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STEROIDAL ALKALOID GLYCOSIDES FROM SOLANUM ORBIGNIANUM

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Abstract—Aerial parts of *Solanum orbignianum* afforded a new steroidal alkaloid glycoside, leptinidine 3–O– β –D-glucopyranoside, together with the known alkaloids leptinidine, leptinine I and leptinine II. Their structures were established by spectroscopic methods. © 1998 Elsevier Science Ltd. All rights reserved

INTRODUCTION

Solanum species have already been studied chemically and found to contain mixtures of steroidal alkaloid glycosides. From the aerial parts of Solanum orbignianum Sendt., along with leptinidine (1), we have isolated three steroidal alkaloid glycosides, leptinidine $3-O-\beta$ -D-glucopyranoside (2), leptinine I (3) and leptinine II (4). Compound 2 is a new steroidal alkaloid glycoside. Leptinines I and II were previously isolated from Solanum chacoense [1], and their structures deduced on the basis of acid hydrolysis and biogenetic analysis [2]; their spectroscopic data are unknown. In this paper, we report the isolation of the compounds 1-4, and their structures elucidation by spectroscopic methods, including two-dimensional NMR techniques.

RESULTS AND DISCUSSION

Leptinidine (1) was identified by its mass spectrum (EIMS) and mainly by comparison of its ¹³C NMR spectrum (Table 2) with literature values [3].

Compound 2 was obtained as an amorphous powder and gave ion peaks at m/z 576 $[M+H]^-$ and m/z 412 $[M-C_6H_{11}O_5]^+$, in the FAB mass spectrum, suggesting a glycosidic structure. Peaks at m/z 413, m/z 166 and m/z 220 in the EIMS spectrum, were diagnostic fragments for a solanidane skeleton containing a hydroxyl in either rings E or F [4]. The ¹H and ¹³C NMR spectra (Tables 1 and 2) indicated that the aglycone was leptinidine [3]. The chemical shifts

for the aglycone moiety of 2 agreed with those of 1, except for C-3 ($\delta_{\rm H}$ 3.95, $\delta_{\rm C}$ 78.5). The ¹³C NMR spectrum of 2 permitted the assignment of the sugar moiety linked at C-3 of the aglycone. The sugar moiety of 2 was determined as β -D-glucopyranose, based on the chemical shifts of the carbons, together with the coupling constant ($J_{1.2}$ =7.8 Hz) for the anomeric proton. This was confirmed by acid hydrolysis of 2, which yielded glucose as sugar component and leptinidine (1), as aglycone. The ¹H and ¹³C NMR spectroscopic signals of 2 were assigned by ¹H-¹H 2D COSY, HETCOR and DEPT experiments. Thus, compound 2 was characterized as leptinidine 3-O- β -D-glucopyranoside.

The solanidane alkaloid glycosides 3 and 4 were identified by their FAB mass, ¹H and ¹³C NMR (Tables 1 and 2) spectral data. Assignment of the proton and carbon resonances of 3 and 4 were achieved by a combination of ¹H-¹H 2D COSY, HETCOR and DEPT experiments. The FAB mass spectra of 3 gave peaks at m/z 868 $[M+H]^+$, m/z 722 [868–146(rhamnose)], m/z 576 [722–146(rhamnose)] and m/z 414 [576–162(hexose)], indicating a triglycoside structure. The solanidane skeleton was characterized by the presence of peaks at m/z 166 and m/z220 in the EIMS spectra [4]. Acid hydrolysis of 3 gave rhamnose and glucose as sugar components, and leptinidine (1) as aglycone. The ¹H and ¹³C NMR spectral data (Tables 1 and 2) for the aglycone moiety of 3 were similar to those of 1, except for C-3 ($\delta_{\rm H}$ 3.85, $\delta_{\rm C}$ 77.8), indicating linkage of the sugar moiety at that position. The anomeric proton signals in the ¹H NMR spectra (Table 1), permitted assignment of the orientation of the rhamnopyranosyl units and glucopyranosyl unit to be α and β , respectively. The

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Table 1. HNMR spectral data for compounds 1-4 (values in parentheses are J in Hz; δ in pyridine- d_{δ})

Н	1	2	3	4
Aglycone moieties				
3	3.55 m	3.95*	3.85 m	3.96*
6	5.35 d(4.9)	5.33 d (4.9)	5.32 d(5.0)	5.32 d (5.2)
16	2.76 m	2.68 m	2.77*	2.76 m
18	0.83 s	$0.92 \ s$	0.94 s	0.94 s
19	1.03 s	0.84 s	$0.98 \ s$	0.98 s
21	0.98 d(6.6)	1.00 d (6.6)	1.00 d (6.2)	1.00 d(6.5)
23	3.95 m	3.95*	3.98 m	3.96*
6 <i>eq</i>	2.89 dd (8.0, 3.0)	2.93 dd (8.0,3.2)	3.06 d (7.8)	3.10 d (9.1)
6ax	1.50 m	1.41*	1.50*	1.50 m
27	0.84 d (6.3)	0.81 d (6.6)	0.82 d (6.2)	0.81 d (6.5)
Sugar moieties				
nner-Glucose				
		5.04 d (7.8)	4.97 d(8.3)	
		4.06 dd (7.8, 8.2)	4.23*	
		4.29 dd (8.2, 8.9)	4.23*	
		4.27 t (9.0)	4.39*	
		3.95*	3.63	
		4.41 dd (5.4,12.0)	4.08 dd (3.2, 12.5)	
		4.56 dd (2.1, 12.0)	4.20	
-O-Rhamnose				
			5.85 brs	
			4.68 dd (3.2, 1.7)	
			4.54 dd (3.2, 9.3)	
			4.35 dd (9.3, 9.4)	
			4.94 dd (9.4, 6.1)	
			1.61 d (6.1)	
-O-Rhamnose				
			6.40 brs	6.29 brs
			4.83 dd (3.2, 1.6)	4.90*
			4.63 dd (3.2, 9.3)	4.61 dd (3.0, 9.1)
			4.37*	4.35*
			4.94 dd (9.4, 6.1)	4.90*
			1.74 d (6.1)	1.70 d (6.1)
ner-Galactose				
				4.92 d (8.3)
				4.70 t (8.8)
				4.80 d (2.4)
O-Glucose				
				5.18 d (7.6)
				3.94 d (7.6)
				4.48 d (10.1)

^{*} Overlapping with other signals

sequence and connectivities of the sugar moiety were determined by comparing the ¹³C NMR spectral data (Table 2) with those reported for solamargine [5]. Thus, the structure of 3 was determined as leptinidine $3-O-[\alpha-L-rhamnopyranosyl-(1 \rightarrow 4)-[\alpha-L-rhamnopyranosyl-(1 \rightarrow 2)]-\beta-D-glucopyranoside].$

The mass spectral data of 4 suggested a triglycoside solanidane alkaloid, due the presence of peaks at m/z 884 [M + H]⁺, and m/z 414 in the FAB mass spectrum,

and m/z 220 and m/z 166 [4] in the EIMS spectrum. Acid hydrolysis of 4 gave glucose, galactose and rhamnose as sugar components, and leptinidine (1) as aglycone. The ¹H and ¹³C NMR data (Tables 1 and 2) for the aglycone moiety of 4 agreed with those of 1, except for C-3 ($\delta_{\rm H}$ 3.96, $\delta_{\rm C}$ 78.2). This confirmed the aglycone to be lepitinidine (1), and indicated that the sugar moiety was linked at C-3. The ¹³C NMR spectroscopic signals (Table 2), together with coupling constants for

Table 2. ¹³C NMR spectral data for compounds 1-4 (δ values in pyridine- d_5)

C	1	2	3	4
Aglycone moieties				
1	37.2	37.3	36.9	37.3
2	31.5	30.1	29.9	29.9
3	71.6	78.5	77.8	78.2
4	42.2	39.2	38.9	38.5
5	140.9	140.9	140.9	140.9
6	121.5	121.8	121.8	121.7
7	31.9	32.2	31.0	30.8
8	31.5	31.6	31.5	31.5
9	50.2	50.2	50.2	50.1
10	36.5	36.8	37.3	36.9
11	20.7 40.3	20.9 40.8	20.8 4 0.7	20.8 40.6
12 13	40.9	41.0	40.9	40.6
14	57.6	57.6	57.5	57.5
15	32.0	31.5	32.2	32.2
16	69.3	69.5	69.3	69.2
17	62.1	62.5	62.2	62.1
18	16.5	16.7	16.6	16.5
19	19.3	19.2	19.1	19.1
20	30.5	30.7	30.8	30.8
21	18.4	18.3	18.0	17.8
22	78.2	78.4	78.4	78.3
23	65.1	64.8	64.6	64.5
24	39.5	39.7	39.7	39.7
25	25.3	25.5	25.3	25.2
26	60.2	60.5	60.2	60.2
27	18.9	19.2	19.1	19.1
Sugar moieties				
Inner-Glucose		102.5	100.2	
1		102.5 75.3	100.2 78.0 ^a	
2 3		73.3 78.5	78.0 77.6ª	
4		71.6	78.4	
5		78.0	76.8	
6		62.2	61.1	
V		02.2	VI.I	
4-O-Rhamnose			102.0	
1			102.9	
2			72.6 72.7	
3 4			72.7 73.7 ^b	
5			70.2°	
6			18.2	
· ·			10.2	
2-O-Rhamnose				
1			101.9	102.1
2			72.3	72.3
3			72.3	72.6
4			73.9 ^b	73.9
5			69.3°	69.6
6			18.4	18.4
Inner-Galactose				
1				100.3
2				76.3
3				84.7
4				70.1
5				74.7
6				62.5

continued

Table 2-continued.

С	ì	2	3	4
3-O-Glucose				
1				105.7
2				74.7
3				77.4ª
4				71.4
5				78.3ª
6				62.5

a-c May be exchanged.

2:
$$R = HO OH OH$$

3:
$$R = \frac{\text{HO}}{\text{HO}} \frac{\text{OH}}{\text{OH}}$$

4:
$$R = \frac{\text{HO}}{\text{HO}} = \frac{\text{OH}}{\text{OH}} = \frac{\text$$

the anomeric protons (Table 1), indicated that the sugar moiety was consisted of a β -D-glucopyranosyl unit, a β -D-galactopyranosyl unit and a α -L-rhamnopyranosyl unit. Finally, connections of the sugar components were determined by comparison of the ¹³C NMR signals with those of solasonine [6]. Therefore, compound 4 was determined to be leptinidine 3-O-

 $[\beta-D$ -glucopyranosyl- $(1 \rightarrow 3)$ - $[\alpha-L$ -rhamnopyranosyl- $(1 \rightarrow 2)$]- β -D-galactopyranoside].

EXPERIMENTAL

General

Mps: uncorr. ¹H NMR (300 MHz) and ¹³C NMR (75.5 MHz) were recorded on a Varian Gemini 2000 spectrometer, in pyridine– d_5 or CDCl₃ with TMS as int. standard. IR spectra were recorded as KBr pellets on a Jasco model IR 700 spectrometer. FABMS were taken in the positive mode on a Fison model Quattro GC-MS spectrometer, using glycerol as matrix. TLC: Silica gel GF₂₅₄ (0.25 mm thick). CC: Silica gel 60 (0.0063–0.200 mesh, Merck); alumina 90 (70–230 mesh ASTM, Merck).

Plant material

Aerial parts of *S. orbignianum* were collected in August 1994 in Porto Rico, State of Paraná, Brazil. A voucher specimen (HUM2483) is deposited at the herbarium of the Universidade Estadual de Maringá, Paraná.

Extraction and isolation

The air-dried and powdered aerial parts (1.5 Kg) were extracted with MeOH (51) and the solvent was evapd under red. pres. The methanolic extract (79.9 g) was dissolved in 0.5 M HCl (400 ml), the soln extracted with hexane-Et₂O 1:1 (3 × 150 ml) and the aq. layer basified with K₂CO₃ powder to pH 10. The basic soln was stirred for 5 h and the resulting ppt. was filtered and dried to give 5.53 g of alkaloid material. This material was sepd into 5 frs over alumina CC with nhexane-CHCl₃ 2:1 (200 ml), hexane-CHCl₃ 1:1 (300 ml), CHCl₃ (150 ml), CHCl₃-MeOH 1:1 (250 ml) and MeOH (200 ml). Recrystallization of the residue from the *n*-hexane-CHCl₃ 1:1 and CHCl₃ eluates with Me₂CO, afforded 750 mg of 1. The residue of the CHCl₃-MeOH 1:1 eluate (1.19 g) was subjected to silica gel CC using mixts of CHCl3-MeOH 9.5:0.5, 9:1, 8:2, 7:3, 6:4 and 1:1 as eluent; 25 frs were collected. Frs 12–14 (11.2 mg) were recrystrallized from Me₂CO to yield 10.0 mg of **2**. The residue of frs 17–22 (350.1 mg) was recrystallized from Me₂CO–MeOH to give 207.8 mg of **3**. The residue (870.1 mg) of the MeOH eluate from alumina CC was subjected to silica gel CC with mixts of CHCl₃–MeOH, 9:1, 8:2, 6:4 and 1:1; 15 frs were collected. Frs 7–10 were recrystallized from Me₂CO to yield 310.0 mg of **3**. Recrystallization of the residue from frs 13–15 with MeOH yielded **4** (270.0 mg).

Acid hydrolysis of the glycoalkaloids

The glycoalkaloid (5 mg) was refluxed with 5 ml of 5% HCl-MeOH soln for 2 hr. The soln was diluted with H₂O and extracted with CHCl₃ and the aglycone was sepd. The aq. filtrate was neutralized with BaCO₃ and evapd. The resulting residue was chromatographed on TLC (silica gel; CHCl₃-MeOH-Me₂CO-H₂O, 3:3:3:1) against ref. sugars. Compounds 2, 3 and 4 yielded leptinidine (1) as aglycone, which was identified by comparison of the EIMS and ¹H NMR spectroscopic data with those of the authentic sample. The sugar components were identified as glucose for compound 2; glucose and rhamnose for compound 3; glucose, galactose and rhamnose for compound 4.

Leptinidine (1)

Crystals from Me₂CO, mp 248–249°, ref [7]: 245–247°. [α]_D²² – 29° (CHCl₃, c 0.30), ref [7]: -24° (CHCl₃). EIMS (probe) 70 eV m/z (rel. int.): 413 [M]⁺ (9), 369 (4), 342 (6), 220 (34), 166 (100). ¹H NMR (300 MHz, CDCl₃): Table 1. ¹³C NMR (75.5 MHz, CDCl₃): Table 2.

Leptinidine $3-O-\beta-D-glucopyranoside$ (2)

Amorphous powder. IR $v_{\text{max}}^{\text{KB}}$ cm⁻¹: 3400 (OH), 1665 (C=C), 1055 (C-O). EIMS (probe) 70 eV m/z (rel. int.): 413 (4), 220 (25), 166 (100). FABMS m/z (rel.

int.): 576 $[C_{33}H_{53}NO_7 + H]^+$ (100), 414 $[C_{27}H_{43}NO_2 + H]^+$ (25), 412 (30). ¹H NMR (300 MHz, pyridine– d_5): Table 1. ¹³C NMR (75.5 MHz, pyridine– d_5): Table 2.

Leptinidine 3–O-[α -L-rhamnopyranosyl-($1 \rightarrow 4$)-[α -L-rhamnopyranosyl-($1 \rightarrow 2$)]- β -D-glucopyranoside], leptinine I (3)

Amorphous powder. [α]₂² – 85° (pyridine, c 0.30). IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3395 (OH), 1665 (C=C), 1050 (C=O). FABMS m/z (rel. int.): 868 [C₄₅H₇₃NO₁₅+H]⁺ (100), 722 (30), 576 (5), 414 [C₂₇H₄₃NO₂+H]⁺ (28). EIMS (probe) 70 eV m/z (rel. int.): 413 (3), 220(27), 166 (100). ¹H NMR (300 MHz, pyridine– d_5): Table 1. ¹³C NMR (75.5 MHz, pyridine– d_5): Table 2.

Leptinidine 3–O-[β -D-glucopyranosyl- $(1 \rightarrow 3)$ -[α -L-rhamnopyranosyl- $(1 \rightarrow 2)$ - β -D-galactopyranoside], leptinine II (4).

Amorphous powder. $[\alpha]_{D}^{22}-60^{\circ}$ (pyridine, c 0.30). IR $\nu_{\text{max}}^{\text{KBr}}\text{cm}^{-1}$: 3400 (OH), 1667 (C=C), 1055 (C=O). FABMS m/z (rel. int.): 884 $[C_{45}H_{73}NO_{16}+H]^+$, 722 (20), 414 $[C_{27}H_{43}NO_2+H]^+$. EIMS (probe) 70 eV m/z (rel. int.): 413 (6), 220 (27), 166 (100). ¹H NMR (400 MHz, pyridine- d_5): Table 1. ¹³C NMR (75.5 MHz, pyridine- d_5): Table 2.

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