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TRITERPENOIDS FROM OWENIA CEPIODORA

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Abstract—The limonoid, 28-deoxonimbolide, and three protolimonoids, 24*S*,25-dihydroxytirucall-7-en-3-one, 3-oxo-tirucalla-7, 24-dien-21-al and 21,24*R*-epoxy-25-hydroxytirucall-7-en-3-one were isolated from the leaves and bark of *Owenia cepiodora*. © 1998 Published by Elsevier Science Ltd. All rights reserved

INTRODUCTION

The Meliaceae family produces a class of compounds called limonoids. Limonoids are tetranortriterpenoids with a β - substituted furan ring at C-17 α . Rings A-D of the triterpenoid skeleton may be oxidized and rearrangements may occur giving different types of limonoids. The type of limonoid produced by a species has been used for taxonomic purposes [1]. Owenia is a genus of the Meliaceae found only in Australia and has been placed in the tribe Trichileae of the subfamily Melioideae. Seed of Owenia acidula and Owenia venosa have been examined previously and have yielded the limonoid 6α-acetoxyhavanensin acetate and the triterpenoid 3α-isobutyryl-7α-deacetylglabretal [2]. Glabretaltype compounds are an interesting by-product of limonoid metabolism and have been isolated from Aglaia [3], Guarea [4], Owenia [2], Turraea [3], and Dysoxylum [5-7], species. In the classification of Pennington and Styles [8], these genera belong to different tribes of the subfamily Melioideae. However, in spite of the thorough investigation of Melia and Azadirachta in recent years, glabretaltype compounds have not been found in the Meliaea tribe of the Melioideae subfamily. The limonoid 6αacetoxydeoxyhavanensin is a common limonoid and hence its presence in the Owenia examined previously is not of taxonomic significance.

The leaves and bark of *Owenia cepiodora* F. Muell, were investigated in order to determine their chemical composition.

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RESULTS AND DISCUSSION

The ring C opened limonoid, 28-deoxonimbolide (1) was isolated from the leaves of Owenia cepiodora. The compound has been isolated previously from the leaves of Azadirachta indica (neem) [9]. Melia and Azadirachta are very similar chemically and contain ring C-oxidized limonoids of the nimbin class. These compounds were thought to distinguish Melia and Azadirachta from the rest of the Meliaceae family [1], although one other example of a ring C opened limonoid had been found previously in the Trichilieae tribe of which Owenia is also a member: the isolation of heudebolin from Trichilia heudelotti [10]. This second report confirms that ring-C opened limonoids are not restricted to Melia and Azadirachta.

Three triterpenoids were isolated from the hexane extract of the bark of O. cepiodora. The first compound was identified as 24S,25-dihydroxytirucall-7en-3-one (2). This compound has not been reported previously, although the 3β -hydroxy derivative (2C) has been described [11]. C-3 occurred at δ 216.1 confirming the presence of a ring A ketone, H-7 occurred at δ 5.28 and C-7 and C-8 at δ 117.8 and δ 145.9, respectively. Resonances occurred at δ 73.2 and δ 78.6 in the ¹³C NMR spectrum and these were assigned to C-25 and C-24, respectively. H-24 occurred at δ 3.32. Standard acetylation of 2 yielded the 24-acetate, (2A); forced acetylation of 2 led to the 24,25-diacetate (2B), and NaBH₄ reduction yielded the known compound $3\beta,24S,25$ trihydroxytirucall-7-ene (2C). The stereochemistry at C-24 of the 3/3-hydroxy derivative 2C was determined by the lanthanide complex method of

Nakanishi *et al.* [12–14]. Complexation [4:1 ratio, substrate: Eu(dpm)₃] of **2**C in dry chloroform provided a CD spectrum $\Delta\epsilon$ + 4.5 at 314 nm indicating the C–24 hydroxy group to have the *S*-configuration [11].

The second compound was identified as 3-oxo-tirucalla-7,24-dien-21-al (3) and has been isolated previously from *Simarouba amera* [15]. Reduction of the two carbonyl groups with NaBH₄ yielded the 3β ,21-diol (3A) as expected.

The third compound was identified as 21.24R-epoxy-25-hydroxytirucall-7-en-3-one (4) and has not been reported previously although the 23α -hydroxy analogue, bourjotinolone A(5) has been isolated from *Flindersia bourjotiana* [16]. Mass spectrometry indicated a molecular formula of $C_{30}H_{48}O_3$. ^{13}C NMR spectra were similar to (2), with a keto group at C-3 (δ 216.9) and Δ^7 -double bond (δ 117.9, 145.8). C-24 had been shifted from δ 78.6 in (2) to δ 84.3 and C-25 from δ 73.2 to δ 72.1. The C-21 methyl carbon resonance had been replaced by a methylene carbon resonance at δ 70.6. The H_3 -21 doublet which occurred at δ 0.85 in (2) was replaced by a pair of resonances at δ 4.02 (d,

J = 11.4 Hz) and $\delta 3.42$ (dd, J = 11.4, 2.4 Hz). The molecular formula indicated a ring was necessary in the side chain. Thus structure (4) is proposed. Acetylation was unsuccessful confirming the absence of a primary hydroxy group at C-21. The ¹H NMR spectrum for bourjotinolone A (5) [16] showed H-24 as a doublet at δ 2.9 $(J_{23\beta,24\alpha}=9.0 \text{ Hz})$ and the ¹H NMR spectrum for sapelin C (6) [17] showed H-24 as a doublet at δ 2.9 $(J_{23\beta,24\alpha} = 10.0 \text{ Hz})$. In (4), H-24 appears as a doublet of doublets signal $(J_{23\beta,24\alpha} = 10.5 \text{ Hz},$ $J_{23\beta,24\alpha} = 2.4$ Hz) at δ 3.10. These coupling constants suggest the configuration at C-24 of (4) to be R, as in bourjotinolone A (5) and sapelin C(6). The ¹³C NMR data for bourjotinolone A (5) [18], is given in Table 1 for comparison purposes.

EXPERIMENTAL

Dried leaves (250 g) and bark (121 g) of *Owenia cepiodora* F. Muell. (Voucher Specimen UND.HAM 8) were collected in Rockhampton, Australia. The leaves were crushed and extracted with hexane and CHCl₃. The CHCl₃ extract yielded,

Table 1. ¹³C NMR data for compounds (2), (3), (4) and (5) (75 MHz, CDCl₃)

Carbon	(2)	(3)	(4)	(5)
l	38.5 t	38.3 t	38.5 t	38.6 t
1 2 3 4 5	34.9 <i>t</i>	34.8 t	34.9 1	35.0 t
3	216.0 s	216.7 s	216.9 s	216.8 s
4	47.8 s	47.8 s	47.8 s	47.9 s
5	53.1 d	55.2 d	52.3 d	52.4 a
6	24.3 t	24.3 t	24.3 t	24.4 /
7	117.8 d	118.3 d	17.9 d	118.1 a
8	145.9 s	145.2 s	45.8 s	145.7 s
9	52.3 d	52.2 d	48.4 d	48.5 d
10	34.9 s	35.0 s	35.0 s	35.1 s
11	18.2 t	17.7 t	18.2 t	18.2 t
12	32.9 t	31.9 t	32.9 t	33.0 t
13	43.4 s	43.3 s	43.1 s	43.3 s
14	51.1 s	50.9 s	51.3 s	51.3 s
15	$34.0 \ t$	33.6 t	33.9 t	34.0 t
16	28.2* t	29.2 t	27.2 t	27.4 t
17	48.4 d	48.2 d	43.2 d	44.8 a
18	21.5 q	21.5 q	21.6 q	21.6 q
19	$12.7 \frac{1}{q}$	$12.7 \dot{q}$	12.7 q	12.8 q
20	35.9 d	48.2 d	35.6 d	37.5 d
21	$18.2 \ q$	205.8 d	70.6 t	70.1 t
22	28.4* 1	26.9* t	27.0 t	36.5 t
23	33.6 t	25.9* 1	21.2 t	64.6 d
24	78.6 d	123.5 d	84.3 d	86.5 d
25	73.2 s	132.5 s	72.1 s	74.2 s
26	24.5 q	17.8 q	24.5 q	24.0 g
27	26.5 q	24.5 q	25.8 q	28.5 q
28	23.1 q	25.6 q	23.5 q	24.6 q
29	$21.9 \frac{1}{q}$	22.8 q	22.4 q	22.3 q
30	27.4 q	27.2 q	$27.5 \frac{q}{q}$	27.5 q

[&]quot;Values for the same compound may be interchanged.

after repeated CC over silica gel (Merck 9385), the limonoid 28-deoxonimbolide (1). The hexane extract of the ground bark afforded three triterpenoids, 24S,25-dihydroxytirucall-7-en-3-one (2), 3-oxotirucalla-7,24-dien-21-al (3) and 21,24*R*-epoxy-25-hydroxytirucall-7-en-3-one (4). The structures of the compounds were determined using 2D NMR and MS techniques. ¹H NMR spectra were recorded using a Varian Gemini 300 NMR spectrometer in CDCl₃. ¹³C NMR data for (2), (3), and (4) are given in Table 1.

28-Deoxonimbolide (1) $C_{27}H_{32}O_6$ (found 452.2178, requires 452.2181). The structure was confirmed by comparison of physical data against literature values [9].

24*S*,25-Dihydroxytirucall-7-en-3-one (**2**) (616 mg), HRMS m/z 458.3746 (C₃₀H₅₀O₃ requires 458.3760). ¹H NMR (300 MHz, CDCl₃): δ 0.78 (3H, s), 0.85 (3H, d, J = 6.0 Hz, H₃–21), 0.97 (6H, s, 2 × CH₃), 1.02 (3H, s), 1.08 (3H, s), 1.14 (3H, s), 1.19 (3H, s), 2.73 (1H, td, J = 5.7, 15.4 Hz, H–2ax), 3.32 (1H, m, H–24), 5.28 (1H, m H–7). 1R $\lambda_{\rm max}^{\rm NaCl}$ cm⁻¹: 3450, 1700.

Acetylation of (2) with Ac₂O/pyridine yielded 24-acetoxy-tirucall-7-en-25-ol-3-one (2A). ¹H NMR: δ 0.78 (3H, s), 0.84 (3H, d, J = 6.0 Hz, H₃–21), 0.98 (6H, s), 1.02 (3H, s), 1.09 (3H, s), 1.17 (6H, s), 2.09 (3H, s OAc), 2.73 (1H, td, J = 5.4, 14.4 Hz, H–2ax), 4.77 (1H, dd, J = 2.7, 10.2 Hz, H–24), 5.28 (1H, m H–7).

Acetylation of **2** with Ac₂O/pyridine/dimethylaminopyridine yielded 24*S*,25-diacetoxy-tirucall-7-en-3-one (**2B**). ¹H NMR: δ 0.78 (3H, s), 0.84 (3H, d, J = 6.0 Hz, H₃-21), 0.98 (3H, s), 1.02 (3H, s), 1.09 (3H, s), 1.23 (3H, s), 1.42 (3H, s), 1.46 (3H, s), 1.94 (3H, s, OAc), 2.09 (3H, s, OAc), 2.73 (1H, td, J = 6.6, 14.4 Hz, H-2ax), 5.10 (1H, dd, J = 2.7, 9.9 Hz, H-24), 5.30 (1H, m, H-7).

Reduction of **2** with NaBH₄ yielded 3β ,24*S*,25-trihydroxy-tirucall-7-ene (**2C**). ¹H NMR. δ 0.72 (3H, *s*, H₃–19), 0.80 (3H, *s*, H₃–18), 0.84 (3H, *s*, H₃–30), 0.86 (3H, *d*, *J* = 6.0 Hz, H₃–21), 0.95 (6H, *s* H₃–29, H₃–28), 1.14 (3H, *s*, H₃–26), 1.20 (3H, *s*, H₃–27), 3.25 (1H, *dd*, *J* = 4.5, 10.8 Hz, H–24), 3.33 (1H, *m*, H–3 α , W_{1/2} = 12.0 Hz).

3-Oxo-tirucalla-7,24-dien-21-al (3) (678 mg), HRMS M⁺ at m/z 438.3487 ($C_{30H46}O_2$ requires 438.3498). ¹H NMR: δ 0.81 (3H, s), 0.97 (3H, s), 0.99 (3H, s), 1.02 (3H, s), 1.08 (3H, s), 1.54 (3H, s), 1.65 (3H, s), 2.71 (1H, td, J = 6.3, 14.4 Hz, H-2ax), 5.05 (1H, m H-24), 5.30 (1H, m, H-7), 9.47 (1H, d, H-21, J = 5.4 Hz). IR $\lambda_{\rm max}^{\rm Nacl}$ cm⁻¹: 1730, 1680.

Reduction of **3** with NaBH₄ yielded 3β ,21-dihydroxy-tirucalla-7,24-diene (**3A**), mp 136–138° (lit. 138–139° [12]). ¹H NMR: δ 0.72 (3H, s), 0.81 (3H, s), 0.84 (3H, s), 0.95 (3H, s), 0.97 (3H, s), 1.60 (3H, s), 1.69 (3H, s), 3.22 (1H, dd, H–3 α , J = 4.5, 10.8 Hz), 3.58 (1H, dd, J = 4.8, 11.1 Hz, H–21a), 3.71 (1H, dd, J = 3.0, 11.1 Hz, H–21b), 5.08 (1H, m, H–24), 5.25 (1H, m, H–7).

21,24*R*-Epoxy-25-hydroxytirucall-7-en-3-one (4) (1.0 g), HRMS M⁺ at m/z 456.3609 (C₃₀H₄₈O₃ requires 456.3603). ¹H NMR: δ 0.77 (3H, s), 0.98 (3H, s), 1.02 (3H, s), 1.03 (3H, s), 1.09 (3H, s), 1.13 (6H, s), 2.73 (1H, td, J = 6.6, 14.4 Hz, H–2ax), 3.10 (1H, dd, J = 2.4, 10.5 Hz, H–24), 3.43 (1H, dd, J = 2.4, 11.4 Hz, H–21a), 4.02 (1H, d, J = 11.4 Hz, H–21b), 5.29 (1H, m, H–7). IR $\lambda_{\text{max}}^{\text{NaCl}}$ cm⁻¹: 3420, 1700.

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