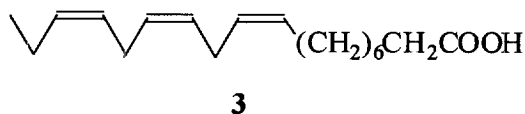
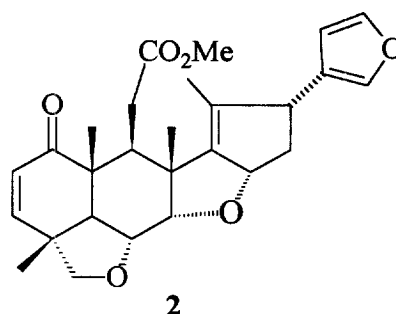
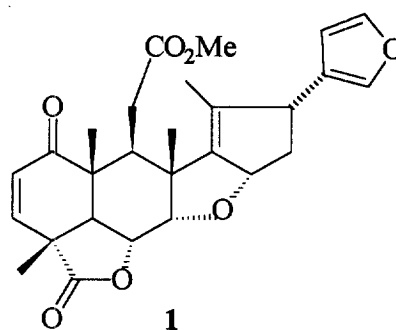
Abstract— α -linolenic acid has been isolated from fresh *A. indica* leaves and an optimised procedure for the isolation of the limonoids nimbolide and 28-deoxonimbolide is described. © 1997 Elsevier Science Ltd

INTRODUCTION

Azadirachta indica A. Juss. commonly called the 'neem tree' in India has been investigated in detail during the last three decades and a large number of biologically active compounds have been isolated from it [1]. The best known among them is azadirachtin, the structure of which was confirmed by Ley *et al.* [2]. The Indian traditional medicine 'Ayurveda' makes extensive use of all parts of *A. indica* [3]. Dry *A. indica* leaves have been traditionally used by Indian farmers to protect grains and legumes from storage pests [4]. One of the major compounds found in the leaves is nimbolide **1**, which was first isolated from fresh leaves by Ekong *et al.* [5]. Later, isolation and structural elucidation using ^1H and ^{13}C NMR data of **1** and 28-deoxonimbolide **2** was carried out by Kraus and associates [6]. A recent report by Rochanakij *et al.* [7] showed that **1** possessed an antimalarial effect.

Extensive studies conducted at RRL, Trivandrum, a few years ago, established the efficacy of neem leaf extract as an antifeedant for the control of two pests, viz. *Longitarsus nigripennis* (Coleoptera, a pest of black pepper commonly called 'Pollu beetle') and *Helopeltis antonii* (Coleoptera, a pest of the cashew tree commonly called 'tea mosquito bug'). This finding was of significance as commercially available azadirachtin-containing antifeedants did not show much effect on *L. nigripennis*. Nimbolide and 28-deoxonimbolide were found to be the major components in the antifeedant fraction [8].

In view of its therapeutic and antifeedant effects, development of an extraction protocol designed for obtaining large quantities of **1** and **2** was sought. While Ekong [5] reported the isolation of **1** from fresh neem leaves using hexane as solvent, the com-



munication did not contain further details. Bokel *et al.* [6] isolated **1** and **2** from dry leaves using diethyl ether extraction followed by repeated chromatographic separations including reversed-phase chromatography.

After extensive studies using fresh and dry leaves with various solvents, we have been able to identify the best conditions for obtaining **1** and **2**. These studies have also led to the isolation of the nutrient, α -linolenic acid **3** from fresh *A. indica* leaves. Dry leaf extract did not contain α -linolenic acid.

RESULTS AND DISCUSSION

Thin layer chromatography of dry *A. indica* leaf extract with hexane indicated the presence of a large number of compounds compared to that of fresh leaves. Extraction of *A. indica* leaves shade-dried for 48 hr (temp. $\approx 30^\circ$, average loss of water 18–20%) with hexane was found to be most suitable in terms of total yield, ease of separation and best yields of **1**, **2** and **3**. After chromatographic removal of very low polarity compounds and carotenes, α -linolenic acid and palmitic acid were obtained as a mixture. Separation was carried out by conversion to the methyl ester followed by chromatography on AgNO_3 -impregnated silica gel. Further elution provided nimbolide and 28-deoxonimbolide which were identified by comparison of mp, mass and NMR spectra.

EXPERIMENTAL

General. All mps are uncorrected and were determined on a Melttemp II hot stage melting point apparatus. ^1H (90 MHz) and ^{13}C (22.4 MHz) NMR spectra were recorded using tetramethylsilane as an int. standard.

Extraction and isolation. 545 g of finely cut leaves of *A. indica* shade-dried for 48 hr was extracted with hexane in a Soxhlet apparatus for 24 hr. Hexane was removed under red. pres. to yield 6.63 g of crude extract which was subjected to CC on silica gel (270 g). Elution with hexane removed fats and carotenes. Further elution with 10% EtOAc–hexane gave 1.1 g of a mixt. of α -linolenic acid and palmitic acid. Elution

with 25% EtOAc–hexane provided 0.716 g of nimbolide {crystallised from CH_2Cl_2 –hexane mixt., mp = 245° (lit. [6] mp = 245 – 247°), MS, ^1H and ^{13}C NMR data identical with the reported values} and 0.525 g of 28-deoxonimbolide (mp = 170° , ^1H and ^{13}C NMR data identical with reported values [6]).

The acid mixt. was treated with MeOH and *p*-toluenesulphonic acid and stirred at room temp. for 16 hr. MeOH was removed under red. pres. and the product extracted with CH_2Cl_2 . Removal of solvent followed by CC on AgNO_3 -impregnated silica gel provided 0.225 g of methyl palmitate and 0.711 g of the methyl ester of α -linolenic acid. TLC, MS, ^1H NMR spectrum were identical with that of an authentic sample of methyl linolenate obtained from Aldrich.

Acknowledgements—MSN thanks Dr A. D. Damodaran, Director, RRL for his keen interest in the programme and Prof. M. V. George for a gift of authentic methyl linolenate. Financial support from DST, India and IFS, Sweden are gratefully acknowledged.

REFERENCES

1. Devakumar, C. and Sukh, Dev, in *Neem Research and Development*, No. 3, ed. N. S. Randhava and B. S. Parmar. Society for Pesticide Science, India, 1993.
2. Bilton, J. N., Broughton, H. B., Jones, P. S., Ley, S. V., Lidert, Z., Morgan, E. D., Rzepa, H. S., Sheppard, R. N., Slawin, A. M. Z. and Williams, D. J., *Tetrahedron*, 1987, **43**, 2805.
3. Govindachari, T. R., *Current Science*, 1992, **63**, 117.
4. Manjunath, B. L. ed., *The Wealth of India, A dictionary of Indian Raw Materials and Industrial Products*, Council of Scientific and Industrial Research, New Delhi, India, Vol. 1. 1948, p. 140.
5. Ekong, D. E. U., *Journal of Chemical Society Chemical Communication*, 1967, 808.
6. Bokel, M., Cramer, R., Gutzeit, H., Reeb, S. and Kraus, W., *Phytochemistry*, 1990, **46**, 775.
7. Rochanakij, S., Thebtarononth, Y., Yenjai, Ch., and Yutharang, Y., *Southeast Asian Journal of Tropical Medicine and Public Health*, 1985, **16**, 16.
8. Babu, P. B. S., Nair, M. S., Ramesha, S. and Nair, V., Indian Pat. Appl. No. 1278/DEL/92.