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PHENYLALKANOIDS FROM PIPER MARGINATUM:

BÁRBARA VIVIANA DE O. SANTOS, EMIDIO V. L. DA-CUNHA MARIA CÉLIA DE O. CHAVES* and Alexander I. Gray†

Laboratório de Tecnologia Farmacêutica, Universidade Federal da Paraíba, Cx. Postal 5009, 58051-970, João Pessoa, PB, Brazil; † Phytochemistry Research Laboratories, Department of Pharmaceutical Sciences, University of Strathclyde, 204 George Street, Glasgow G1 1XW, UK.

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Abstract—Phenylpropanoids, and a new phenyloctanoid were isolated from the roots of *Piper marginatum* and their structures were elucidated by spectroscopic methods. The compounds were: 3,4-methylenedioxy-1-(2E-octenyl)-benzene, 2,6-dimethoxy-3,4-methylenedioxy-1-(2-propenyl)-benzene, 1-(1E-propenyl)-2,4,6-trimethoxybenzene; apiole; isoasarone. © 1998 Elsevier Science Ltd. All rights reserved

INTRODUCTION

Many plants of the genus Piper are used as food flavoring agents and for the treatment for numerous diseases [1-3]. P. marginatum has previously only been shown to produce the phenylpropanoid, croweacin (2methoxy-3,4-methylenedioxy-1-(2-propenyl)-benzene [4]. Herein we describe the isolation and structure elucidation of 3,4-methylenedioxy-1-(2E-octenyl)benzene(1) and 1-(1E-propenyl)-2,4,6-trimethoxybenzene (2) from the dried ground root of P. marginatum. The plant popularly known as "mavaisco", is used in the Brazilian state of Paraíba as a food flavouring agent (seeds) and as an antidote for snakebite (root) [5]. We named the new compounds marginatine (1) and pipermargine (2), respectively. Complete ¹H and ¹³C NMR assignments for apiole (4) and isoasarone (5) are given here for the first time [6].

RESULTS AND DISCUSSION

The ¹H NMR spectrum of compound **1** showed the presence of an ABX system, with signals centred at δ 6.69 (1H, brd, J = 1.6 Hz), 6.64 (1H, brdd, J = 7.9, 1.6 Hz) and 6.74 (1H, d, J = 7.9 Hz), of an aromatic ring and a singlet at δ 5.92 (2H) for a methylenedioxy group indicating a piperonyl moiety; a series of signals indicating a long chain with integrations totalling 15 H. Analysis of the ¹³C NMR spectrum confirmed the

presence of a methylenedioxy group (δ 101.0) and a piperonyl substituent. It also showed evidence of a C_8 side-chain with one double bond (δ 14.3, 22.7, 29.4, 31.7, 32.7, 39.0, 129.0 and 132.3). EI mass spectrometry gave a $[M]^+$ at m/z at 232 which is com-

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^{*} Author to whom correspondence should be addressed.

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patible with the molecular formula $C_{15}H_{20}O_2$. The NMR HMBC spectrum gave evidence for one methylene group between the double bond of the side chain and the piperonyl moiety. The large J value obtained for the olefinic protons, H-2′ and H-3′, pointed to an E configuration for the double bond in the 2-octenyl side-chain.

The ¹H NMR spectrum of **2** indicated the symmetrical nature of the molecule. It showed one s at δ

6.15 (2H) for H-3 and H-5; one s at δ 3.83 (6H) attributable to two methoxy groups (2-C-OMe and 6-C-OMe); a further methoxy at δ 3.82 (3H) was assigned to the 4-C-OMe; one allylic methyl dd centred at δ 1.90 (C-3') was coupled to two trans olefinic protons with signals centred at δ 6.57 (H-1') and δ 6.47 (H-2'). ¹³C NMR chemical shifts of the three methoxy groups (δ 55.9 for 2-C-Ome, 6-C-Ome and 55.5 for 4-C-Ome) indicated that all these moieties should have at least one of the ortho carbons unsubstituted. This fact, together with protons H-3 and H-5 being equivalent, confirms the substitution pattern for the aromatic ring proposed for compound 2. EI mass spectrometry gave the $[M]^+$ at m/z 206 which is consistent with the molecular formula C₁₂H₁₄O₃. All the NMR assignments presented for all the compounds were based on 2D experiments such as HMBC (with J optimised for 7 Hz), ¹H-¹H COSY, NOESY and H-C COBI. Complete ¹H and ¹³C NMR data for compounds 1 and 2 are given in Tables 1 and 2, respectively. The complete spectroscopy analysis of 3 is in accordance with the literature [7] while complete and unambiguous ¹H and ¹³C NMR data for the compounds 4 and 5 are presented here for the first time (Table 3).

EXPERIMENTAL

General methods

CC: silica gel; Prep. and analyt. TLC: silica gel 60 PF $_{254}$. Compounds were detected using UV light and/or spraying with H_2SO_4 -vanillin reagent. IR: liquid film; EIMS: GC-MS at 70 eV (Column HP-1, Init. temp. 70°, rate 20°, final temp. 220°); NMR: 400.14 MHz for 1H and 100.62 MHz for ^{13}C . Chemical shifts are reported in ppm relative to the solvent (CDCl $_3$) at 27°.

Table 1. 13C and 1H NMR (CDCl₃) data for marginatine (1) with correlations based on H-C COBIdec and HMBC

	Н	$^1\!J$	2J	3J
[_	135.3	_	_
!	6.69 (1 H, brd, J = 1.6 Hz)	109.2	_	121.3, 145.9
3	_	147.8	_	_
	_	145.9	_	_
	6.74(1H, d, J = 7.9 Hz)	108.3	145.9	135.3, 147.8
)	6.64(1H, brdd, J = 1.6, 7.9 Hz)	121.3	_	39.0, 109.2, 145.9
′	3.25 (2H, brd, J = 4.8 Hz)	39.0	129.0, 135.3	109.2, 121.3, 132.3
<i>'</i>	5.52 (1H, dm, J = 15.4 Hz)	129.0	_	32.7
′	5.50 (1H, dm, J = 15.4 Hz)	132.3	_	39.0
,	2.02(2H, m)	32.7	132.3	129.0
<i>i'</i>	1.38(2H, m)	29.4	31.7	_
5′	1.30(2H, m)	31.7	_	_
"	1.30(2H, m)	22.7	_	_
′	0.90(3H, m)	14.3	22.7	31.7
OCH ₂ O	5.92 (2H s)	101.0		145.9, 147.8

Table 2. ¹³C and ¹H NMR (CDCl₃) data for pipermargine (2) with correlations based on H-C COBIdec and HMBC

	Н	1J	2J	3J
1	_	108.7	_	_
2	_	159.0	_	_
3/5	6.15 (2H, s)	91.0	159.0, 159.6	91.0, 108.7
4	_ ` ` ′	159.6		
6	_	159.0	_	_
1'	6.57 (1H, dq, J = 1.3, 16.0 Hz)	121.2	121.2	20.2, 159.0
2'	6.47 (1 H, dq, J = 6.0, 16.0 Hz)	128.2	20.2, 128.2	108.7
3'	1.90 (3H, dd, J = 1.3, 6.0 Hz)	20.2	121.2	128.2
C2/6-OMe	3.38 (6H, s)	55.9	_	159.0
C4–OMe	3.82(3H, s)	55.5	_	159.6

Table 3. ¹³C and ¹H NMR (CDCl₃) data for apiole (4) and isoasarone (5)

	Apiole		Isoasarone		
	Н	С	Н	С	
1	_	110.8	_	120.3	
2	_	136.5	6.70(1H, s)	114.3	
3	_	139.0		143.3	
4	_	135.2	_	148.2	
5	_	139.2	6.54(1H, s)	98.4	
6	6.31 (1H, s)	108.5	_ ` ` ` ` `	151.6	
1'	3.31 (1H, dm, J = 6.4 Hz)	34.3	3.30 (2H, dm, J = 6.4 Hz)	33.8	
2′	5.94 (1H, m)	137.6	5.97 (1H, m)	137.5	
3′	5.05(2H, m)	115.6	5.04(2H, m)	115.4	
C2-OMe	3.88(3H,s)	60.4	_ ` ` ` ` `	_	
C3-OMe	_ ` ´ ´	_	3.83(3H, s)	56.8	
C4-OMe	_	_	3.88(3H, s)	56.5	
C5-OMe	3.86(3H, s)	57.1	_ ` ´ ′	_	
C6-OMe	_	_	3.81(3H, s)	56.8	
OCH ₂ O	5.96 (2H, s)	101.7		_	

Plant material

The root of *P. marginatum* Jacq. was collected in September 1993 near the city of João Pessoa, PB, Brazil. A voucher specimen (Agra 1500-JPB) is deposited at the Herbarium Lauro Pires Xavier of the Universidade Federal da Paraíba.

Extraction and isolation

The powdered root of *P. marginatum* (4.2 kg) was moistened with a soln. of 40% conc. NH₄OH in H₂O and extracted with EtOH at room temp. The resultant extract (170 g) was treated with a soln. of 2% HCl in H₂O and extracted with CHCl₃. This extract (22 g) was then subjected to cc (column 1) over silica gel, elution with hexane, hexane–CHCl₃ mixtures, CHCl₃, CHCl₃–MeOH mixtures and finally MeOH (100 fractions). The fractions 10–15 (1.0 g) eluted with hexane–CHCl₃ (1:4) were subjected to a second column (col-

umn 2) over silica gel using hexane and CHCl₃, (15 fractions). Fractions 2–3 (70 mg) eluted with pure hexane, were subjected to prep. TLC developed with pure hexane. Fraction 1 (48 mg) was marginatine (1). The fractions 18-19 of column 1 (80 mg) when subjected to prep. TLC developed with hexane-CHCl₃ (9:1) gave pipermargine (2) (20 mg). A column of the fractions 23-26 from column 1 (1.02 g) (column 3) eluted with hexane and CHCl3 mixtures of increasing polarity gave 14 fractions. Fractions 3-4 (27 mg) contained 2,6-dimethoxy-3,4-methylenedioxy-1-(2-propenyl)-benzene (3), which was previously isolated from P. sarmentosum [7]. The fraction 8 of column 3 when purified by prep. TLC developed with hexane-EtOAc (9:1) gave apiole (26 mg) (4). Fractions 11–14 of column 3 purified in the same way gave isoasarone (6.6 mg) (5).

Marginatine (1). Pale yellow oil. IR ν_{max} cm⁻¹: 2933, 2854, 2360, 1488, 1442, 1241, 1041; UV λ_{max} nm: 288, 233; EIMS m/z (rel. int.): 232 [M⁺] (75), 161 (68), 131 (100); NMR: Table 1.

Pipermargine (2). Pale yellow oil. IR ν_{max} cm⁻¹: 2923, 2846, 1604, 1457, 1203, 1126; UV λ_{max} nm: 268 nm; IEMS m/z (rel. int.): 208 [M⁺] (100), 180 (51); NMR: Table 2.

2,6-Dimethoxy-3,4-methylenedioxy-1-(2-propenyl)-benzene (3). Pale yellow oil. Spectroscopy data identical to literature [7].

Apiole (4). Pale yellow oil. IR v_{max} cm⁻¹: 2923, 2846, 1511, 1457, 1203, 1033; UV λ_{max} nm: 280 nm; EIMS m/z (rel. int.): 222 [M⁺] (100), 207 (47), 195 (20), 191 (17), 178 (33); NMR: see Table 3.

Isoasarone (5). Pale yellow oil. IR v_{max} cm⁻¹: 2923, 2846, 1511, 1458, 1203, 1033; UV λ_{max} nm): 291, 350; EIMS m/z (rel. int.): 208 [M⁺] (100), 193 (52), 177 (16), 164 (19); NMR: see Table 3.

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