



SEED OIL ALKANES FROM LEGUMINOSAE SPECIES: EVEN-CARBON NUMBER PREFERENCE

ALICIA L. LAMARQUE,* RENÉE H. FORTUNATO† and CARLOS A. GUZMÁN

Cátedra de Química Orgánica. Facultad de Ciencias Exactas, Físicas y Naturales. INBIV, CONICET, Universidad Nacional de Córdoba. Av. Vélez Sarsfield 299. 5000 Córdoba, Argentina; †CONICET at Instituto de Recursos Biológicos, INTA, 1712 Castelar, Buenos Aires, Argentina

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Key Word Index—Caesalpinoideae, Mimosoideae, Papilionoideae, Leguminosae; seed oil alkanes; *n*-alkanes; even-carbon number alkanes.

Abstract—GC and GC-MS have been used to study the distribution of *n*-alkanes in the seed oil from 60 samples of 43 Leguminosae species. The occurrence of high percentages of low *M_n*, even-carbon numbered alkanes was the most important feature in relation to earlier reports on plant hydrocarbons. The main seed oil alkane in almost all species investigated was hexadecane followed by octadecane, eicosane and tetradecane in decreasing order. All samples studied from the three subfamilies presented almost the same pattern with little quantitative and qualitative differences. The presence of this distinctive carbon range of aliphatic paraffins suggests that the inner conditions could lead to a change in chain-length specificity and/or activity of some enzyme systems. © 1998 Elsevier Science Ltd. All rights reserved

INTRODUCTION

The long, straight chain hydrocarbons occurring in higher plants have been studied extensively [1–4]. The alkane distribution patterns have also been determined in species growing in different environmental conditions [5–7] and, in some cases, this fractions has been used as a taxonomic aid [8–10]. These studies relate mainly to hydrocarbons from cuticular waxes.

Seed oil and internal hydrocarbons include a small but significant proportion of alkanes, which may participate in maintaining inner cell structure and may act as energy storage components. However, few investigations of these components have been reported [11, 12].

Although a wide range of alkane patterns differing in their *M_n* distribution has been found in living organisms, odd-carbon numbered *n*-alkanes are the major constituents in higher plants. The biosynthesis of such components has been extensively investigated [13–20].

All the cited cases, which report no alternation of odd and even-carbon number alkanes or flatter distribution with an increase in the relative proportion of the even-carbon number members of the series, were obtained from prolonged extraction of leaf waxes or other tissues or from macerated plant material. In those cases the intracellular lipids would have been included in the lipid fractions examined [21–23].

At present, there are no studies of the Leguminosae or related families, where the even-carbon numbered alkanes from seed lipids are dominant.

The findings presented here are concerned with the unusual composition of the seed oil alkanes of 43 species belonging to the three subfamilies of the Leguminosae, as revealed by the application of IR, GC and GC-MS techniques.

RESULTS AND DISCUSSION

The alkane composition of the seed oils of the species studies (Table 1), is presented in Table 2. Nineteen *n*-alkanes were identified. IR spectra confirmed their identity as paraffins (2825, 2854, 1481 and 1375 cm^{-1}), and the lack of absorption at 1720–1740 cm^{-1} , 3600–3400 cm^{-1} or 1100–1250 cm^{-1} confirmed the absence of carbonyl, hydroxy or internal oxygen-containing components.

The mass spectra of the *n*-alkanes showed for each $[M]^+$ at *m/z* 198, 209, 226, 239, 254, 267, 281, 295, 309, 323, 337, 351, 365, 379, 393, 407, 421, 435 and 449 which correspond to C_{14} – C_{32} respectively. There was no evidence of branched or unsaturated components.

Qualitative and quantitative variations were observed among the species. The main features of the alkane distribution patterns were as follows: (i) The even-numbered compounds exceeded their odd-carbon neighbours (62.8–99.2% of total *n*-alkanes). (ii)

* Author to whom correspondence should be addressed.

Table 1. Collection data (Caesalpinoideae, Mimosoideae and Papilionoideae Subfamilies)

Taxon	Voucher No	Population site
Caesalpinoideae Subfamily		
<i>Caesalpinieae</i> Tribe		
<i>Caesalpinia gilliesii</i> Wall. ex Hook (1)	RHF 5030 (BAB)	
<i>Caesalpinia gilliesii</i> (2)	BAB 91963	Córdoba : Capital
<i>Caesalpinia gilliesii</i> (3)	BAB 91953	Córdoba : Mina Clavero
<i>Cercidium praecox</i> (R. et P) Harms subsp. <i>praecox</i>	RHF 5036 (BAB)	Santiago del Estero : 20 km from Sumampa
<i>Cassieae</i> Tribe		
<i>Senna alata</i> L.	CTES 29436	Corrientes : Capital
<i>Chamaecrista venturiana</i> Irwin et Barneby	RHF 5377 (BAB)	Bolivia
<i>Cercideae</i> Tribe		
<i>Bauhinia forficata</i> Link subsp. <i>pruinosa</i> (Benth.) Fortunato et Wunderlin.	BAB 91931	Córdoba : Capital
Mimosoideae Subfamily		
<i>Mimoseae</i> Tribe		
<i>Prosopis alba</i> Griseb. (1)	18'88	Santiago del Estero :
<i>Prosopis alba</i> (2)	532'90	Lat. 28°57' Long. 66°57'
<i>Prosopis nigra</i> (Griseb.) Hier. (1)	62'88	Lat. 25°05' Long. 64°09'
<i>Prosopis nigra</i> (2)	68'88	Salta
<i>Prosopis ferox</i> Griseb.	76'89	Salta : Bosque de Tintin
<i>Prosopis caldenia</i> Burkart	542'90	Lat. 32°20' Long. 65°10'
<i>Prosopis kuntzei</i> Harms	k09	Loreto : Río Namby
<i>Prosopis flexuosa</i> DC.	575'91	La Rioja : Tinogasta
<i>Prosopis argentina</i> Burkart	38'88	Mendoza : Tucunucu
<i>Anadenanthera macrocarpa</i> Humb. et Bonpl. ex Willd.	BAB 91929	Salta : Capital
<i>Anadenanthera colubrina</i> (Vell.) Brenam	CTES 29437	Corrientes : Capital
<i>Parapiptadenia excelsa</i> (Griseb.) Burkart f. <i>jujuyensis</i> Burkart	BAB 91927	Salta : Capital
<i>Desmanthus virgatus</i> (L.) Willd.	BAB 91962	Córdoba : Saldán
<i>Acacieae</i> Tribe		
<i>Acacia caven</i> (Mol.) Mol. var. <i>caven</i> (1)	CORD 186	Córdoba : La Calera
<i>Acacia caven</i> (2)	BAB 91947	Córdoba : Villa Allende
<i>Acacia aroma</i> Gill. ex Hook. (1)	CORD 188	Córdoba : Route 28, km 873
<i>Acacia aroma</i> (2)	CORD 191	San Juan : Route 12, km 87
<i>Acacia aroma</i> (3)	CORD 196	San Juan : Route 12, km 66
<i>Acacia aroma</i> (4)	BAB 91936	Córdoba : Villa Allende
<i>Acacia aroma</i> (5)	BAB 91948	Córdoba : Villa Allende
<i>Acacia furcatispina</i> Burkart (1)	CORD 187	San Juan : Route 12, km 87
<i>Acacia furcatispina</i> (2)	BAB 91930	Córdoba : El Diquecito
<i>Acacia atramentaria</i> Benth. (1)	BAB 91932	Córdoba : Capital
<i>Acacia atramentaria</i> (2)	BAB 91930	Córdoba : Route 38, km 49
<i>Acacia parviceps</i> (Speg.) Burkart	BAB 91928	Salta : Capital
<i>Acacia horrida</i> (L.) Willd.	BAB 91946	Chile : Route 5, km 80
<i>Ingeae</i> Tribe		
<i>Inga uruguensis</i> Hook. et Arn.	CTES 29439	Chaco : Río Paraná
<i>Enterolobium contortisiliquum</i> (Vell.) Morong	BAB 91925	Salta : Rosario de Lerma
Papilionoideae Subfamily		
<i>Dalbergieae</i> Tribe		
<i>Tipuana tipu</i> (Benth.) Kuntze	BAB 91954	Córdoba : Capital
<i>Robinieae</i> Tribe		
<i>Sesbania punicea</i> (Cav.) Benth	BAB 91933	Córdoba : Capital
<i>Indigoferae</i> Tribe		
<i>Indigofera parodiana</i> Burkart	RHF 5338 (BAB)	Córdoba : Río Tercero
<i>Indigofera suffruticosa</i> Mill.	RHF 5391 (BAB)	Bolivia : 5 km from Coroico
<i>Desmodieae</i> Tribe		
<i>Desmodium cuneatum</i> Hook. et Arn.	RHF 5341 (BAB)	Córdoba : Villa La Bolsa

Table 1—continued.

Taxon	Voucher No	Population site
<i>Desmodium tortuosum</i> (Sw) DC	RHF 5334 (BAB)	Córdoba: Parador El Condor
<i>Desmodium neomexicanum</i> Gray	BAB 91951	Córdoba: Mina Clavero
<i>Phaseoleae</i> Tribe		
<i>Collaea speciosa</i> (Lois.) DC.	RHF 5373 (BAB)	Bolivia
<i>Galactia latisilicua</i> Desv. (1)	BAB 91934	Córdoba: Capital
<i>Galactia latisilicua</i> (2)	BAB 91956	Córdoba: Carlos Paz
<i>Galactia latisilicua</i> (3)	BAB 91971	Córdoba: Villa Los Aromos
<i>Cologania ovalifolia</i> H.B.K.	BAB 91920	Salta: Route to Corralito
<i>Centrosema virginianum</i> (L.) Benth. (1)	BAB 91924	Salta: Campo Quijano
<i>Centrosema virginianum</i> (2)	BAB 91973	Córdoba: Agua de Oro
<i>Rhynchosia edulis</i> Griseb. (1)	BAB 91966	Córdoba: Cabalango
<i>Rhynchosia edulis</i> (2)	BAB 91970	Córdoba: Villa Los Aromos
<i>Rhynchosia edulis</i> (3)	BAB 91955	Córdoba: Capital
<i>Rhynchosia edulis</i> (4)	BAB 91974	Córdoba: Agua de Oro
<i>Rhynchosia senna</i> Hook. et. Arn.	BAB 91965	Córdoba: Mina Clavero
<i>Aeschynomeneae</i> Tribe		
<i>Aeschynomene rudis</i> Benth.	RHF 5375 (BAB)	Bolivia: Coroico
<i>Stylosanthes montevidensis</i> Vogel.	RHF 5332 (BAB)	Córdoba: Altas Cumbres
<i>Stylosanthes gracilis</i> H.B.K.	RHF 5336 (BAB)	Córdoba: Río Tercero
<i>Adesmieae</i> Tribe		
<i>Adesmia volckmanni</i> Phil.	RHF 5215 (BAB)	Neuquén: 38 km from Junín de los Andes. Route 40
<i>Galegeae</i> Tribe		
<i>Glycyrrhiza astragalina</i> Gill. ex Hook. et Arn.	RHF 5213 (BAB)	Neuquén: Junín de los Andes
<i>Vicieae</i> Tribe		
<i>Lathyrus magellanicus</i> Lam. var. <i>glauescens</i> Speg.	RHF 4889 (BAB)	Tierra del Fuego

n-Hexadecane was the principal alkane in all species except in *Cologania ovalifolia* where *n*-octadecane was the dominant component. (iii) *n*-Octadecane, *n*-eicosane and *n*-tetradecane were present in significant amounts. (iv) Some species of *Acacia* (*A. aroma*, *A. atramentaria* and *A. parviceps*) presented a flatter distribution, by way of an increase in the odd-carbon numbered and long chain components. This feature was also encountered in *Sesbania punicea*. (v) The genus *Rhynchosia* (Phaseoleae tribe) was characterized by the presence of significant levels of dotriacontane (4.5–7.2%), an alkane not found in the other groups.

No variation was found in the alkane patterns of the same species originating from different localities. This feature could demonstrate the predominance of genetic factors over external conditions in these species.

Earlier investigation on the unsaponifiable matter from some species of this plant family found a mixture of C₁₆–C₃₇ *n*-alkanes, with the ratio of odd to even number alkanes much less than that found in other higher plants [11]. Our results are in agreement with these findings, but the ratios found by us were much lower.

It has been postulated that two pathways must operate in the biosynthesis of hydrocarbons. As well

as the constitution of epicuticular waxes varying between species [8–10, 23], as a result of plant age [24], and of the environmental conditions in which plants grow or to which they are exposed [1, 5, 7, 25], the inner tissue conditions could lead to a change in chain-length specificity and/or activity of some enzyme systems. Our results suggest that the preponderance of even and short carbon numbered alkanes in the seed oils of Leguminosae could be strongly tissue dependent, apparently as a result of tissue-specific differences in alkane biosynthesis.

Finally, the presence of this class of alkanes in Leguminosae seed oils suggests that this plant material should provide a good source for studying short chain alkane biosynthesis.

EXPERIMENTAL

Plant material

Details of plant material are given in Table 1. Voucher specimens are retained at the Herbarium of the Instituto de Recursos Biológicos, INTA, Castelar (BAB), at the Instituto de Botánica del Nordeste, Corrientes (CTES), at the Herbarium of the Museo Botánico (Facultad de Ciencias Exactas, Físicas y Naturales of the Universidad Nacional de Córdoba)

Table 2. Alkane composition (% total) of seed oils from Caesalpinoideae, Mimosoideae and Papilionoideae (Leguminosae) samples

Taxon	C14	C15	C16	C17	C18	C19	C20	C21	C22	C23	C24	C25	C26	C27	C28	C29	C30	C31	C32
<i>Caesalpinoideae</i>																			
<i>Caesalpinia gilliesii</i> (1)	10.3	tr	30.8	tr	24.4	5.0	14.9	tr	10.9	tr	3.2	tr	tr	tr	tr	tr	tr	tr	—
<i>Caesalpinia gilliesii</i> (2)	10.5	tr	30.6	tr	24.2	4.9	14.7	tr	11.1	tr	3.2	tr	tr	tr	tr	tr	tr	tr	—
<i>Caesalpinia gilliesii</i> (3)	10.5	tr	31.0	tr	24.7	4.7	15.1	tr	10.6	tr	3.2	tr	tr	tr	tr	tr	tr	tr	—
<i>Cercidium praecox</i>	8.4	1.9	26.2	2.2	20.3	3.5	11.6	tr	3.3	tr	3.5	2.1	5.5	9.2	tr	1.8	tr	tr	—
<i>Senna alata</i>	12.8	tr	26.2	tr	23.1	tr	11.8	tr	10.5	tr	6.8	tr	3.0	tr	5.0	tr	tr	tr	—
<i>Chamaecrista venturiana</i>	11.4	tr	30.5	tr	21.9	tr	11.9	tr	7.4	tr	6.5	3.0	3.8	1.7	0.9	0.6	tr	tr	—
<i>Bauhinia forficata</i>	16.2	tr	22.9	tr	14.4	tr	7.7	tr	6.0	tr	4.8	5.0	4.0	4.1	8.4	6.3	tr	tr	—
<i>Mimosoideae</i>																			
<i>Prosopis alba</i> (1)	12.0	0.6	28.1	tr	18.7	2.3	10.7	1.7	8.5	tr	3.0	tr	3.3	1.6	4.6	3.1	1.5	tr	—
<i>Prosopis alba</i> (2)	12.7	0.5	30.4	0.9	21.5	0.6	8.1	0.5	7.3	tr	3.6	tr	3.2	2.2	1.5	4.6	2.1	tr	—
<i>Prosopis nigra</i> (1)	11.7	1.3	32.8	1.0	24.1	0.8	8.6	0.9	8.7	tr	3.7	tr	2.8	tr	0.9	0.9	1.1	tr	—
<i>Prosopis nigra</i> (2)	18.7	1.0	28.7	0.8	18.5	0.4	11.8	1.2	11.1	0.4	3.1	0.4	1.6	0.8	0.4	0.6	0.4	tr	—
<i>Prosopis ferox</i>	5.4	0.3	27.2	0.3	26.2	0.1	16.5	tr	9.6	tr	5.9	tr	4.0	1.3	1.8	1.1	tr	tr	—
<i>Prosopis caldenia</i>	13.0	1.4	33.2	0.9	21.4	1.3	12.3	0.9	6.5	tr	3.4	tr	1.8	1.3	1.1	1.2	tr	tr	—
<i>Prosopis kuntzei</i>	6.9	1.2	26.4	1.6	23.3	1.5	13.8	1.2	7.9	1.6	4.6	1.0	2.8	1.0	4.4	0.6	tr	tr	—
<i>Prosopis flexuosa</i>	10.9	1.5	23.9	1.9	20.1	1.2	12.1	1.1	6.3	0.1	3.5	1.5	2.6	7.3	2.7	2.6	0.4	tr	—
<i>Prosopis argentina</i>	10.0	1.0	27.8	1.3	23.0	0.9	15.2	0.6	8.9	tr	5.0	tr	2.7	tr	1.6	tr	1.6	tr	—
<i>Anadenanthera macrocarpa</i>	10.8	3.6	19.2	1.9	19.1	6.7	12.4	1.6	6.5	1.4	4.1	1.6	2.1	2.6	2.7	1.7	1.9	tr	—
<i>Anadenanthera colubrina</i>	10.8	4.0	29.2	5.1	21.1	3.5	10.8	1.6	6.2	tr	3.4	tr	2.1	0.5	1.3	tr	0.1	tr	—
<i>Parapiptadenia excelsa</i>	6.9	1.6	19.1	2.3	14.1	6.3	8.6	2.1	3.9	1.6	2.0	6.2	5.6	6.1	3.9	5.3	4.3	tr	—
<i>Desmanthus virgatus</i>	11.8	1.5	28.9	1.8	21.9	1.3	13.3	1.1	7.9	tr	4.9	tr	2.7	0.4	1.5	0.1	0.7	tr	—
<i>Acacia caven</i> (1)	10.6	1.9	28.5	1.5	23.1	1.3	12.9	1.3	7.3	0.4	3.5	0.2	2.0	1.5	1.4	1.9	0.6	tr	—
<i>Acacia caven</i> (2)	10.9	1.8	28.7	1.5	22.8	1.3	12.4	1.3	7.5	0.4	3.8	0.2	2.2	1.5	1.3	1.6	0.7	tr	—
<i>Acacia aroma</i> (1)	8.1	1.5	18.3	3.9	12.8	3.6	7.8	0.3	4.9	0.2	3.2	1.2	3.0	6.7	10.8	9.8	3.8	tr	—
<i>Acacia aroma</i> (2)	8.2	1.5	18.3	3.8	12.9	3.6	7.9	0.3	4.9	0.2	3.2	1.2	3.1	6.5	10.7	9.9	3.7	tr	—
<i>Acacia aroma</i> (3)	8.0	1.5	18.5	3.7	13.5	3.5	7.6	0.3	4.7	0.2	3.3	1.2	3.2	6.7	10.9	9.5	3.6	tr	—
<i>Acacia aroma</i> (4)	8.3	1.5	18.9	3.6	12.8	3.7	7.8	0.3	4.8	0.3	3.2	1.2	3.0	6.9	10.5	9.3	3.8	tr	—
<i>Acacia aroma</i> (5)	8.1	1.6	18.1	3.5	12.7	3.6	7.9	0.3	4.9	0.3	3.1	1.2	3.2	6.8	10.8	9.9	3.9	tr	—
<i>Acacia furcatispina</i> (1)	11.9	1.6	26.5	1.4	22.4	1.5	13.0	1.1	7.8	0.3	5.1	0.3	3.0	1.0	1.1	1.0	0.9	tr	—
<i>Acacia furcatispina</i> (2)	11.8	1.6	27.0	1.5	22.0	1.5	13.0	1.1	7.7	0.3	5.2	0.3	3.1	1.0	1.0	1.0	0.8	tr	—
<i>Acacia atramentaria</i> (1)	6.9	1.2	15.5	1.6	10.9	1.9	7.5	2.3	4.7	1.9	4.6	4.3	3.5	6.4	12.6	9.2	4.9	tr	—
<i>Acacia atramentaria</i> (2)	7.2	1.3	15.8	1.6	10.7	1.8	7.3	2.3	4.5	1.8	4.7	4.4	3.6	6.7	12.3	9.1	4.8	tr	—
<i>Acacia parviceps</i>	7.6	0.1	17.4	0.2	15.7	0.2	9.9	0.5	6.6	0.3	3.5	9.2	6.2	9.4	4.2	8.9	tr	tr	—
<i>Acacia horrida</i>	16.7	0.6	27.6	0.5	19.5	0.5	11.4	0.5	7.3	tr	4.4	0.1	2.7	2.7	2.5	1.9	0.9	tr	—
<i>Inga uruguensis</i>	21.7	1.8	28.9	1.3	19.3	1.0	10.7	1.2	6.1	1.1	3.0	tr	1.5	tr	0.7	1.0	0.4	tr	—
<i>Enterolobium contortisiliquum</i>	11.2	1.1	25.2	1.8	18.8	2.0	11.8	2.1	7.4	0.2	5.7	0.4	3.9	1.1	7.1	tr	tr	tr	—
<i>Tipuana tipu</i>	18.9	0.8	32.4	0.7	20.2	0.4	10.9	0.5	6.2	tr	3.3	tr	2.6	0.4	0.9	0.6	0.9	tr	—
<i>Sesbania punicea</i>	10.7	2.3	16.2	2.0	9.0	2.5	5.9	2.5	4.2	4.0	3.8	6.7	5.2	10.3	6.9	6.8	0.9	tr	—
<i>Indigofera parodiana</i>	6.9	2.1	25.2	1.2	22.7	0.9	16.8	0.9	10.2	tr	5.5	tr	3.1	tr	2.6	tr	1.6	tr	—
<i>Indigofera suffruticosa</i>	19.8	0.4	33.2	0.5	21.0	tr	11.3	0.2	5.7	tr	2.7	tr	1.4	1.5	0.9	1.2	tr	tr	—
<i>Desmodium cuneatum</i>	12.1	0.4	27.7	0.4	22.9	0.3	12.9	0.3	7.4	tr	4.6	tr	3.1	0.5	2.1	2.1	1.2	1.9	—
<i>Desmodium tortuosum</i>	8.9	0.6	28.5	0.5	23.3	0.3	13.8	0.3	7.9	0.1	4.5	0.1	2.6	2.1	1.9	1.9	1.8	0.9	—
<i>Desmodium neomexicanum</i>	12.1	0.5	28.0	0.5	23.1	0.3	11.9	0.3	7.6	0.1	4.6	0.1	2.7	1.3	2.0	2.0	1.6	1.3	—
<i>Collaea speciosa</i>	14.7	2.2	31.0	1.3	19.9	1.2	11.7	0.4	6.7	tr	3.4	tr	1.9	tr	1.2	0.5	1.1	2.6	—
<i>Galactia latisilicua</i> (1)	8.9	0.2	27.7	0.3	24.6	0.1	14.8	0.2	8.0	tr	5.2	tr	4.3	0.5	1.6	3.0	0.5	tr	—
<i>Galactia latisilicua</i> (2)	8.5	0.1	27.9	0.3	24.2	0.1	14.6	0.1	8.3	tr	5.7	tr	3.8	0.8	1.9	2.9	0.5	tr	—
<i>Galactia latisilicua</i> (3)	8.6	0.1	26.8	0.3	24.2	0.1	14.0	0.2	8.1	tr	4.8	tr	5.0	0.3	3.2	3.0	0.5	tr	—
<i>Cologania ovalifolia</i>	14.4	0.8	15.6	1.0	33.1	1.1	12.2	0.5	7.3	tr	4.3	tr	2.9	2.9	1.8	1.8	tr	tr	—
<i>Centrosema virginianum</i> (1)	7.0	0.3	27.6	0.3	23.9	0.2	15.2	0.1	9.9	tr	6.2	tr	3.6	0.6	2.0	0.6	1.3	1.1	—
<i>Centrosema virginianum</i> (2)	6.7	2.4	26.9	2.8	21.2	1.9	12.8	0.6	8.8	tr	6.1	tr	3.9	1.1	2.4	0.8	1.3	tr	—
<i>Rhynchosia edulis</i> (1)	11.5	0.7	23.4	0.9	15.9	0.3	10.3	0.3	7.4	tr	5.3	tr	3.5	4.2	7.2	1.7	tr	tr	7.0
<i>Rhynchosia edulis</i> (2)	13.4	1.0	24.6	0.9	17.9	0.5	10.6	0.3	6.4	tr	4.4	tr	3.2	4.5	3.3	1.5	tr	tr	7.2
<i>Rhynchosia edulis</i> (3)	11.3	0.4	26.8	0.3	20.1	0.3	12.4	0.2	7.5	tr	4.0	tr	2.2	2.6	2.6	2.1	1.7	tr	5.2

Table 2—continued.

<i>Rhynchosia edulis</i> (4)	10.5	1.8	26.9	1.8	20.0	1.5	6.3	1.1	9.8	tr	5.6	tr	3.3	1.6	1.8	1.6	1.7	tr	4.5
<i>Rhynchosia senna</i>	10.4	0.2	26.5	0.2	20.3	0.1	11.8	0.1	6.3	tr	3.2	tr	1.6	5.5	2.9	2.5	1.2	tr	6.9
<i>Aeschynomene rudis</i>	20.3	1.6	31.7	1.0	19.1	0.8	10.4	0.9	5.9	tr	3.5	tr	2.5	tr	1.3	tr	0.8	tr	—
<i>Stylosanthes montevidensis</i>	6.8	0.7	26.3	1.5	25.2	1.0	16.3	0.8	9.7	tr	5.2	tr	3.0	tr	2.0	1.2	tr	tr	—
<i>Stylosanthes gracilis</i>	10.7	1.0	26.7	1.5	21.5	0.7	13.4	0.5	8.3	tr	4.8	tr	2.7	1.5	2.9	3.3	0.4	tr	—
<i>Adesmia volckmanni</i>	9.6	0.9	29.2	1.5	23.8	0.9	14.1	0.9	8.0	tr	4.3	tr	2.3	1.0	1.8	0.8	0.6	tr	—
<i>Glycyrrhiza astragalina</i>	14.5	1.0	28.3	1.0	19.2	0.9	10.7	0.7	8.4	tr	6.1	tr	3.9	0.2	2.3	1.6	0.9	tr	—
<i>Lathyrus magellanicus</i>	10.6	2.6	28.9	2.6	23.5	1.8	13.1	0.2	7.4	tr	4.2	tr	2.2	1.2	1.2	0.2	tr	tr	—

tr, trace less than 0.1%.

(CORD), and *Prosopis* specimens are deposited at the National Genebank of *Prosopis* (Facultad de Ciencias Agropecuarias, Universidad Nacional de Córdoba) (BNG).

Extraction of oil

Clean and dried samples of seeds were ground in a disintegrator. The powdered seeds were extracted with *n*-hexane in a Soxhlet apparatus for 12 h. The extracted oils were filtered and the solvent removed under reduced pressure in a rotary evaporator at 40°.

Isolation of unsaponifiable matter

The unsaponifiable fraction was obtained by direct saponification of the total lipids using methanolic KOH (IM) and recovered with CHCl₃.

Alkane isolation

The CHCl₃ soln of the unsaponifiable fraction was applied uniformly along a line 1.5 cm from one edge of (20 × 20 cm) 2 mm thickness plates coated with silica gel 60 G and developed three times with hexane–Et₂O (9:1) as developing solvent. The hydrocarbon zone was scraped from the plates and extracted with *n*-hexane. Further purification of the alkane fraction was carried out using AgNO₃–silica gel (1:9) prep. TLC using *n*-hexane–Et₂O (19:1) as developing solvent. IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 2825, 2854, 1481 and 1375.

GC, GC-MS

GC was performed using a CBPI (25 m × 0.25 mm i.d.) column programmed from 100°–300° at 5° min⁻¹ using a FID with N₂ as carrier gas.

Mass spectrometric analysis was performed using a PERKIN ELMER Q MASS 910 gas chromatograph-mass spectrometer. A SE 30 (30 m × 0.25 mm i.d.) column was used with He (0.8 ml min⁻¹) as carrier gas and was temperature programmed from 60°–280° at 4° min⁻¹. Injector temperature at 290°. All spectra were obtained at 70 eV and the ion source temperature was 200°, with a filament current of 1 mA. Each peak was scanned over the range of 10–500 mass units.

The alkanes were identified by their IR spectra, comparisons of their gas chromatographic *R*_s with respect to reference samples run under identical conditions and by their respective mass spectrometric fragmentation patterns.

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