

XANTHONES FROM THE TWIGS OF *MAMMEA SIAMENSIS*

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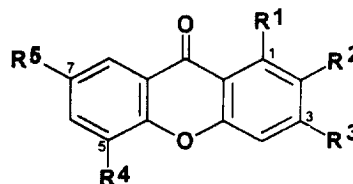
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Key Word Index—*Mammea siamensis*; Guttiferae; 1,2-dimethoxy-5-hydroxyxanthone; xanthones.

Abstract—1,2-Dimethoxy-5-hydroxyxanthone, a new xanthone, was isolated from the twigs of *Mammea siamensis*, in addition to six known xanthones (5-hydroxy-1-methoxy-, 1,3-dimethoxy-5-hydroxy-, 2,5-dihydroxy-1-methoxy-, 1,7-dihydroxy-, 1,3,7-trihydroxy- and 3,5-dihydroxy-1-methoxyxanthone). Structures for these compounds were deduced from their spectral data. © 1998 Elsevier Science Ltd. All rights reserved

INTRODUCTION

Mammea siamensis (Miq.) T. Anders. (*Ochrocarpus siamensis* Anders.), a small, evergreen tree up to 15 m tall and 10–30 cm in diameter, is native to Myanmar, Thailand, Laos, Cambodia and Vietnam [1]. The genus *Mammea* of the family Guttiferae is reported to contain various types of xanthones [2–10]. Previously, two phenylcoumarins, 6-butyryl-5-hydroxy-4-phenylseselin and 6-butyryl-5,7-dihydroxy-8-(3,3-dimethylallyl)-4-phenylcoumarin, were isolated from the flowers of *M. siamensis* [11], and ochrocarpus proanthocyanidin 1 has been isolated from its leaves and found to have piscicidal and molluscicidal activities [12]. Fractionation of the MeOH extract of the twigs of *M. siamensis* resulted in the isolation of a novel xanthone 3, and the six known analogues, 1, 2 and 4–7.



	R ₁	R ₂	R ₃	R ₄	R ₅
1	OCH ₃	H	H	OH	H
2	OCH ₃	H	OCH ₃	OH	H
3	OCH ₃	OCH ₃	H	OH	H
4	OH	H	H	H	OH
5	OCH ₃	OH	H	OH	H
6	OH	H	OH	H	OH
7	OCH ₃	H	OH	OH	H

RESULTS AND DISCUSSION

The MeOH extract of the twigs of *M. siamensis* was partitioned against petroleum ether and CHCl₃, respectively. An LC-MS dereplication process of the CHCl₃–MeOH extract showed marginal cytotoxic activity in the BC1 (human breast cancer) (ED₅₀ = 8.3 µg/ml) and ZR-75-1 (hormone-dependent human breast cancer) (ED₅₀ = 8.6 µg/ml) cell lines, and chromatography of this extract by semi-preparative RP-

HPLC or by silica gel columns resulted in the isolation of xanthones 1–7.

Compounds 1–2, and 4–7 were identified by spectroscopic data as 5-hydroxy-1-methoxyxanthone (1) [2, 3], 1,3-dimethoxy-5-hydroxyxanthone (2), 1,7-dihydroxyxanthone (euxanthone) (4) [2, 3, 13], 2,5-dihydroxy-1-methoxyxanthone (5) [14], 1,3,7-trihydroxyxanthone (6) [3], and 3,5-dihydroxy-1-methoxyxanthone (7) [15], respectively.

Compound 3, 1,2-dimethoxy-5-hydroxyxanthone, is a yellow amorphous solid. The HREI-MS gave a [M]⁺ at *m/z* 272.0675 (C₁₅H₁₂O₅). The ¹H NMR

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spectrum displayed two methoxyl groups (δ 3.77, *s*; 4.15, *s*), five aromatic protons (δ 7.11, *d*; 7.27, *t*; 7.34, *d*; 7.51, *dd*; 8.08, *dd*), and a hydroxyl group (δ 12.67, *br s*). The COSY experiment showed an *ortho* correlation ($J = 7.9$ Hz) between the *dd* at δ 8.08 and the *t* at δ 7.26. A weak *meta* correlation ($J = 1.6$ Hz) was also observed between the resonances at δ 8.08 and at δ 7.51. These correlations suggested that these three protons were on the same aromatic ring. Consequently, the *ortho* coupled ($J = 9.2$ Hz) doublets at δ 7.34 and at δ 7.11 in the COSY spectrum were placed on the other aromatic ring. The assignments of these five protons were in accordance with those of 2,5-dihydroxy-1-methoxyxanthone (**5**), which is the 2-hydroxy analogue of this xanthone. In the NOE experiment, irradiation of the methoxyl resonance at δ 3.77 enhanced the resonance at δ 7.34 (5.1%). On the other hand, irradiation of the methoxyl resonance at δ 4.15 or the proton at δ 12.67 did not show any NOE effect. These NOE results indicated that the methoxyl group (δ_{H} 3.77, δ_{C} 56.8) was adjacent to the proton at δ 7.34. Unambiguous carbon assignments for this xanthone were not possible because of sample size. However, following the establishment, using 2D NMR techniques, of the unambiguous assignments of 2,5-dihydroxy-1-methoxyxanthone (**5**), tentative assignments were made for **3** which established the structure as 1,2-dimethoxy-5-hydroxyxanthone.

Compounds **1**, **2**, and **5** were evaluated for their cytotoxic activity against various human cancer cell lines, i.e., Lu-1 (human lung cancer), KB (human oral epidermoid carcinoma), LNCaP (hormone-dependent human prostate cancer), and ZR-75-1, using the procedures described previously [16], and deemed to be inactive in these assay systems (ED_{50} values 20 $\mu\text{g}/\text{ml}$). However, compound **2** has been reported to inhibit type A and type B monoamine oxidases [17], and compound **6** was reported to possess tuberculostatic activity [18].

EXPERIMENTAL

General

Mps: Fisher-Johns melting apparatus (uncorr). UV: Beckman DU-7 spectrometer. IR: Midac Collegian FT-IR spectrometer. HREIMS and EIMS: Finnigan MAT 90 mass spectrometer. ^1H -, ^{13}C -NMR, NOE, and COSY: Varian XL-300 NMR spectrometer (299.9 MHz for ^1H and 75.4 MHz for ^{13}C), using TMS as an int. st. Selective INEPT spectra: Nicolet NMC-360 NMR spectrometer (360 MHz for ^1H and 90.8 MHz for ^{13}C). HMBC: General Electric GE-500 (500.12 MHz).

Plant material

Mammea siamensis (Miq.) T. Anders. was collected in Saraburi, Thailand in October 1991. A voucher

specimen (AA768) is deposited at the Field Museum of Natural History, Chicago, Illinois, U.S.A.

Extraction and isolation

Dried and ground twigs of *M. siamensis* (3.63 kg) were extracted with 95% MeOH (13 l $\times$ 4). The combined extract was concentrated *in vacuo* (356 g), redissolved in 1 l water and extracted with petroleum ether (3 $\times$ 1 l). The aq. MeOH fr. was then extracted with CHCl_3 (1:1), and the CHCl_3 -MeOH fr. was washed with 1% NaCl soln to remove tannins. This CHCl_3 -MeOH fr. was dried *in vacuo* (12.9 g) and subjected to Si gel CC, eluting with a mixture of CHCl_3 -MeOH. Fr. 6 (100 mg) from the CHCl_3 -MeOH (100:0) eluate was further purified by semi-prep. HPLC (Kromasil[®] C₁₈ 250 $\times$ 20 mm), using a gradient mobile phase [$\text{H}_2\text{O}:\text{CH}_3\text{CN}$ (65:35) to $\text{H}_2\text{O}:\text{CH}_3\text{CN}$ (0:100) in 50 min], and the detector was set at 280 nm to yield the xanthenes **1** (6.8 mg), **2** (6.2 mg), **3** (2.9 mg), and **4** (7.2 mg) with retention times of 13.2, 14.3, 16.2, and 31.8 min, respectively. Frs 15-17 from the CHCl_3 -MeOH (95:5) eluate were combined (600 mg) and chromatographed on Si gel CC, using CHCl_3 -MeOH (99:1) as an eluent, to afford the xanthenes **5** (36.0 mg) and **6** (9.5 mg). Fr. 18 (600 mg) from the CHCl_3 -MeOH (95:5) eluate was further purified on a Si gel CC, eluting with CHCl_3 -MeOH (90:10), to yield the xanthone **7** (1.2 mg).

LC-MS dereplication procedure

The CHCl_3 -MeOH extract of *M. siamensis* twigs, prepared as indicated above, was subjected for dereplication analysis, employing a previously published protocol, with the published chromatographic conditions being used, and the ZR-75-1 cytotoxicity assay used to monitor activity [19, 20].

1,2-Dimethoxy-5-hydroxyxanthone (**3**). Yellow amorphous residue. HREI-MS m/z 272.0675 for $\text{C}_{15}\text{H}_{12}\text{O}_5$ (calcd 272.0685). EI-MS m/z (rel. int.): 272 [$\text{M}]^+$ (100), 258 (11), 257 (76), 253 (16), 243 (33), 239 (15), 229 (42), 113 (38). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 255 (2.66), 372 (1.49). IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm^{-1} : 3459 (OH), 1640 ($>\text{C}=\text{O}$), 1584. ^1H NMR (299.9 MHz, pyridine- d_5): 3.77 (3H, *s*, OCH_3 -2), 4.15 (3H, *s*, OCH_3 -1), 7.11 (1H, *d*, $J = 9.2$ Hz, H-4), 7.27 (1H, *t*, $J = 7.9$ Hz, H-7), 7.34 (1H, *d*, $J = 9.2$ Hz, H-3), 7.51 (1H, *dd*, $J = 1.6$, 7.9 Hz, H-6), 8.08 (1H, *dd*, $J = 1.6$, 7.9 Hz, H-8), 12.67 (1H, *br s*, OH-5). ^{13}C NMR (75.4 MHz, pyridine- d_5): 56.8 (*s*, OCH_3 -2), 61.6 (*s*, OCH_3 -1), 113.4 (*d*, C-4), 116.2 (*d*, C-8), 117.5 (*s*, C-9a), 120.4 (*d*, C-6), 120.6 (*d*, C-3), 124.0 (*s*, C-8a), 124.2 (*d*, C-7), 135.6 (*s*, C-4a), 145.9 (*s*, C-10a), 147.8 (*s*, C-5), 149.0 (*s*, C-1), 151.4 (*s*, C-2), 176.5 (*s*, C-9).

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