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TWO PRENYLATED FLAVANONES FROM STEM BARK OF ERYTHRINA BURTTII

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Key Word Index—*Erythrina burttii*; Leguminosae; stem bark; flavanones; abyssinone V-4′-methyl ether; burttinone.

Abstract—From the stem bark of *Erythrina burttii*, two new flavanones were isolated and characterised as 5,7-dihydroxy-4'-methoxy-3',5'-di-(3-methylbut-2-enyl)flavanone (trivial name, abyssinone V-4'-methyl ether) and 5,7-dihydroxy-4'-methoxy-3'-(3-hydroxy-3-methylbut-1-enyl)-5'-(3-methylbut-2-enyl)flavanone (trivial name, burttinone). In addition, seven known compounds were identified. Structures were determined on the basis of spectroscopic evidence. © 1998 Published by Elsevier Science Ltd. All rights reserved

INTRODUCTION

The genus *Erythrina* comprises over 100 species distributed in tropical and subtropical regions of the world [1]. It is known for the production of alkaloids and C-prenylated flavoids [1, 2].

Erythrina burttii is a flat-topped tree growing 3.5–18 m tall and is found in Kenya and Tanzania [3]. From the stem bark of this species, we have isolated two new C-prenylated flavanones, along with seven known compounds. The isolation and characterization of these compounds is presented here.

RESULTS AND DISCUSSION

The first compound (1) showed a [M]⁺ at m/z 422.2094 corresponding to the molecular formula $C_{26}H_{30}O_5$. The presence of a flavanone skeleton was evident from the UV (λ_{max} 287, 328 nm), ¹H (δ 5.33 for H-2, 2.76 and 3.11 for H-3) and ¹³C (δ 79.3 for C-2, 43.0 for C-3 and 196.3 for C-4) NMR spectra. The ¹H (Table 1) and ¹³C (Table 2) NMR spectra further showed the presence of two hydroxyl (one chelated), a methoxyl and two 3-methylbut-2-enyl substituents on the flavanone skeleton. In the EI-mass spectrum, the fragment ion at m/z 153 [C₇H₈O₄]⁺, caused by RDA-fragmentation of the C-ring showed that the A-ring possessed the two hydroxyl groups [1], and, therefore, the methoxyl and the two 3-methylbut-2-

enyl substituents should be located on the B-ring. The NMR spectra further revealed that the two 3methylbut-2-enyl groups are symmetrically substituted on the B-ring, being either at C-2' and C-6' or C-3' and C-5', with the methoxyl at C-4'. The ¹³C NMR resonance value for the methoxyl (δ 60.9) was shifted downfield compared with the normal value (δ 55.0-56.5). This shift requires that both *ortho*-positions are substituted [5], allowing the placement of the two 3-methylbut-2-enyl groups at C-3' and C-5'. These observations were strongly suggestive of structure 1 and this was confirmed by HMOC and HMBC experiments. The CD spectrum showed a positive Cotton effect at 328 nm and a negative one at 287 nm, consistent with the S-configuration at C-2 [6, 7]. Compound 1 is thus a C-4'-methyl ether derivative of abyssinone V (2) [4], which was also isolated in the present study. Hence, the trivial name abyssinone V-4'-methyl ether is given to this new compound.

The third isolate (3), which analysed for $C_{26}H_{30}O_6$ by HR mass spectrometry, was also a flavanone derivative, as was evident from the UV. ¹H (Table 1) and ¹³C (Table 2) NMR spectra. Comparison of the ¹H and ¹³C NMR data of this compound with those of 1 and 2 showed identical A-ring substitution. This was supported by the EI-mass spectrum which showed a fragment ion at m/z 153, consistent with the presence of two hydroxyl substituents on this ring. Two *meta*-coupled protons at δ 7.37 and 7.09 could be assigned to H-2' and H-6' of the B-ring with substituents at C-3', C-4' and C-5'. This was supported by the HMBC spectrum (Fig. 1), which showed correlation between H-2' and H-6' to C-2 (δ 79.0). In the ¹³C NMR spectrum, the downfield chemical shift value of the meth-

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oxyl group (δ 61.2) requires that it should be located at C-4′, with the other substituents being at C-3′ and C-5′. One of these substituents is clearly a 3-methylbut-2-enyl group (Tables 1 and 2). The other substituent was established to be a *trans*-3-hydroxy-3-methylbut-1-enyl unit. This is apparent from the ¹H NMR spectrum which showed a pair of doublets at δ 6.84 and 6.40 (J=16.2 Hz) corresponding to the two vinyl protons [8], while the presence of a tertiary hydroxyl group on this substituent was deduced from the ¹³C NMR (δ 71.8 for C-3″ and 29.5 for C-4″/5″)

and mass $(m/z \ 420 \text{ for } [M-H_2O]^{\top})$ spectra. The nature of the substituent at C-3' and of the substitution pattern in the B-ring was confirmed by HMBC (Fig. 1). The CD spectrum of this compound showed similar cotton effects to that of compound 1, again suggesting S-configuration at C-2. These data are consistent with structure 3 for this new compound; the trivial name, burttinone, has been assigned.

The UV, mass, ¹H and ¹³C NMR data of compound 4 were consistent with a 3,9-dihydroxypterocarpan with a 3-methylbut-2-enyl substituent at either C-2 or

Table 1. ¹H NMR data of flavanones isolated from *Erythrina* burttii (300 MHz, CDCl₃)

our mile, ebelg							
Н	1	2	3	6			
2	5.33 dd	5.30 dd	5.28 dd	5.30 dd			
3	2.76 dd	2.74 dd	2.71 dd	2.77 dd			
	3.11 dd	3.09 dd	3.04 dd	3.06 dd			
6	5.98 s	5.97 s	5.99 s	5.99 s			
8	5.98 s	5.97 s	5.99 s	5.99 s			
2'	7.09 s	7.05 s	7.37 d	6.91 d			
61	$7.09 \ s$	7.05 s	7.09 d	6.75 d			
1"	3.39 d	3.35 d	6.84 d	3.37 d			
2"	5.29 t	5.28 t	$6.40 \ d$	5.28 t			
4"	1.73 s	1.76 s	$1.45 \ s$	$1.73 \ s$			
5"	1.74 s	1.76 s	1.45 s	1.75 s			
I "'	3.39 d	3.35 d	3.35 d				
2‴	5.29 t	5.28 t	5.23 /				
4""	1.73 s	1.76 s	$1.70 \ s$				
5‴	1.74 s	$1.76 \ s$	1.70 s				
4′-OMe	3.80 s		3.71 s	3.81 s			
5-OH	12.05 s	$12.07 \ s$	12.02 s	12.03 s			

J values: H-2/H-3 2.7, 13.2 Hz; H-3_{gem} - 17.1 Hz; H-1"/H-2", H-1"'/H-2" 7.2 Hz; in **3** and **7** H-2'/H-6' 2.1 Hz; in **3** H-1"(H-2" 16.2 Hz.

Table 2. ¹³C NMR data of flavanones isolated from *Erythrina* burttii (75 MHz, CDCl₃)

7.4.1.1.1 (7.5.1.1.1.1.1.1.1.1.1.1.1.1.1.1.1.1.1.1.						
C	1	2	3	6		
2	79.3	79.3	79.0	79.0		
3	43.0	43.0	43.1	43.1		
4	196.3	196.4	195.9	196.1		
5	164.2	164.1	164.1	164.2		
6	96.6	96.6	96.7	96.7		
7	165.3	165.7	166.0	164.8		
8	95.6	95.6	95.7	95.6		
9	163.3	163.3	163.1	163.2		
10	103.0	102.7	102.6	103.1		
1'	133.7	129.7	134.0	134.7		
2'	125.9	126.1	122.6	111.3		
3'	135.4	127.6	130.7	149.1		
4'	156.4	153.3	155.9	145.3		
5'	135.4	127.6	135.6	135.4		
6'	125.9	126.1	127.1	119.3		
1"	28.4	29.6	121.2	28.2		
2"	122.5	121.5	139.0	121.9		
3"	133.0	134.8	71.8	133.4		
4"	25.7	25.8	29.5	25.7		
5"	17.9	17.9	29.5	17.9		
I'''	28.4	29.6	28.3			
2"	122.5	121.5	122.3			
3"	133.0	134.8	133.0			
4'''	25.7	25.8	25.7			
5′″	17.9	17.9	17.8	******		
OMe	60.9	***	61.2	61.2		

C-assignments confirmed by HMQC and HMBC experiments.

C-8. The position of attachment of this substituent was fixed from the long-range HMBC correlation spectrum, which showed interactions of H-1 to C11a and also to the methylene carbon of the 3-methyl-2-enyl group. These correlations clearly establish that the isolated compound is calopocarpin (4), which has earlier been reported as a phytoalexin [9]. The CD spectrum which showed a positive Cotton effect at 293 nm and the high negative optical rotation ($[\alpha]_D = -262.5^\circ$) are consistent with the 6aR:11aR absolute configuration of 4 [7, 10].

The remaining compounds isolated from *E. burttii* were identified as neorautenol (5) [9], sigmoidin B-4′-methyl ether (6) [11], bidwillon A (7) [12], isobavachalcone (8) [13] and erythrinasinate (9) [14, 15].

EXPERIMENTAL

General

Mps: uncorr. Analytical TLC: Merck pre-coated silica gel 60 F₂₅₄; compounds were detected under UV light (254 nm). CC: silica gel 60 (70–230) mesh and Sephadex LH-20. EIMS: direct inlet, 70 eV. ¹H NMR (300 MHz) and ¹³C NMR (75.48 Mz) were recorded on ARX 300 (Bruker) spectrometer using TMS as int. standard. HMQC and HMBC spectra were acquired using standard Bruker software.

Plant material

Stem bark of *E. burttii* Ball. f. was collected near Emali town, on the Nairobi-Mombasa road, Kenya, in November 1994. The plant was identified at the Herbarium, Botany Department, University of Nairobi, where a voucher specimen is deposited.

Extraction and isolation

Dried and ground stem bark (800 g) was extracted with CHCl₃ by cold percolation. The concd extract (20 g) was fractionated on silica gel (200 g) eluting

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with hexane containing increasing amounts of EtOAc. The fr. eluted with 5% EtOAc in hexane was purified by further chromatography on silica gel (2% Me₂CO in hexane) to give **9** (248 mg). The fr. eluted with 10% EtOAc gave **1** (860 mg), while 15% EtOAc gave **5** (23 mg). The fr. eluted with 20% EtOAc was further subjected to CC on silica gel (hexane–Me₂CO, 9:1) to give **2** (178 mg). Similar treatment of the fr. eluted with 25% EtOAc gave **4** (137 mg), while 30% EtOAc gave **6** (256 mg). The fr. eluted with 35% EtOAc was chromatographed on Sephadex LH-20 (CHCl₃-MeOH. 1:1) to give **8** (29 mg). The fr. eluted with 40% EtOAc showed two major spots on TLC which were sepd by prep. TLC (benzene–CH₂Cl₂–EtOAc, 2:2:1) to give **3** (39 mg) and **7** (36 mg).

Abyssinone V-4'-methyl ether (1). Needles from MeOH, mp 146–147°. [α]_D = 71.5° (MeOH, c 0.1). found [M]⁺ 422.2094; $C_{26}H_{30}O_5$ requires 422.2085. UV λ_{max} (MeOH) nm: 287, 328. CD (MeOH, c 0.045): [Θ]₃₂₈ + 9571, [Θ]₂₈₇ = 32.746. ¹H NMR: Table 1. ¹³C NMR: Table 2. EIMS (probe) 70 eV, m/z (rel. int): 422 [M]⁺ (85), 257 (45), 243 (100), 201 (87). 187 (48), 179 (38), 171 (32), 153 (70), 69 (77).

Abyssinone V(2). Oil. [α]_D 0° (MeOH, c 0.1). UV λ_{max} (MeOH) nm: 285, 328. ¹H NMR: Table 1. ¹³C NMR: Table 2. EIMS (probe) 70 eV, m/z (rel. int): 408 [M] (70), 391 (17), 337 (11), 243 (100), 185 (24), 153 (68), 69 (59).

Burttinone (3). Oil. Found [M]⁻ 438.2044 C₂₆H₃₀O₆ requires 438.2034. [α]_D -67.5° (MeOH, c 0.1). UV λ_{max} (MeOH) nm: 258sh, 288, 329. CD (MeOH; c 0.038): [Θ]₃₂₉ +9916, [Θ]₂₈₈ -34364. ¹H NMR: Table 1. ¹³C NMR: Table 2. EIMS (probe) 70 eV, m/z (rel. int.): 438 [M]⁻ (4), 420 [M -H₂O]⁺, (30), 253 (13), 241 (25), 179 (20), 173 (23), 153 (50), 69 (100).

Calopocarpin (4). Amorphous. Found [M]⁺ 324.1361 C₂₀H₂₀O₄ requires 324.1356. [α]_D – 262.5 (MeOH, c 0.1). UV λ_{max} (MeOH) nm: 238sh, 282sh, 286, 293. CD (MeOH, c 0.035): [Θ]₂₉₃ + 28 706, [Θ]₂₃₈ – 98 896. ¹H NMR identical with lit. [9]. ¹³C NMR (acetone-d₆, 75 MHz): δ 131.9 (d, C-1), 121.1 (s, C-2), 155.8 (s, C-3), 104.0 (s, C-4), 155.0 (s, C-4a), 66.5 (t, C-6), 39.5 (d, C-6a), 119.4 (s, C-6b), 125.0 (d, C-7), 107.6 (d, C-8), 156.8 (s, C-9), 98.3 (d, C-10), 160.7 (s, C-10a), 78.8 (d, C-11a), 112.1 (s, C-11b), 29.2 (t, C-1'), 121.8 (d, C-2'), 134.9 (s, C-3'), 25.8 (q, C-4'), 17.9 (q, C-5'). EIMS (probe) 70 eV, m/z (rel. int): 324 [M]⁺ (100), 269 (39), 147 (7).

Neorautenol (5). Amorphous. [α]_D -243.0° (MeOH, c 0.1). UV λ_{max} (MeOH) nm: 232sh, 282sh, 286, 291sh. ¹H NMR identical with lit. [9]. ¹³C NMR (CDCl₃, 75 MHz): δ 128.3 (d, C-1), 116.4 (s, C-2), 154.6 (s, C-3), 104.7 (s, C-4), 156.3 (s, C-4a), 66.5 (t, C-6), 39.4 (d, C-6a), 119.4 (s, C-6b), 125.0 (d, C-7), 107.6 (d, C-8), 156.9 (s, C-9), 98.4 (d, C-10), 160.7 (s. C-10a), 78.7 (d, C-11a), 112.2 (s, C-11b), 78.7 (s, C-2'), 27.9 (q, 2'-Me), 28.2 (q, 2'-Me), 129.2 (d, C-3'), 121.6 (t, C-4'). EIMS (probe) 70 eV, m/z (rel. int): 322 [M]⁻ (29), 307 [M – Me]⁺ (100), 185 (44), 154 (28).

4'-O-Methylsigmoidin B (6). Amorphous. Found

[M]⁺ 370.1404 $C_{21}H_{22}O_6$ requires 370.1410. [α]_D -73.5° (MeOH, c 0.1). UV λ_{max} (MeOH) nm: 289, 328. CD λ_{max} (MeOH, c 0.039): [Θ]₃₂₈ +10.441, [Θ]₂₈₉ -39.667 nm. ¹H NMR: Table 1. ¹³C NMR: Table 2. EIMS (probe) 70 eV, m/z (rel. int): 370 [M]⁺ (100), 301 (8), 205 (51), 153 (46), 69 (41), 43 (54).

Bidwillon A (7). Colourless oil. [α]_D 0° (MeOH, c 0.1). UV λ_{max} (MeOH) nm: 285, 334. ¹H and ¹³C NMR identical with lit. [12]. EIMS (probe) 70 eV, m/z (rel. int): 409 (100) [M+1]⁺, 408 (83) [M]⁺, 390 (5), 273 (75), 217 (19), 161 (22).

Isobavachalcone (8). Amorphous. UV and ¹H NMR identical to lit [13]. ¹³C NMR (acetone-d₆, 75 MHz): δ 128.3 (s, C-1), 132.4 (d, Hz, C-2/6), 117.4 (d, C-3/5), 161.7 (s, C-4), 116.8 (s, C-1'), 165.8 (s, C-2'), 108.7 (s, C-3'), 163.4 (s, C-4'), 108.7 (d, C-5'), 130.9 (d, C-6', 193.7 (s, C=O), 119.1 (d, C-α), 145.6 (d, C-β), 26.5 (t, C-1"), 124.0 (d, C-2"), 132.1 (s, C-3"), 17.9 (q, Me-4"), 26.5 (q, Me-5"). EIMS (probe) 70 eV, m/z (rel. int): 324 [M]* (100), 281 (60), 269 (18), 149 (66).

Erythrinasinate (9). Amorphous. UV, ${}^{1}H$ and ${}^{13}C$ NMR identical to lit. [14, 15]. EIMS (probe) 70 eV, m/z (rel. int): 586 [M]⁺ (100), 558 (33), 194 (54), 177 (42).

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REFERENCES

- Nkengfack, A. E., Kouam, J., Vouffo, W. T., Fomum, Z. T., Dagne, E., Sterner, O., Browne, L. M. and Ji, G., *Phytochemistry*, 1993, 32, 1305.
- Mears. J. A., in *Chemotaxonomy of the Leg-uminosae* eds. J. B. Harborne, D. Boulter and B. L. Turner, p. 73. Academic Press, London, 1971.
- Gillett, J. B., Polhill, R. M. and Verdcourt, B., Flora of Tropical East Africa, Leguminosae (Part 3) Subfamily Papilionoideae, Whitefriars Press, London, 1971, p. 541.
- Promsattha, R., Tempesta, M. S., Fomum, Z. T. and Mbafor, J. T., J. Nat. Prod., 1988, 51, 611.
- Panichpol, K. and Waterman, P. G., *Phyto-chemistry*, 1978, 17, 1363.
- 6. Gaffield, W., Tetrahedron. 1970, 26, 4093.
- Kamat, V. S., Chuo, F. Y., Kubo, I. and Nakanishi, K., *Heterocycles*, 1981, 15, 1163.
- 8. Gomez-Garibays, F., Quijano, L., Calderon, P. J.

- S., Morales, S. and Rios, T., *Phytochemistry*, 1988, **27**, 2971.
- Ingham, J. L. and Tahara, S., Z. Naturforsch., 1985, 40c, 482.
- 11. Maillard, M., Hamburger, M., Gupta, M. P. and Hostettmann, K., *Planta Med.*, 1989, 55, 281.
- 12. Iinuma, M., Tanaka, T., Mizuno, M., Yama-

- moto, H., Kobayashi, Y. and Yonemori, S., Chem. Pharm. Bull., 1992, 40, 2749.
- 13. Kobayashi, M., Noguchi, H. and Sankawa, U., *Chem. Pharm. Bull.*, 1985, **33**, 3811.
- 14. Fomum, Z. T., Ayafor, J. F., Nkengfack, A. E., Wandji, J. and Fomban, W. G., *Phytochemistry*, 1986, 25, 757.
- Nkengfack, A. E., Sanson, D. R. and Tempesta, M. S., J. Nat. Prod., 1989, 52, 320.