

PII: S0031-9422(97)00576-1

16β,17-DIHYDROXYAPHIDICOLAN-18-OIC ACID, A MINOR DITERPENOID METABOLITE OF *CEPHALOSPORIUM APHIDICOLA*

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(Received in revised form 28 April 1997)

Key Word Index—Cephalosporium aphidicola; fungus; diterpenoid; aphidicolin.

Abstract—The structure of 16β , 17-dihydroxyaphidicolan-18-oic acid, a minor metabolite of *Cephalosporium aphidicola*, has been established by X-ray crystallography. © 1997 Published by Elsevier Science Ltd. All rights reserved

INTRODUCTION

The tumour inhibitory diterpenoid, aphidicolin (1) is produced by the fungus, Cephalosporium aphidicola [1]. During the preparation of quantities of aphidicolin for chemical transformations, we have isolated a number of minor metabolites related to aphidicolin [2, 3]. These included the acid, 16β ,18-dihydroxyaphidicolan-17-oic acid (2) [3]. We have now isolated an isomeric acid by chromatography of some aphidicolin residues.

RESULTS AND DISCUSSION

The new metabolite 3, C₂₀H₃₂O₄, had spectroscopic properties which characterized the oxygen functions as a carboxylic acid, a tertiary alcohol and a primary alcohol [v_{max} 3583, 3400 (br), 1688 cm⁻¹; δ_{C} 182.6, CO_2H ; 74.4; C-OH; 67.8, δ_H 3.21 and 3.33, each doublets, J = 11 Hz, CH₂OH]. Methylation of the acid with caesium fluoride and methyl iodide in dimethylformamide [4] gave a separable mixture of the methyl ester 4 and the formate 5 confirming the presence of a primary alcohol and carboxylic acid in the metabolite. The formate still retained the hydroxyl absorption of the tertiary alcohol. The ¹³C NMR spectra of the acid and its derivatives (see Table 1) were similar to that of 16β , 18-dihydroxyaphidicolan-17-oic acid (2). However, in the 'H NMR spectrum there was no nuclear Overhauser enhancement (NOE) of the primary alcohol signals (δ_H 3.21 and 3.33) on irradiation of either C-Me signals ($\delta_{\rm H}$ 0.80 or 1.07). However, there were mutual enhancements of the methyl group signals (4% at δ 1.07 on irradiation at

 δ 0.80 and 3% at δ 0.80 on irradiation at δ 1.07). This led to a probable structure of 16β ,17-dihydroxyaphidicolan-18-oic acid (3) for the metabolite. However, there was also the possibility that the stereochemistry might not be identical (e.g. at C-16) with that of aphidicolin and hence the structure was confirmed by X-ray crystallography of the methyl ester (see Fig. 1).

5 R' = Me, R² = CHO

EXPERIMENTAL

General. ¹H and ¹³C NMR spectra were determined at 500 and 125 MHz, respectively; IR spectra were determined as nujol mulls. Silica for chromatography was Merck 9385.

Isolation of compound 2. Chromatography of the

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Table 1.	¹³ C NMR	signals of	compounds 3,	4,	and 5 deter-
	min	ed at 125 l	MHz in CDCl ₃		

C	3	4	5
1	38.0	37.2	37.2
2	18.9	18.3	18.3
3	35.7	33.2	33.1
4	39.9	39.6	39.6
5	41.6	39.8	39.9
6	26.6	26.1	26.0
7	27.2	26.7	26.6
8	40.5	40.8	41.0
9	49.6	48.8	48.8
10	47.6	47.6	47.5
11	33.0	32.6	32.4
12	41.6	41.5	41.3
13	31.6	31.1	31.2
14	25.3	24.7	24.4
15	28.7	29.6	28.6
16	74.4	74.5	73.1
17	67.8	67.7	69.1
18	182.6	179.8	179.6
19	17.2	16.6	16.6
20	15.2	15.0	14.9
ЭМе		51.4	51.8
ЭСНО			181.6

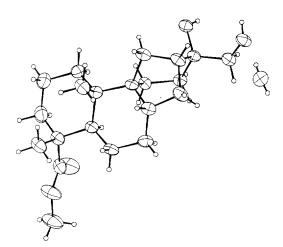


Fig. 1. X-Ray Crystal Structure of Compound 4.

residues (3 g) from the crystallization of aphidicolin (10 g) on silica gave in the frs eluted with EtOAcpetrol (4:1) 16 β ,17-dihydroxyaphidicolan-18-oic acid (210 mg) which crystallized from EtOAcpetrol as needles, mp 233.5–235°. (Found: C, 71.2; H, 9.5. C₂₀H₃₂O₄ requires C, 71.4; H, 9.6%) IR $v_{\rm max}$ cm⁻¹: 3583, 3450 (br), 1688. ¹H NMR (CDCl₃-pyridine- d_s): δ 0.80 (3H, s, H-19), 1.07 (3H, s, H-20), 3.21 and 3.33 (each 1H, d, J = 11 Hz. H-17).

Methylation with caesium fluoride and methyl iodide. The acid (160 mg) in dry DMF (4 ml) was treated with dry caesium fluoride (310 mg) and MeI (320 mg) at room temp. for 24 hr. The soln was poured into H₂O

and the product recovered in EtOAc. The extract was dried over Na2SO4 and the solvent evapd to give a gum which was chromatographed on silica. Elution with EtOAc-petrol (3:7) gave methyl 17-formyloxy- 16β -hydroxyaphidicolan-18-oate (5) (22 mg) which crystallized from EtOAc-petrol as needles, mp 143°. (Found: C, 69.9; H, 9.1. C₂₂H₃₄O₅ requires C, 69.8; H, 9.0%). IR v_{max} cm⁻¹: 3583, 1722 (br). ¹H NMR (CDCl₃): δ 0.91 (3H, s, H 19), 1.12 (3H, s, H-20), 3.58 (3H, s, OMe), 3.99 (2H, br s, H-17), 8.06 (1H, s, OCHO). Further elution with EtOAc-petrol (1:1) gave methyl 16β,17-dihydroxyaphidicolan-18-oate (4) (56 mg) which crystallized from EtOAc-petrol as needles, mp 129-131°. (Found: C, 70.2; H, 9.7. C₂₁H₃₄O₄. 0.5 H₂O requires C, 70.2; H, 9.8%) IR v_{max} cm⁻¹ 3580, 3274, 1719, ¹H NMR (CDCl₃): δ 0.99 (3H, s, H-19), 1.19 (3H, s, H-20), 3.39 and 3.42 (each 1H, d, J = 11Hz, H-17), 3.65 (3H, s, OMe).

Crystallographic data and structure determination. $C_{21}H_{34}O_4$. 0.5 H_2O , M_r 359.49, orthorhombic, space group $C222_1$ (no 20), a=7.808(4), b=10.958(7), c=46.702(12) Å, $\alpha=\beta=\gamma=90^\circ$, V=3996(3) Å³, Z=8, $D_{\rm calc}$ 1.20 g cm⁻³, F(000) 1576, monochromated Mo-K α radiation $\lambda=0.71073$ Å, $\mu=0.08$ mm⁻¹.

Data were collected using a crystal size ca $0.40 \times 0.30 \times 0.30$ mm on an Enraf-Nonius CAD4 diffractometer. A total of 1990 unique reflections were measured for $2 < \theta < 25^\circ$ and 0 < h < 9, 0 < k < 12, 0 < l < 55. 1403 Reflections with $I > 2\sigma(I)$ were used in the refinement. There was no crystal decay and no absorption correction was applied.

The structure was solved by direct methods using SHELXS 86 and SHELXL 93. The non-hydrogen atoms were refined anisotropically by full matrix least squares. Hydrogen atoms were included in riding mode with $U_{\rm iso}$ (H) = $1.2 U_{\rm eq}$ (C) or $1.5 U_{\rm eq}$ (C) for methyl groups. Hydroxyl groups were fixed at idealised geometry but with the torsion angle defining the hydrogen atom position, refined with $U_{\rm iso}$ (H) equal to $1.5 U_{\rm q}$ (O). There was a molecule of H₂O lying on a crystallographic 2-fold rotation axis for which the hydrogen atom was fully refined. The final R indices were $R_1 = 0.055$, $wR_2 = 0.138$ and R indices (all data) $R_1 = 0.085$, $wR_2 = 0.175$. The tables of crystallographic data have been deposited with the Cambridge Crystallographic Data Centre.

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