



TETRANORTRITERPENOIDS FROM *KHAYA SENEGALENSIS*

T. R. GOVINDACHARI* and G. N. KRISHNA KUMARI

Centre for Agrochemical Research, SPIC Science Foundation, 110 Mount Road, Madras 600 032, India

(Received in revised form 18 June 1997)

Key Word Index—*Khaya senegalensis*; Meliaceae; Tetranortriterpenoids; Mexicanolide; Khivorin; Prep. HPLC.

Abstract—Preparative HPLC of an ethanol extract of seeds of *Khaya senegalensis* yielded three new tetranortriterpenoids of the mexicanolide type. These compounds were identified as 2-hydroxymexicanolide, 6-deoxydestigloylswietenine and 2,3-dihydroxy-3-deoxymexicanolide. In addition, mexicanolide, 3 β -hydroxy-3-deoxymexicanolide, 3 β -hydroxy-3-deoxycarapin, 6-hydroxy methyl angolensate, 3-acetyl-7-keto khivorin, 3-deacetyl khivorin and 3,7-dideacetyl khivorin were also isolated. © 1998 Elsevier Science Ltd. All rights reserved

INTRODUCTION

Taylor and co-workers [1, 2] have investigated extensively the genus *Khaya* A. Juss (Meliaceae) which is the main source of African mahogany. Three structural types of tetranortriterpenoids are encountered in this genus: khivorin, a D-seco compound, mexicanolide and methyl angolensate which are B-seco compounds. From *K. senegalensis*, the most distinctive species of the genus *Khaya*, fifteen compounds belonging to the above mentioned types have been isolated from various parts of the tree [3]. In a recent publication [4] three new tetranortriterpenoids were reported which are again mexicanolide derivatives.

Extensive biological work has been carried out on C-seco-tetranortriterpenoids of the type of azadirachtin [5, 6] but, very little work has been done on B-seco-tetranortriterpenoids. It was found that nymania-3, a B-seco compound [7] isolated in this laboratory [8] from *Dysoxylum malabaricum* was a very potent insect antifeedant. Since *K. senegalensis* is a good source of B-seco compounds, it was decided to isolate various compounds present in this source and examine their biological activities. Mexicanolide was the principal compound isolated from the 30–40% hexane–ethyl acetate eluent in VLC (0.6%). The more polar compounds isolated from the ethyl acetate eluate were conveniently separated by prep. HPLC. Nine compounds were isolated of which three were new [9].

RESULTS AND DISCUSSION

The three new compounds were assigned structures on the basis of extensive spectral analysis. Compound

1, C₂₇H₃₂O₈ (HRMS), showed IR peaks at 3688, 3566 cm⁻¹ (OH), 1732 cm⁻¹ (CO) and 875 cm⁻¹ (furan). The ¹H NMR spectrum (Table 1) indicated four tertiary methyls (δ 0.85, 1.0, 1.3), a β -substituted furan ring (δ 7.6, 7.43 and 6.51) and a carbomethoxy group (δ 3.75, s, 3H). The ¹³C NMR spectrum (Table 2) showed the presence of two six membered ketones (δ 212.8, 209.6) and a tetra substituted double bond (δ 135.3 s and 123.6 s). A study of the NMR data of compound **1** indicated a mexicanolide skeleton. The ¹³C NMR spectrum of compound **1** showed only two CH signals (DEPT 90) in the δ 40–60 region in comparison to mexicanolide. The presence of a singlet at δ 80.3 indicated that one of the CHs in mexicanolide was replaced by —C—OH which was in agreement with the HRMS data. In the ¹H NMR spectrum, the two protons at C-30 appeared as geminally coupled doublets at δ 2.12 and 3.35 (J = 13.3 Hz) showing the absence of a proton at C-2. Consequently the hydroxyl was placed at position C-2 and this was supported by the downfield shift of the β -carbon at C-30 compared to mexicanolide. Hence compound **1** was assigned the structure, 2 α -hydroxymexicanolide.

The NMR data of compound **2** also showed a similar skeleton. The ¹H NMR spectrum showed a sharp singlet at δ 3.64 (s, 1H) for a CHOH which was connected to a carbon at δ 85.5 (d) in the HETCOR spectrum. This indicated the presence of a secondary hydroxyl group. This suggested the replacement of a keto group by a hydroxyl group either at C-1 or C-3. The placement of the hydroxyl group at position C-3 was confirmed by COLOC. C-3 showed ³J_{CH} correlation to the gem dimethyl protons at C-4. C-1 at δ 219 showed correlation to the 19-Me. Thus compound **2** was assigned the structure 2 α ,3 β -dihydroxy-3-

* Author to which correspondence should be addressed.

Table 1. ^1H NMR spectral data of compounds **1**, **2** and **3** (200 MHz, δ values in CDCl_3)

H	1	2	3
2	—	—	3.42 <i>m</i>
3	—	3.64 <i>s</i>	3.80 <i>d</i> ($J = 9.3$ Hz)
5	2.8 <i>m</i>	3.16 <i>dd</i> ($J = 3.9, 9.4$ Hz)	3.31 <i>m</i>
6	2.5 <i>m</i> , 2H	2.36 <i>m</i>	2.40 <i>m</i>
9	2.12 <i>m</i>	1.96 <i>m</i>	2.21 <i>m</i>
11	1.8 <i>m</i>	1.81 <i>m</i>	1.75 <i>m</i>
12	1.8 <i>m</i>	1.1 <i>m</i>	1.75 <i>m</i>
	1.3 <i>m</i>	1.05 <i>m</i>	1.48 <i>m</i>
14	—	—	2.28 <i>m</i>
15	3.5 <i>m</i>	4.13 <i>d</i> ($J = 21.1$ Hz) 3.48 <i>d</i> ($J = 21.1$ Hz)	2.94 <i>m</i>
17	5.25 <i>s</i>	5.59 <i>s</i>	5.74 <i>s</i>
21	7.60 <i>s</i>	7.57 <i>s</i>	7.76 <i>s</i>
22	6.51 <i>s</i>	6.50 <i>d</i> ($J = 1.5$ Hz)	6.48 <i>s</i>
23	7.43 <i>d</i> ($J = 1.6$ Hz)	7.41 <i>d</i> ($J = 1.5$ Hz)	7.43 <i>d</i> ($J = 1.6$ Hz)
30	3.3 <i>d</i> ($J = 13.3$ Hz) 2.12 <i>d</i> ($J = 13.3$ Hz)	3.48 <i>d</i> ($J = 13.7$ Hz) 1.64 <i>d</i> ($J = 13.7$ Hz)	5.72 <i>d</i> ($J = 7.5$ Hz)
18Me	1.00 <i>s</i>	1.05 <i>s</i>	1.11 <i>s</i>
19Me	1.00 <i>s</i>	1.24 <i>s</i>	1.15 <i>s</i>
28Me	0.85 <i>s</i>	0.68 <i>s</i>	0.84 <i>s</i>
29Me	1.30 <i>s</i>	0.83 <i>s</i>	0.76 <i>s</i>
COOMe	3.75 <i>s</i>	3.71 <i>s</i>	3.72 <i>s</i>

Table 2. ^{13}C NMR spectral data of mexicanolide, compounds **1**, **2** and **3*** (50 MHz, δ values in CDCl_3)

	Mexicanolide	1	2	3
C-1	212.8	212.8	219.2	218.3
C-2	57.9	80.3	79.6	51.2
C-3	210.8	209.6	85.5	76.3
C-4	49.3	49.2	39.5	39.7
C-5	40.1	40.1	39.4	40.5
C-6	32.1	32.2	33.2	33.1
C-7	173.5	173.6	174.9	174.2
C-8	133.7	135.3	132.7	138.6
C-9	50.4	50.2	51.4	56.7
C-10	54.2	52.9	52.5	50.1
C-11	18.4	18.5	18.6	20.8
C-12	28.6	28.7	28.4	29.2
C-13	37.9	38.1	38.0	36.8
C-14	125.2	123.6	125.7	45.3
C-15	32.8	32.8	35.3	34.4
C-16	169.7	169.8	171.4	169.9
C-17	80.5	80.7	80.1	77.1
C-18	17.4	17.4	17.4	21.9
C-19	17.9	21.9	16.8	15.6
C-20	120.6	120.4	120.6	120.6
C-21	142.6	141.6	141.6	141.9
C-22	109.8	109.9	110.0	109.6
C-23	141.4	142.8	142.5	142.8
C-28	17.9	17.9	23.6	22.4
C-29	21.9	17.8	19.6	20.5
C-30	36.2	44.2	43.9	123.3
COOCH ₃	52.0	52.8	52.0	52.2

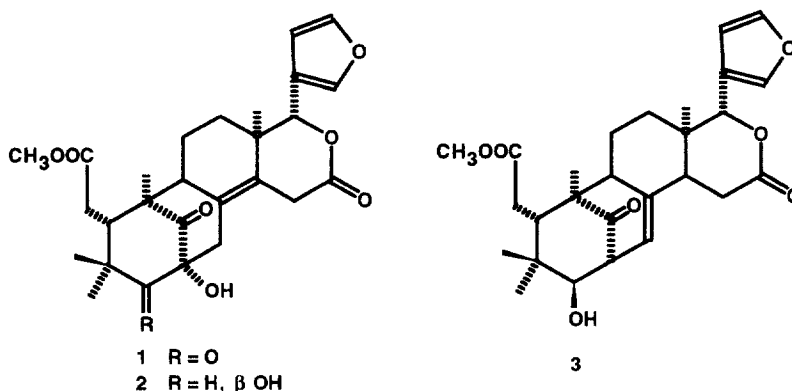
* Assignments were supported by DEPT experiments.

deoxymexicanolide. The 3-acetate of this compound was previously reported from *K. ivorensis* [10].

Compound **3** also showed the mexicanolide skeleton but with only one keto group in a six membered ring. The ^{13}C NMR spectrum showed a trisubstituted double bond (δ 136.6 *s* and 123.3 *d*). This was also supported by the presence of an olefinic proton at δ 5.72 (*d*, $J = 7.5$ Hz) in the ^1H NMR spectrum. The J value suggested a double bond between C-8 and C-30. Thus compound **3** was tentatively given the structure 6-deoxydestigloylswietenine. The 3-tiglate of this compound was previously reported from *Soyimida febrifuga* [11].

EXPERIMENTAL

Dried fruits of *Khaya senegalensis* were collected and identified by Dr D. B. Gude (Geneticist, Forest Research Centre, Wada, Pune) and a voucher specimen was deposited by him in the herbarium of Forest Research Centre, Pune, Maharashtra. Dried seeds (1.015 kg) were powdered and extracted with *n*-hexane at room temp. The defatted seed powder was then extracted with EtOH and the residue (74.2 g) after removal of the solvent was suspended in H_2O and extracted with EtOAc. The EtOAc fr after removal of solvent (50 g) was adsorbed on 100 g of 60–120 silica gel and applied to a column of 400 g silica gel (200–400) and eluted rapidly with 10–100% EtOAc in hexane. The frs from 30 and 40% EtOAc in hexane eluate gave a solid which was identified as mexican-



olide. The frs from 50% EtOAc and EtOAc eluent are similar on HPLC except for the variable amounts.

The residue from EtOAc fraction (10.5 g) was subjected to prep. HPLC using MeOH–H₂O (60–40) as eluent in a Shimpack reverse phase prep. column 25 cm $\times$ 4.6 mm with a flow rate of 10 ml min⁻¹. Details of the prep. HPLC procedure are presented in another communication [9]. In all, nine compounds were isolated in pure form out of which three were new.

Compound 1. Yield: 150mg, mp 212°; $[\alpha]_D^{25}$ –64° (MeOH; c 0.5). UV λ_{max} nm (log ϵ): 230 (3.27), 280 (2.62). IR ν_{max} cm⁻¹: 3688, 3566, 1732, 875. ¹H NMR and ¹³C NMR: Tables 1 and 2. HRMS C₂₇H₃₂O₈ 484.213195 (obs) 484.209718 (cal).

Compound 2. Yield: 100 mg mp 120° $[\alpha]_D^{25}$ +79.54° (MeOH; c 0.088). UV λ_{max} nm (log ϵ): 224 (3.07), 280 (2.82). IR ν_{max} cm⁻¹: 3685, 3560, 1720, 875. ¹H NMR and ¹³C NMR: Tables 1 and 2. HRMS (M+H) C₂₇H₃₅O₈ 487.2328 (obs) 487.2332 (cal)

Compound 3. Yield: 10 mg, mp 198°, ¹H NMR and ¹³C NMR: Tables 1 and 2. HRMS (M+H) C₂₇H₃₅O₇ 471.2360 (obs) 471.2383 (cal).

Acknowledgements—Thanks are due to the Department of Biotechnology, Govt. of India for funding a research project on Bioactive Natural Products from Meliaceae and Rutaceae. We thank Forest Research Centre, Wada, Maharashtra, India for the supply of plant material. We also thank Indian Institute of Chemical Technology, Hyderabad for HRMS Data.

REFERENCES

- Adesida, G. A., Adesogan, E. K., Okorie, D. A. and Taylor D. A. H., *Phytochemistry*, 1971, **10**, 1845.
- Taylor, D. A. H., *Progress in the Chemistry of Organic Natural Products*, eds, Herz, W., Grisebach, H., Kirby, G. W. Springer Verlag, Wien New York, 1984, p. 1.
- Adesogan, E. K. and Taylor, D. A. H., *Journal of Chemical Society(C)*, 1968, 1974.
- Olmo, L. R. V., Silva, M. F. das G. F. da., Fo, E. R., Vieira, P. C., Fernandes, J. B., Marsaioli, A. J., Pinheiro, A. L. and Vilela, E. F., *Phytochemistry*, 1996, **42**, 831.
- Champagne, D. E., Koul, O., Isman, M. B., Scudder, G. G. E. and Towers, G. H. N., *Phytochemistry*, 1992, **31**, 377.
- Govindachari, T. R., Narasimhan, N. S., Suresh, G., Partho, P. D. and Gopalakrishnan, G., *Journal of Chemical Ecology*, 1996, **22**, 1453.
- MacLachlan, L. K. and Taylor, D. A. H., *Phytochemistry*, 1982, **21**, 1701.
- Suresh, G., in *Biotechnological perspectives in Chemical Ecology of Insects*, ed. Ananthakrishnan, T. N. Oxford IBH Pub. Co. Pvt. Ltd, New Delhi, India, 1996, p. 63.
- Govindachari, T. R., Krishna kumari, G. N. and Suresh, G., *Journal of Liquid Chromatography and Related Technologies*, 1997 (In press).
- Adesogan, E. K., Powell, J. W. and Taylor, D. A. H., *Journal of Chemical Society (C)*, 1970, 1710.
- Rao, M. M., Krishna, E. M., Gupta, P. S. and Singh, P. P., *Indian Journal of Chemistry, Sec. B*, 1978, **16B**(9), 823.