



ACYLATED KAEMPFEROL GLYCOSIDES FROM *LAURUS NOBILIS* LEAVES

CHRISTEL FIORINI, BRUNO DAVID,* ISABELLE FOURASTÉ† and JOSEPH VERCAUTEREN‡

Centre de Recherche des Substances Naturelles, Institut de Recherche Pierre Fabre BP 92, F-81603 Gaillac Cedex, France; † Laboratoire de Pharmacognosie, Faculté des Sciences Pharmaceutiques, 35, Chemin des Maraîchers F-31076 Toulouse Cedex, France; ‡ Laboratoire de Pharmacognosie, Faculté des Sciences Pharmaceutiques, Université Victor Segalen Bordeaux 2-146, rue Léo Saignat, F-33076 Bordeaux Cedex, France

(Received in revised form 9 June 1997)

IN HONOUR OF THE RETIREMENT OF PROFESSOR ANDRÉ CAVÉ

Key Word Index—*Laurus nobilis*; Lauraceae; flavonol glycosides; acylated kaempferol glycosides.

Abstract—*Laurus nobilis* leaves yielded four non-polar flavonoids: kaempferol-3-*O*- α -L-(3'',4''-di-*E*-*p*-coumaroyl)-rhamnoside, kaempferol-3-*O*- α -L-(2'',4''-di-*E*-*p*-coumaroyl)-rhamnoside, kaempferol-3-*O*- α -L-(2''-*E*-*p*-coumaroyl)-rhamnoside and a new product kaempferol-3-*O*- α -L-(2'',4''-di-*Z*-*p*-coumaroyl)-rhamnoside. Structural elucidation was achieved by UV, 1D- and 2D-NMR experiments, mass spectrometry, acid hydrolysis and saponification. © 1998 Elsevier Science Ltd. All rights reserved

INTRODUCTION

Apollo's Laurel (*Laurus nobilis* L.), a common Mediterranean aromatic plant, has been analysed for alkaloids [1] and essential oils of its leaves [2–5]. Its leaves also contain glycosylated flavonoids (derived from quercetin and kaempferol), (–)-epicatechin, (+)-catechin, (+)-gallocatechin, (+)-epigallocatechin and procyanidins (B2, B4, B5 and B7) [6]. The present study reports on the isolation of four non-polar flavonoids from leaves [7].

RESULTS AND DISCUSSION

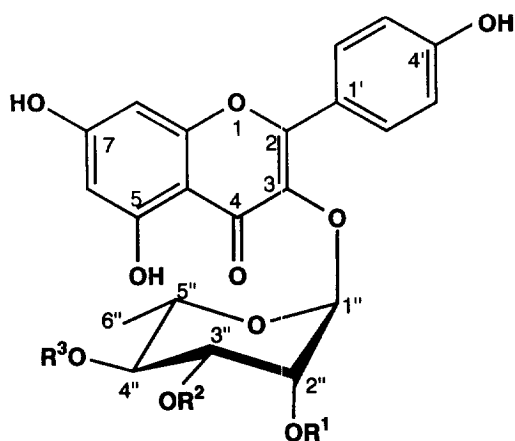
Flavonoids **1–4** were isolated from a MeOH–H₂O (7:3) extract of dried leaves. They were identified by 1D- and 2D-NMR experiments (¹H NMR, ¹³C J-MOD, ¹H-¹H COSY, HMQC and HMBC), FAB-mass spectrometry and by chemical correlations (acid hydrolysis and saponification).

Flavonoids **1**, **2** and **3** all possessed a molecular mass of 724 and flavonoid **4** of 578. Acid hydrolysis (2M H₂SO₄) indicated the aglycone of these heterosides to be kaempferol and saponification proved the presence of rhamnosyl and *p*-coumaroyl moieties.

¹H and ¹³C NMR experiments (Tables 1 and 2) showed that the differences between these kaempferol-3-*O*- α -L-rhamnoside derivatives were due to the number, the position and configuration (*E* or *Z*) of the *p*-coumaroyl residues on the rhamnose moiety. The ¹H NMR spectrum and the ¹H-¹H COSY experiment allowed us to assign accurately all of the glycosidic protons and to determine the positions of acylation by the *p*-coumaroyl residues from the downfield chemical shift of the corresponding proton(s). All the heteronuclear correlations in the HMBC spectrum confirmed these assignments. The double bond *Z* configuration was deduced from the lowest value (12.8 Hz) of the coupling constants of the ethylenic protons and the *E* configuration from the largest one (16.0 Hz). The HMBC correlation between C-3 (δ 135.1) and the anomeric (H-1'') proton (δ 5.53) proved the position of rhamnosyl substitution to be on the 3-hydroxyl group. Finally, the ¹³C and ¹H NMR signals were fully assigned to the four acylated flavonoids, which are thus reported to be: kaempferol-3-*O*- α -L-(3'',4''-di-*E*-*p*-coumaroyl)-rhamnoside (**1**), kaempferol-3-*O*- α -L-(2'',4''-di-*E*-*p*-coumaroyl)-rhamnoside (**2**), kaempferol-3-*O*- α -L-(2'',4''-di-*Z*-*p*-coumaroyl)-rhamnoside (**3**) and kaempferol-3-*O*- α -L-(2''-*E*-*p*-coumaroyl)-rhamnoside (**4**).

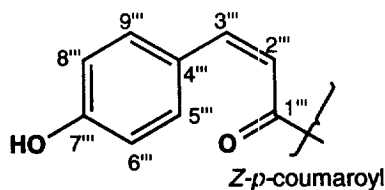
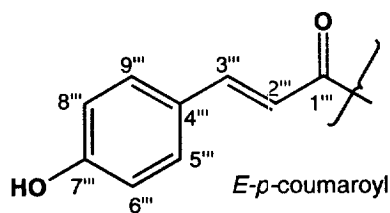
Monoacylated flavonoids are quite usual but diacylation with aromatic acids is not a frequent structural

* Author to whom correspondence should be addressed.



1 $R^1=H$, $R^2=R^3= E\text{-}p\text{-coumaroyl}$

2 $R^2=H$, $R^1=R^3= E\text{-}p\text{-coumaroyl}$



3 $R^2=H$, $R^1=R^3= Z\text{-}p\text{-coumaroyl}$

4 $R^2=R^3=H$, $R^1= E\text{-}p\text{-coumaroyl}$

feature. Flavonoids **1** and **2** have recently been isolated from *Ocotea vellosiana*, belonging also to the Lauraceae family [8] and **4** from *Platanus acerifolia* (Platanaceae) [9]. Compound **3** is new and also the first di-*Z*-cinnamoyl flavonoid rhamnoside reported to date in the plant kingdom.

EXPERIMENTAL

General. UV: usual shifts reagents used according to standard procedures [9]. FAB-MS: positive mode, glycerol; NMR: 500 (^1H) and 125 (^{13}C) MHz in CD_3OD or $\text{DMSO}-d_6$.

Plant material. Leaves were collected in March 1995 from trees growing in Manas (south-west of France) and were identified by Prof. I. Fourasté.

Extraction and isolation. Dried leaves (3 kg) were extracted at reflux temp. with 2×12 l of H_2O -MeOH (3:7). The extracts were bulked and concd under red.

press. The water-insoluble green ppt. (122 g) was purified by silica gel filtration. Elution was carried out successively with 3×0.5 l hexane, 3×0.5 l CHCl_3 and 3×0.5 l Et_2O . The Et_2O eluates (8.1 g) containing apolar flavonoids were subjected to CC (Büchi 19014, 46 cm $\times$ 3.6 cm) on silica gel 60 (Lichroprep 15–25 μm) eluted with CH_2Cl_2 -MeOH (19:1). Six frs were selected based on TLC analysis of the initial frs (100 ml): F1 (0.2 g), F2 (2.4 g), F3 (0.8 g), F4 (3.4 g), F5 (0.3 g), F6 (0.8 g). Compound **1** (60 mg) crystallized from F2 in CH_2Cl_2 -MeOH (19:1) at room temp.

Flavonoids **2**, **3**, and **4**, respectively, were isolated from F3, F4 and F5 which were purified by prep. CC on Lichroprep (15–25 μm) eluted with CH_2Cl_2 -MeOH (19:1). Final purification was achieved by prep. HPLC (Nucleosil C-18, 5 μm , 20 $\times$ 250 mm); H_2O -MeOH (8:17) affording 17 mg of **2**, 62 mg of **3** and 10 mg of **4**.

Structural elucidation. Acid hydrolyses were carried out by dissolving 2 mg of the appropriate compound in 50 μl MeOH and 50 μl 2 M H_2SO_4 . After 20 min at 100°, MeOH evapn and addition of 1 ml H_2O , the aq. phase was extracted with 3×250 μl Et_2O . The aq. phase was neutralized with Ba_2CO_3 and analysed by HPLC: Nucleosil NH₂, 5 μm , 250 mm $\times$ 4.6 mm; MeCN- H_2O (83:17). Sugars were detected by light diffusion detector (Eurosep).

Saponifications were carried out by dissolving 2 mg of flavonoid in 100 μl 1% NaOH. After 5 min at 100°, the mixts were neutralized with 1 N HCl. The aq. phase was extracted with 3×50 μl Et_2O and analysed by TLC: silica gel, toluene- Et_2O -10% HOAc (1:1:3), upper phase.

Compound 1 (kaempferol-3-*O*- α -L-(3'',4''-di-*E*-*p*-coumaroyl)-rhamnoside). Yellow powder. Alkaline hydrolysis gave kaempferol-3-*O*- α -L-rhamnoside, and *p*-coumaric acid. UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 267, 300 sh, 313; (AlCl_3) 280, 312, 322 sh, 400 sh; ($\text{AlCl}_3 + \text{HCl}$) 278 sh, 307, 327 sh, 395 sh; (NaOAc) 274, 317, 325 sh, 368; ($\text{NaOAc} + \text{H}_3\text{BO}_3$) 268, 303 sh, 317. FAB-MS: m/z 725 $[\text{M} + \text{H}]^+$.

Compound 2 (kaempferol-3-*O*- α -L-(2'',4''-di-*E*-*p*-coumaroyl)-rhamnoside). Yellow powder. Alkaline hydrolysis gave kaempferol-3-*O*- α -L-rhamnoside and *p*-coumaric acid. UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 268, 300 sh, 313; (AlCl_3) 275, 315 sh, 395 sh; ($\text{AlCl}_3 + \text{HCl}$) 275, 300 sh, 317, 400 sh; (NaOAc) 275, 320 sh, 377; ($\text{NaOAc} + \text{H}_3\text{BO}_3$) 265, 301 sh, 318. FAB-MS: m/z 725 $[\text{M} + \text{H}]^+$.

Compound 3 (kaempferol-3-*O*- α -L-(2'',4''-di-*Z*-*p*-coumaroyl)-rhamnoside). Yellow powder. Alkaline hydrolysis gave kaempferol-3-*O*- α -L-rhamnoside and *p*-coumaric acid. UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 266, 301 sh, 313; (AlCl_3) 276, 307, 320, 390 sh; ($\text{AlCl}_3 + \text{HCl}$) 278, 308, 323 sh, 392 sh; (NaOAc) 274, 317, 325 sh, 368; ($\text{NaOAc} + \text{H}_3\text{BO}_3$) 268, 303 sh, 317. FAB-MS: m/z 725 $[\text{M} + \text{H}]^+$.

Compound 4 (kaempferol-3-*O*- α -L-(2''-*E*-*p*-coumaroyl)-rhamnoside). Yellow powder. Alkaline hydrolysis gave kaempferol-3-*O*- α -L-rhamnoside and *p*-coum-

Table 1. ¹H NMR data of compounds 1–4*

H	1	2	3	4
6	6.24 <i>d</i> (1.98)	6.18 <i>d</i> (1.75)	6.18 <i>d</i> (1.75)	6.18 <i>d</i> (2.1)
8	6.41 <i>d</i> (1.98)	6.38 <i>br s</i>	6.34 <i>br s</i>	6.33 <i>d</i> (2.1)
2',6'	7.86 <i>d</i> (8.73)	7.77 <i>d</i> (8.7)	7.74 <i>d</i> (8.8)	7.78 <i>d</i> (8.8)
3',5'	7.07 <i>d</i> (8.7)	7.00 <i>d</i> (8.7)	6.97 <i>d</i> (8.7)	6.94 <i>d</i> (8.8)
1''	5.8 <i>d</i> (1.32)	5.58 <i>br s</i>	5.53 <i>br s</i>	5.47 <i>d</i> (1.6)
2''	4.44 <i>m</i>	5.47 <i>br s</i>	5.52 <i>br s</i>	5.52 <i>dd</i> (1.6; 3.4)
3''	5.40 <i>dd</i> (3.15; 10.05)	4.00 <i>dd</i> (3.1; 9.7)	4.13 <i>dd</i> (3.4; 9.8)	3.93 <i>dd</i> (3.4; 9)
4''	5.21 <i>t</i> (10.05)	4.82 <i>t</i> (9.7)	4.86 <i>t</i> (9.8)	3.39 <i>t</i> (9)
5''	3.3 <i>m</i>	3.2 <i>m</i>	3.44 <i>m</i>	3.39 <i>m</i>
6''	0.87 (6.27)	0.76 <i>d</i> (6.15)	0.83 <i>d</i> (6.2)	0.98 <i>d</i> (5.83)
2'''	6.2 (16.02)	6.31 <i>d</i> (15.9)	5.74 <i>d</i> (12.8)	6.33 <i>d</i> (15.9)
2'''	6.31 <i>d</i> (16.02)	6.44 <i>d</i> (15.9)	5.84 <i>d</i> (12.8)	
3'''	7.52 <i>d</i> (16.02)	7.48 <i>d</i> (15.9)	6.86 <i>d</i> (12.8)	7.62 <i>d</i> (15.9)
3'''	7.65 <i>d</i> (16.02)	7.60 <i>d</i> (15.9)	6.89 <i>d</i> (12.8)	
5''' ,9'''	7.42 <i>d</i> (8.58)	7.61 <i>d</i> (8.7)	7.64 <i>d</i> (8.6)	7.44 <i>d</i> (8.7)
5''' ,9'''	7.45 <i>d</i> (8.58)	7.59 <i>d</i> (8.7)	7.66 <i>d</i> (8.6)	
6''' ,8'''	6.78 <i>d</i> (8.58)	6.82 <i>d</i> (8.7)	6.75 <i>d</i> (8.6)	6.78 <i>d</i> (8.7)
6''' ,8'''	6.88 <i>d</i> (8.58)	6.79 <i>d</i> (8.7)	6.72 <i>d</i> (8.6)	

* At 500 MHz in CD₃OD except for compound 1 which was dissolved in DMSO-*d*₆.
J (Hz) in parentheses.

Table 2. ¹³C NMR data of compounds 1–4*

C	1	2	3	4
2	157.0	157.0	159.3	159.2
3	133.1	132.6	135.1	135.6
4	177.1	176.8	179.2	179.4
5	160.9	160.9	163.1	163.2
6	98.8	97.5	99.8	100.0
7	165.4	165.5	166.6	166.3
8	93.7	93.8	95.0	94.9
9	156.4	156.4	158.6	158.6
10	103.6	103.3	105.7	105.8
1'	120.0	119.9	122.4	122.5
2'	130.2	130.3	131.8	131.8
3'	115.5	115.2	116.7	116.7
4'	159.7	160.1	161.8	161.7
5'	115.5	115.2	116.7	116.7
6'	130.2	130.3	131.8	131.8
1''	100.4	98.9	101.1	100.5
2''	67.8	70.9	72.9	73.4
3''	70.6	65.7	68.5	70.7
4''	69.4	72.4	74.3	73.6
5''	67.1	67.7	69.8	72.1
6''	16.7	16.7	17.6	17.8
1''' ,1'''	165.7	165.6	167.0; 167.4	168.3
2''' ,2'''	113.0; 113.4	113.34; 113.7	115.8	114.9
3''' ,3'''	145.1	144.4; 144.6	145.9	147.1
4''' ,4'''	124.6; 124.5	124.7	127.5	127.2
5''' ,9''' ; 5''' ,9'''	130.1	130.28; 130.1	133.9	131.3
6''' ,8''' ; 6''' ,8'''	115.5	115.51	116.1	116.8
7''' ,7'''	159.7	159.8; 159.7	160.1	161.3

* At 500 MHz in DMSO-*d*₆ for 1 and CD₃OD for 2, 3 and 4.

aric acid. UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 266, 293 sh, 314; (AlCl₃) 275, 302, 322, 393 sh; (AlCl₃+HCl) 273, 303 sh, 323, 395 sh; (NaOAc) 275, 323 sh, 372; (NaOAc+H₃BO₃) 265, 303 sh, 314. FAB-MS: 578 *m/z* [M+H]⁺.

REFERENCES

1. Pech, B. and Bruneton, J., *Journal of Natural Products*, 1982, **45**, 560.
2. Zola, A., Le Vanda, J. P. and Guthbrod, F., *Plantes Médicinales et Phytothérapie*, 1977, **11**, 241.
3. Riaz, M., Ashraf, C. M. and Chaudhary, F. M., *Pakistani Journal of Scientific Research*, 1989, **32**, 33.
4. Zheng, K. L., Ying, F. H., Guo, P. G. and Yu, H. G., *Acta Botanica Sinica*, 1990, **32**, 878.
5. Knackstedt, J. and Herrman, K., *Zeitschrift für Lebens Untersuchung Forschung*, 1981, **173**, 288.
6. Sakar, M. K. and Engelshewe, K., *Zeitschrift für Lebens Untersuchung Forschung*, 1985, **180**, 494.
7. Fiorini, C., Ph.D. Thesis, Institut National Polytechnique, Toulouse, 1996.
8. Garcez, W. S., Yoshida, M. and Gottlieb, O. R., *Phytochemistry*, 1995, **39**, 815.
9. Kouadji, M., Morand, J. M. and Garcia, J., *Journal of Natural Products*, 1993, **56**, 1618.