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# A SESQUITERPENE FROM GARDENIA SOOTEPENSIS

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Abstract—From the twigs of *Gardenia sootepensis* a new sesquiterpene with a guaiane skeleton named sootepdienone was isolated, in addition to known flavones and benzoic acid derivatives. © 1998 Elsevier Science Ltd. All rights reserved

#### INTRODUCTION

The genus Gardenia Ellis consists of more than 80 species spread among the tropical forests of certain regions of the world. Seventeen species of this genus occur in Thailand [1]. One of these is G. sootepensis, local name: Kam-mok-luang, which grows only in the northern part of Thailand. Although there are a number of reports on the isolation of triterpenes [2-6], iridoid glycosides [7–13], quinic acid derivatives [8, 14-15], a keto fatty acid [16] and flavones [6, 17-24] from many Gardenia species, the chemical constituents of G. sootepensis have not been extensively investigated. The flowers were found to contain  $\beta$ -sitosterol, a highly conjugated diterpene carboxylic acid, 7,4'-dihydroxyflavone and long-chain aliphatic compounds [6], while the fruits contain a quinic acid lactone (quinide) [25]. In an examination of twigs, we have now isolated a new sesquiterpene together with benzoic acid, 4-hydroxy-3-methoxybenzoic acid, 4hydroxy-3,5-dimethoxybenzoic acid, 5.7,4'-trihydroxy-6-methoxyflavone [26-28] and 5,7,3'-trihydroxy-6,4',5'-trimethoxyflavone [29–31].

#### RESULTS AND DISCUSSION

A crude MeOH extract of the ground twigs of G. sootepensis was partitioned between 20% aq. NaOH and ether. The ether-soluble material was repeatedly chromatographed on silica gel to give compound 1.

The alkali-soluble fraction was separated into phenolic and acidic fractions which were chromatographed on silica gel. The phenolic fraction yielded 5,7,4'-trihydroxy-6-methoxyflavone and 5,7,3'-trihydroxy-6,4',5'-trimethoxyflavone. The acidic fraction afforded benzoic acid, 4-hydroxy-3-methoxybenzoic acid and 4-hydroxy-3,5-dimethoxybenzoic acid.

Compound 1, sootepdienone, was obtained as an optically active, colourless solid. Strong peaks at 3420 and 1680 cm $^{-1}$  in the IR spectrum indicated the presence of hydroxyl and unsaturated carbonyl functionalities. The UV spectrum ( $\lambda_{\rm max}$  245, 300 nm) indicated an extended conjugated system. The HRMS and  $^{\rm 1}H$  and  $^{\rm 13}C$  NMR data (Table 1) established the molecular formula to be  $C_{1s}H_{22}O_2$ . The  $^{\rm 1}H$  NMR spectrum was assigned by decoupling and 2D  $^{\rm 1}H^{-\rm 1}H$  COSY experiments and the  $^{\rm 13}C$  NMR signals were assigned from DEPT, HMQC and GSHMBC spectra. The  $^{\rm 1}H^{-\rm 1}H$  correlations enabled us to establish the following subunits.

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The  $^{13}$ C NMR signals for quaternary carbons at  $\delta_{\rm C}$ 74.4 (C-11) and 209.3 (C-3) indicated the presence of a tertiary hydroxy group and an unsaturated keto function, respectively. An olefinic proton at  $\delta_{\rm H}$  6.75 (brd, J = 1.7 Hz, H-6) in the <sup>1</sup>H NMR spectrum together with four signals of  $sp^2$  carbons at  $\delta_C$  167.1 (C), 160.4 (C), 137.4 (C) and 118.2 (CH) confirmed the presence of trisubstituted and tetrasubstituted double bonds. The <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra further showed the presence of four methyl groups [ $\delta_H$  0.71 (3H, d, J = 6.6 Hz, H-14;  $\delta_C$  14.5), 1.41 (6H, s, H-12 and H-13;  $\delta_C$  28.6 and 28.8) and 1.70 (3H, d, J = 1.8Hz, H-15;  $\delta_C$  8.3], two methine groups [ $\delta_H$  3.33 (1H, m, H-1;  $\delta_C$  43.6) and 2.23 (1H, m, H-10;  $\delta_C$  32.9)] and three methylene groups [( $\delta_H$  2.50, 1H, dd, J = 6.6 and 18.6 Hz, H-2 $\alpha$ ;  $\delta_{\rm H}$  2.24, 1H, dd, J=2.1 and 18.6 Hz, H-2 $\beta$ ;  $\delta_C$  39.7), ( $\delta_H$  2.58, 1H, m; H-8 $\alpha$ ;  $\delta_H$  2.35, 1H, ddd, J = 2.7, 7.8 and 16.5 Hz, H-8 $\beta$ ;  $\delta_C$  26.5) and ( $\delta_H$ 2.15, 1H, m; H-9 $\alpha$ ;  $\delta_{\rm H}$  1.48, 1H, m; H-9 $\beta$ ;  $\delta_{\rm C}$  35.1)].

The connections of the subunits were determined from GSHMBC data. The carbonyl carbon at  $\delta_{\rm C}$  209.3 showed cross peaks with Me-15 and H-2 indicating the connection of the carbonyl carbon to C-2 and C-4. The  $^3J_{\rm C}$  H connection between the  $sp^3$  secondary carbon at  $\delta_{\rm C}$  35.1 (C-9) and Me-14 revealed the link between C-9 and C-10. The quaternary hydroxylated carbon at  $\delta_{\rm C}$  74.4 (C-11) showed correlations with Me-12, Me-13 and H-6 whereas the quaternary carbon at  $\delta_{\rm C}$  160.4 (C-7) showed  $^3J_{\rm C}$  H connection with Me-12

and Me-13, and H-9 $\alpha$  and  ${}^2J_{C-H}$  connection with H-8α, H-8β. Thus, the hydroxylated carbon C-11 carries two methyl groups and links with C-7 of the trisubstituted double bond. The relative stereochemistry of 1 was determined by 1H NMR, NOE difference and NOESY spectra. Irradiation of the H-1 signal caused a strong enhancement of the H-10 signal (as well as H-2 $\alpha$  and H-8 $\alpha$ ), and irradiation of Me-14 gave enhancements of H-2 $\beta$  and H-8 $\beta$ , suggesting that Me-14 was trans to H-1. H-1α had homoallylic coupling (J = 1.8 Hz) with Me-15 and weak  ${}^4J_{\text{H-H}}$  coupling with H-6; H-8 $\alpha$  (quasi-axial) had strong (J = 1.7 Hz) allylic coupling with H-6, whereas H-8 $\beta$  (quasi-equatorial) showed no coupling. These results are consistent with the assigned structure, 11-hydroxy-4,6-guaiadien-3one. A number of hydroxyguaiadienones have been reported from natural sources, including three with the  $\Delta^{4,5}$ -guaian-3-one structure [32–34]. Nevertheless, sootepdienone represents a new member of the class.

#### **EXPERIMENTAL**

General

<sup>1</sup>H NMR: 400 and 600 MHz, CDCl<sub>3</sub> solns with TMS as int. standard; <sup>13</sup>C NMR: 100 and 150 MHz. Multiplicities were determined by DEPT experiments. Known compounds were identified by comparison with their reported spectroscopic data, and the pos-

Table 1. 1H NMR and 13C NMR data of compound 1

Н	$\delta$	J (Hz)	C	δ	DEPT
1	3.33 m		]	43.6	СН
2α	2.50 dd	18.6, 6.6	2	39.7	CH <sub>2</sub>
$2\beta$	2.24 dd	18.6, 2.1	3	209.3	C
6	6.75 brd	1.7	4	137.4	C
8α	2.58 m		5	167.1	C
$8\beta$	2.35 ddd	16.5, 7.8, 2.7	6	118.2	CH
9α	2.15 m		7	160.4	C
9β	1.48 m		8	26.5	CH,
10	2.23 m		9	35.1	CH <sub>2</sub>
12	1.41 s		10	32.9	CH
13	1.41 s		11	74.4	C
14	0.71 d	6.6	12	28.6	$CH_3$
15	$1.70 \ d$	1.8	13	28.8	$CH_3$
			14	14.5	$CH_3$
			15	8.3	$CH_3$

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ition of OMe groups confirmed by NOE difference spectra.

#### Plant material

Twigs of *Gardenia sootepensis* were collected from Doi Sootep, Chiang Mai, Thailand. The voucher specimen (Max 93-1139) has been deposited at the herbarium of the Department of Biology, Faculty of Science, Chiang Mai University.

## Extraction and isolation

The ground dried twigs (1.45 kg) were consecutively extracted with hexane, CH<sub>2</sub>Cl<sub>2</sub> and MeOH. The concentrated MeOH extract (2 g) was partitioned between 20% aq. NaOH and Et<sub>2</sub>O. The Et<sub>2</sub>O-soluble fraction was evaporated to dryness to give a yellow viscous oil, which was fractioned by quick CC using gradient elution: CH<sub>2</sub>Cl<sub>2</sub>-hexane, EtOAc-CH<sub>2</sub>Cl<sub>2</sub> and MeOH-EtOAc. Frs 4 and 5, upon prep. TLC using EtOAc-CH<sub>2</sub>Cl<sub>2</sub> (1:9), afforded compound 1. The aq. layer was neutralized with 20% aq. HCl and further extracted with EtOAc (3×100 ml). The combined organic extracts were concd to dryness in vacuo to afford a residue (331 mg) which was chromatographed on a column of silica gel and eluted with CH<sub>2</sub>Cl<sub>2</sub>hexane, EtOAc-CH<sub>2</sub>Cl<sub>2</sub> and MeOH-EtOAc. The third eluate (38 mg) was then subjected to prep. TLC on silica gel using MeOH-CH<sub>2</sub>Cl<sub>2</sub> (1:19) to give two bands. After eluting with EtOAc, 5,7,4'-trihydroxy-6methoxyflavone (7 mg) was obtained from the lower band. The upper band afforded 5,7,3'-trihydroxy-6,4',5'-trimethoxyflavone (10 mg). The neutral aq. layer was further acidified with 20% aq. HCl and then extracted with EtOAc. Upon evapn, the crude extract was obtained as a brown viscous oil. The oil was partially soluble in Et<sub>2</sub>O. Chromatographic sepn of the Et<sub>2</sub>O-soluble fraction afforded benzoic acid, 4hydroxy-3-methoxybenzoic acid and 4-hydroxy-3,5dimethoxybenzoic acid.

## Compound 1

Colourless solid, mp 47–50°. [ $\alpha$ ]<sub>D</sub><sup>20</sup> +12° (c. 0.1 CHCl<sub>3</sub>). IR (CHCl<sub>3</sub>)  $\nu_{max}$  3240, 1680 cm<sup>-1</sup>; UV  $\lambda_{max}$  EtOH nm ( $\epsilon$ ) 245 (5900), 296 (10,500); <sup>1</sup>H NMR and <sup>13</sup>C NMR (CDCl<sub>3</sub>): Table 1; MS, m/z (rel. int.); 234 [M<sup>+</sup>] (11.5), 219 (13.1), 216 (26.2), 173 (15.5), 43 (100): HRMS Calc. for  $C_{15}H_{22}O_2$  234.1620; Found: 234.1612.

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#### REFERENCES

- Suvatti, C., in Flora of Thailand, Vol. 2, Royal Institute, Bangkok, 1978.
- 2. Davies, N. W., Miller, J. M., Naidu, R. and Sotheeswaran, S., *Phytochemistry*, 1992, 31, 159.
- 3. Reddy, G. C. S., Ayengar, K. N. N. and Rangaswami, S., *Phytochemistry*, 1975, **14**, 307.
- Reddy, G. C. S., Ayengar, K. N. N. and Rangaswami, S., Phytochemistry, 1973, 12, 1831.
- Reddy, G. C. S., Ayengar, K. N. N. and Rangaswami, S., *Indian Journal of Chemistry*, 1975, 13, 749.
- 6. Liang, H., Zheng, H. and Chen S., Yunnan Zhiwu Yanjin, 1991, 13, 95.
- 7. Watanabe, N., Nakajima, R., Watanabe, S., Moon, J., Inagaki, J., Sakata, K., Yagi, A. and Ina, K., *Phytochemistry*, 1994, 37, 457.
- 8. Nishizawa, M., Izuhara, R., Kaneko, K., Koshihara, Y. and Fujimoto, Y., *Chemical and Pharmaceutical Bulletin*, 1988, 36, 87.
- 9. Takeda, Y., Nishimura, H., Kadota, O. and Inouye, H., Chemical and Pharmaceutical Bulletin, 1976, 24, 2644.
- Inouye, H., Takeda, Y. and Nishimura, H., Phytochemistry, 1974, 13, 2219.
- 11. Endo, T. and Taguchi, H., Chemical and Pharmaceutical Bulletin, 1970, 18, 1066.
- Inouye, H., Saito, S. and Shigu, T., *Tetrahedron Letter*, 1970, 41, 3581.
- Inouye, H., Saito, S., Taguchi, H. and Endo, T., Tetrahedron Letter, 1969. 28, 2347.
- Nishizawa, M., Izuhara, R., Kaneko, K. and Fujimoto, Y., Chemical and Pharmaceutical Bulletin, 1987, 35, 2133.
- 15. Nishizawa, M. and Fujimoto, Y., Chemical and Pharmaceutical Bulletin, 1986, 34, 1419.
- Mahmood, C., Daulatabad, J. D., Mulla, G. M. M., Mirajkar, A. M. and Hosamami, K. M., Phytochemistry, 1991, 30, 2399.
- 17. Gunatilaka, A., Sirimanne, S., Sotheeswaran, S. and Sriyana, H., *Phytochemistry*, 1982, **21**, 805.
- Chhabra, S. C., Gupta, S. R., Sharma, C. S. and Sharma, N. D., *Phytochemistry*, 1977, 16, 1109.
- Chhabra, S. C., Gupta, S. R. and Sharma, N. D., Phytochemistry, 1977, 16, 399.
- Chhabra, S. C., Gupta, S. R., Seshadri, T. R. and Sharma, N. D., *Indian Journal of Chemistry*, 1976, 14B, 651.
- Gupta, S. R., Seshadri, T. R., Sharma, C. S. and Sharma, N. D., *Indian Journal of Chemistry*, 1975, 13, 785.
- Krishnamurti, M., Seshadri, T. R. and Sharma,
  N. D., Indian Journal of Chemistry, 1972, 10, 23.
- Krishnamurti, M., Seshadri, T. R. and Sharma,
  N. D., Indian Journal of Chemistry, 1971, 9, 189.
- Rao, R., Venkataraman, K., Chakrabarti, P., Sanyal, A. K. and Bose, P. K., *Indian Journal of Chemistry*, 1970, 8, 398.
- 25. Wang, G., Zhao, S., Chen, D., Yang, L., Wu, N.

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and Zheng, Q., Journal of Structural Chemistry, 1996, 15, 400.

- 26. Bacchi, M. and Carrier, M., *Planta Medica*, 1980, **38**, 267.
- 27. Mues, R., Timmermann, B. N., Oho, N. and Mabry, T. J., *Phytochemistry*, 1979, **18**, 1379.
- 28. Liu, H., Ho, D. K. and Cassady, J. M., *Journal of Natural Products*, 1992, **55**, 357.
- 29. Marco J. A., Barbera, O., Rodriguez, S., Domingo, C. and Adell, J., *Phytochemistry*, 1988, 27, 3155.
- 30. Liu, Y. and Mabry, T. J., *Phytochemistry*, 1982, **21**, 209.
- 31. Liu, Y. and Mabry, T. J., *Phytochemistry*, 1981, **20**, 309.
- 32. Singh, P. and Suri, A., *Phytochemistry*, 1990, **29**, 3944.
- Zdero, C., Bohlmann, F., Solomon, J. C., King R. M. and Robinson, H., *Phytochemistry*, 1989, 28, 531
- 34. Bohlmann, F., Zdero, C., King, R. M. and Robinson, H., *Phytochemistry*, 1983, 22, 1201.