

PII: S0031-9422(97)00782-6

REVISION OF STRUCTURE OF RANGIFORMIC ACID

MICHAEL H. BENN,* STEPHEN D. LORIMER† and NIGEL B. PERRY†

Chemistry Department, University of Calgary, Calgary, Alberta, Canada T2N 1N4; † Plant Extracts Research Unit, New Zealand Institute for Crop and Food Research Limited, Department of Chemistry, University of Otago, P.O. Box 56, Dunedin, New Zealand

(Received 23 June 1997)

Key Word Index—Cladia retipora; lichen; (+)-rangiformic acid; structural revision; biological activity.

Abstract—On the basis of new NMR evidence, the structure of the lichen-substance, (+)-rangiformic acid, should be revised to (2S,3S)-(+)-1-methoxycarbonylheptadecane-2,3-dicarboxylic acid. Its biological activities against a range of organisms are recorded. © 1998 Elsevier Science Ltd. All rights reserved

INTRODUCTION

While screening New Zealand plants and lichens for bioactive constituents we noted antimicrobial activity in ethanolic extracts of *Cladia retipora* [1]. This lichen had been reported to contain atranorin, usnic acid and rangiformic acid [2]; upon fractionation of lichen extracts we obtained the same three compounds. The antimicrobial properties of usnic acid and antranorin are well known [3–5], but little seems to have been reported about rangiformic acid, other than that it showed modest antibacterial activity towards *Staphylococcus aureus* [3, 4]. We report herein a reinvestigation of the structure of rangiformic acid and some further biological activities.

RESULTS AND DISCUSSION

Rangiformic acid was first isolated by Paterno from the lichen, Cladonia rangiformis [6]. It was subsequently recognized to be a mono-methyl ester of a heptadecane-1,2,3-tricarboxylic acid, nor-rangiformic acid [7, 8], whose stereochemistry was shown to be 2S,3S by enantiospecific synthesis [9]. A regio-isomeric methyl ester, (+)-isorangiformic acid, was subsequently discovered in another lichen, Lecanora stenotropa [10]. As isorangiformic acid yielded a 6-membered cyclic anhydride (as shown by IR $\nu_{\rm max}$ 1758 and 1790 cm⁻¹), it was identified as (2S,3S)-2-methoxy-carbonylheptadecane-1,3-dicarboxylic acid (1) [10]. Rangiformic acid is therefore either 2 or 3. It was first

deduced to be the 3-methoxycarbonyl compound (2) on the basis of its negative ion mass spectrum [11] and this conclusion was subsequently supported by the following argument.

In accordance with expectation, rangiformic acid formed a 5-membered cyclic anhydride (v_{max} 1774 and 1844 cm⁻¹), which must be 4 or 5. Selective irradiation of the methoxyl protons was said to result in a carbonyl ¹³C resonance changing from a complex signal to a doublet, a result interpreted as revealing that the anhydride was 4 and that rangiformic acid was (2S,3S)-3-methoxycarbonylheptadecane-1,2-dicarboxylic acid (2) [12].

This analysis requires the ester carbonyl carbon to have a significant $^2J_{\rm C,H}$ coupling to H-3, with negligible $^3J_{\rm C,H}$ couplings to H-2 and 4. As it is known that $^2J_{\rm C,H}$ and $^3J_{\rm C,H}$ are often comparable in magnitude [13–15], this requirement seemed improbable. In our hands, repetition of Huneck and Steglich's selective proton-carbon decoupling experiment on the anhydride did not simplify the carbonyl carbon to a doublet but instead gave a quartet-like multiplet. This left open the matter of whether rangiformic acid was 2 or 3.

Unlike the situation with CHCl₃-d or benzene- d_6 as solvents, in pyridine- d_5 solution at 400 and 300 MH₂, rangiformic acid gave ¹H NMR spectra in which the resonances for H-1, 2, 3, 4 and 5 were resolved. In the corresponding ¹³C-NMR spectra, the three carbonyl groups were also well separated. By COSY, DEPT and HETCOR spectra, the ¹H and ¹³C resonances for the C-1 to C-5 terminal unit were assigned (Table 1).

Our first approach to the assignment of the carbonyl resonances used selective INEPT (SELINEPT) [16, 17] measurements, optimised for a $J_{C,H}$ of 5 Hz.

^{*} Author to whom correspondence should be addressed.

Short Report

1.
$$R^1 = R^3 = H$$
, $R^2 = CH_2$

2.
$$R^1 = R^2 = H$$
, $R^3 = CH_3$

3.
$$R^2 = R^3 = H$$
, $R^1 = CH_3$

6.

$$C_{12}H_{25}$$
 $O=C$
 H
 O
 $C_{12}H_{25}$
 H
 O
 $C_{02}CH_3$

4.

5.

Irradiation of the methoxyl protons resulted in selective enhancement of the resonance at $\delta_{\rm C}$ 173.5, i.e. this was identified as the carbonyl of the ester functionality. Similarly irradiation of H-1A ($\delta_{\rm H}$ 3.33) or H-1B ($\delta_{\rm H}$ 3.08) resulted in the enhancement of both the ester carbonyl and another at $\delta_{\rm C}$ 176.5, whilst irradiation of H-2 ($\delta_{\rm H}$ 4.03) enhanced all three carbonyl resonances and of H-3 ($\delta_{\rm H}$ 3.50) the two carboxylic acid carbonyl resonances at $\delta_{\rm C}$ 176.5 and 177.2. If we exclude esterification of the carboxyl group at C-2 (which corresponds to isorangiformic acid) and accept that $^4J_{\rm C,H} \ll ^3J_{\rm C,H}, ^2J_{\rm C,H}$ [13, 14], these results indicate that the esterified carboxyl is at C-1, not C-3.

This conclusion was supported by HMBC spectra [18, 19] which revealed long-range coupling of H-1A

and H-1B to C-2 ($\delta_{\rm C}$ 44.4), C-3 ($\delta_{\rm C}$ 47.6), the ester carbonyl ($\delta_{\rm C}$ 173.5) and another at $\delta_{\rm C}$ 176.5, H-2 to C-1 ($\delta_{\rm C}$ 33.6), C-3, C-4 ($\delta_{\rm C}$ 29.3) and the carbonyls at $\delta_{\rm C}$ 173.5, 176.5 and 177.2, H-3 to C-1, C-2, C-4 and $\delta_{\rm C}$ 176.5 and 177.2, as well as the other correlations shown in Table 1. Thus, it is concluded that rangiformic acid has the structure 3, not 2.

Some years ago, at a time when rangiformic acid was recognized to be a monomethyl ester of norrangiformic acid but the site of methylation was unknown, it was suggested that it might be 3, formed from another lichen product, caperatic acid (6) [20], by dehydration and reduction [21]. Our demonstration that rangiformic acid is 3 revives this biosynthetic proposal.

Rangiformic acid (3) showed marginal activity

Short Report 1651

Table 1. ¹H and ¹³C NMR assignments for (+)-rangiformic acid (3)*

Н	$\delta_{ extsf{H}}$	HMBC correlations (to δ_C)	C	δ_{C} .
1A	3.33 (dd, 10.2, 16.7)	44.4, 47.6, 173.5, 176.5	1	33.6
1B	3.08 (dd, 4, 16.7)	44.4, 47.6, 173.5, 176.5		
2	4.03 (ddd, 4, 4, 10.2)	29.3, 33.6, 47.6, 173.5, 176.5, 177.2	2	44.4
3	3.50 (ddd, 4, 4, 6.7)	29.3, 33.6, 44.4, 176.5, 177.2	3	47.6
4A	2.15(m)		4	29.3
4B	1.84(m)			
5 A	1.69 (m)		5	28.3
6-14	1.2-1.4 (m)		614	30-30.5
15	ca 1.25 (n.o)		15	32.5
16	ca 1.25 (n.o)		16	23.3
17	0.87 (<i>dist.t</i> , ca 7)	23.3, 32.5	17	14.6
OCH ₃	3.63(s)	173.5	OCH_3	51.9
			I-CO ₂	173.5
CO₂H	12.2 (2H, brs)		2-CO ₂	176.5
			3-CO ₂	177.2

^{*} In pyridine- d_s ("100 atom","), using traces of residual pyridine for internal reference (δ_H 7.22 (H-2) and δC 123.5 (C-2)).

against Bacillus subtilis but not against Candida albicans, Cladosporium resinae, Escherichia coli, Pseudomonas aeruginosa or Trichophyton mentagrophytes (at 60 μ g per disk). We ascribe the antimicrobial activity of a crude extract of C. retipora [1] to the presence of usnic acid. Rangiformic acid (3) was inactive against P388 leukemia cells (IC₅₀ > 25 μ g ml⁻¹) but showed marginal cytotoxicity against monkey kidney (BSC-1) cells (at 60 μ g per disk).

EXPERIMENTAL

Rangiformic acid was isolated from a collection of *C. retipora* (Labill.) Nyl from the Maungatua tops, near Dunedin, in March 1996 (Plant Extracts Research Unit voucher specimen 960330-01). It had mp $105.5-107.5^{\circ}$ (uncorr.) after recrystallization from EtOAc, [α]_D+22.7° (c 0.13, EtOH). Lit. mp $102-104^{\circ}$ [12], $104-105^{\circ}$ [21], [α]_D+18.2 (EtOH) [22], +14.2° (EtOH) [12]. The anhydride, prep according to ref. [12] and recrystallised from hexane had mp $51-53^{\circ}$ (uncorr.); lit. mp $52-54^{\circ}$ [12]. ¹H and ¹³C-NMR spectra (Table 1) were recorded with Bruker AM-400 and AMX-300 spectrometers. Bioassays were performed as described in refs [1, 23].

Acknowledgements—We thank Dr D. Galloway for lichen identification, Ms G. Ellis (University of Canterbury) for biological assays and Mrs Qiao Wu and Dr R. Yamdagni of the Instrumentation Facility, Chemistry Department, University of Calgary, for measuring the SELINEPT and HMBC spectra. Partial financial support was provided for the work done in Canada by the Natural Sciences and Engineering Research Council of Canada through a grant-in-aid of research to M.H.B.

REFERENCES

- Lorimer, S. D., Barns, G., Evans, A. C., Forster, L. M., May, B. H. C., Perry, N. B. and Tangney, R. S., *Phytomedicine*, 1996, 2, 327.
- 2. Galloway, D. J., Flora of New Zealand, Lichens. Government Printer, Wellington, 1985.
- Asahina, Y. and Shibata, S., Chemistry of Lichen Substances. Japan Soc. Promot Sci., Tokyo, 1954.
- Shibata, S., Especial Compounds of Lichens, in Handbuch der Pflanzenphysiologie, Vol. X, ed. W. Ruhland. Springer-Verlag, Berlin, 1958, p. 560– 623.
- Stoll, A., Brack, A. and Renz, J., Experientia, 1947, 3, 115.
- 6. Paterno, S., Gazz. Chim. Ital., 1882, 12, 256.
- Asahina, Y. and Sasaki, T., Bull. Chem. Soc. Jpn, 1942, 17, 495.
- Aoki, M., Yakugaku Zasshi, 1946, 66, 52. (Chem. Abs., 1951, 45, 6584).
- 9. Åkermark, B., Acta Chem. Scand., 1967, 21, 589.
- 10. Huneck, S., Phytochemistry, 1982, 21, 2407.
- 11. Huneck, S., Z. Naturforsch., 1966, 21B, 888.
- 12. Huneck, S. and Steglich, W., *Phytochemistry*, 1983, **22**, 2855.
- Kalinowski, H.-O., Berger, S. and Braun, S., Carbon-13 NMR Spectroscopy. J. Wiley and Sons, Chichester, 1988.
- 14. Breitmaier, E. and Voelter, W., *Carbon-13 NMR Spectroscopy*, 3rd edn. VCH, Weinheim, 1990.
- Pretsch, E., Clerc, T., Seibl, J. and Simon, W., Tables of Spectral Data for Structure Determination of Organic Compounds (English transl. of revised 2nd German edm). Springer Verlag, Berlin, 1983.
- 16. Bax, A., J. Magn. Res., 1984, 57, 314.
- Cordell, G. A. and Kinghorn, A. D., *Tetrahedron*, 1991. 47, 3521.

- Kessler, H., Bernd, M., Kogler, H., Zarbok, J., Sorensen, O. W., Bodenhausen, G. and Ernst, R. R., J. Amer. Chem. Soc., 1983, 105, 6944.
- Bax, A., Davis, D. G. and Sarker, S. K., J. Magn. Res., 1985, 63, 230.
- 20. Brandänge, S., Josephson, S., Mörch, L. and Vallén, S., Acta Chem. Scand., 1977, 31, 307.
- 21. Turner, W. B., Fungal Metabolites. Academic Press, London, 1971, p. 291.
- 22. Culberson, C. F., Chemical and Botañical Guide to the Lichens. U.N. Carolina Press, 1969.
- Lorimer, S. D., Perry, N. B. and Tangney, R. S., J. Nat. Prod., 1993, 56, 1444.