



EUSCAPHOLIDE AND ITS GLUCOSIDE FROM LEAVES OF *EUSCAPHIS JAPONICA*

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Key Word Index—*Euscaphis japonica*; Staphyleaceae; tetraketide; euscapholide; euscapholide glucoside.

Abstract—A new tetraketide, euscapholide and its glucoside were isolated from the leaves of *Euscaphis japonica*. The structures were elucidated by spectroscopic and chemical analyses. © 1998 Elsevier Science Ltd. All rights reserved

INTRODUCTION

Flavonol glycosides, anthocyanins and compounds positive to Ehrlich reagent have been isolated from the capsule of *Euscaphis japonica* (Thunb.) Kanitz [1,2]. In the course of our studies on the constituents of subtropical plants, we examined the constituents of the leaves of the title plant harvested in Okinawa Prefecture, Japan and isolated a tetraketide, euscapholide (**1**), and its glucoside (**3**). This paper describes the isolation and structural elucidation of the new compounds.

RESULTS AND DISCUSSION

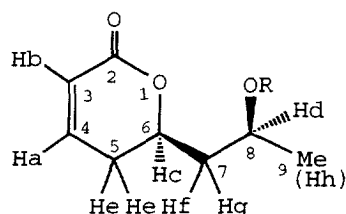
Euscapholide (**1**) and its glucoside (**3**) were isolated from the *n*-BuOH-soluble fraction of the methanolic extract obtained from the leaves of *E. japonica*.

Euscapholide (**1**) was found to have the molecular formula $C_{14}H_{22}O_8$ by HR-EI-mass spectrometry. The IR spectrum showed the presence of a hydroxyl group (3420 cm^{-1}) and a δ -lactone (1700 cm^{-1}). The ^1H NMR spectrum showed the presence of a secondary methyl group [δ 1.26 (H_b)] on one of the carbon atoms (C-8) having an oxygen atom, two methylene groups [δ 1.77 (H_g), 2.03 (H_f) and 2.42

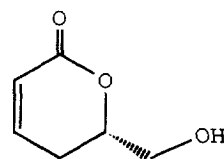
(H_{e2})], a methine on C-8 [δ 4.08 (H_d)], a methine on the carbon having an acyloxy group [δ 4.65 (H_c)] and a conjugated disubstituted double bond with *Z*-geometry [δ 6.02 (H_b) and 6.92 (H_a)]. The ^{13}C NMR spectrum (Table 1) contained the signals for above mentioned functional groups. Acetylation of **1** with acetic anhydride and pyridine gave the monoacetate **2** [δ 2.05 (3H, *s*) and 5.13 (1H, *m*), H_d]. The planar structure was deduced as **1** by analysis of the ^1H - ^1H COSY spectrum. Namely, the cross peaks were followed as $H_b \rightarrow H_a \rightarrow H_e \rightarrow H_c \rightarrow H_f, H_g \rightarrow H_d \rightarrow H_h$. The absolute stereochemistry at C-6 was elucidated as *R* from the fact that **1** showed the opposite Cotton effect [$\Delta\epsilon_{255} + 3.0$] to that of (*S*)-5,6-dihydro-6-hydroxymethyl-2H-pyran-2-one(**7**) [3]. Another asymmetric centre at C-8 was determined to have *S*-configuration by a modified Mosher's method (Fig. 1) [4]. Thus, the structure of euscapholide was elucidated as **1**.

Euscapholide glucoside (**3**) was assigned the molecular formula $C_{14}H_{22}O_8$ by negative ion HR-FAB-mass spectrometry. The ^1H and ^{13}C NMR spectra were very similar to those of euscapholide (**1**), except for the appearance of the signals due to β -glucopyranosyl moiety (Table 1 and Experimental). Thus, compound **3** was presumed to be β -glucopyranoside of **1**. Compound **3** gave the tetraacetate **4** [δ_H 2.00, 2.02, 2.03 and 2.09 (each 3H, *s*)] on acetylation with acetic anhydride and pyridine and euscapholide (**1**), $[\alpha]_D + 116.4^\circ$ (MeOH), upon enzy-

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- 1 R=H
 2 R=COMe
 3 R=Glc
 4 R=Glc (COMe)₄
 5 R=(R)-MTPA
 6 R=(S)-MTPA



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matic hydrolysis. Thus, the structure of euscapholide glucoside was elucidated as 3.

EXPERIMENTAL

General

NMR: ¹H (400 MHz) and ¹³C (100 MHz) with TMS as int. standard; FABMS: PEG-400 as matrix; CC: Dianion HP-20 (Mitsubishi Kasei) and silica gel 60 (230–400 mesh, Merck); TLC: silica gel 60 F₂₅₄ (0.25 mm and 0.5 mm in thickness); prep. HPLC: Cosmosil 10 C₁₈, 20 × 250 mm; solvent: MeOH–H₂O 1:4, 6 ml min⁻¹, detection 210 nm.

Plant material

The plant material was collected at Kunigami-son, Okinawa Prefecture, Japan in July 1994 and identified as *Euscaphis japonica* (Thunb.) Kanitz by one of the authors (A. T.). A voucher specimen (9407-4) was deposited in the Herbarium of Institute of Pharmaceutical Sciences, Hiroshima University School of Medicine.

Table 1. ¹³C NMR data for euscapholide (1) and its glucoside (3). (CD₃OD, 100 MHz)

C	1	3
2	166.9 (164.2)*	167.1
3	121.3 (121.2)	121.3
4	148.3 (145.4)	148.3
5	30.2 (29.3)	30.4
6	77.5 (76.8)	77.1
7	44.7 (43.5)	42.6
8	64.8 (65.0)	74.6
9	23.6 (23.6)	22.2
1'		104.2
2'		75.2
3'		78.2
4'		71.6
5'		77.9
6'		62.8

* Chemical shifts in parentheses are for chloroform-d.

Isolation

Dried leaves (2.9 kg) of *E. japonica* were extracted (×2) with MeOH (45 l) at room temp for three weeks. The combined methanolic extract was concentrated *in vacuo*. The residue was dissolved in 90% aq. MeOH (2.1 l) and the solution washed with *n*-hexane (1 l × 3). The aq. MeOH layer was concentrated *in vacuo* and the residue was suspended in H₂O (1 l). The suspension was extracted with EtOAc (1 l × 3) and *n*-BuOH (1 l × 3), successively. The *n*-BuOH extract was concentrated *in vacuo* to give a residue (77 g) which was chromatographed on Dianion HP-20 (a highly-porous synthetic resin [Φ = 78 mm, L = 536 mm] with stepwise increases of MeOH content in H₂O [0 (4 l), 10 (5 l), 30 (6 l), 40 (6.5 l), 50 (6.5 l), 70 (6.5 l) and 100 (6 l)]. Frs. of 500 ml being collected.

Frs 14–17 were combined and concentrated *in vacuo* to give a residue (10.7 g) which was chromatographed over silica gel (CHCl₃–MeOH with increasing amounts of MeOH). The fractions which contained a spot with *R_f* 0.35 on TLC (CHCl₃–MeOH–H₂O 15:6:1) were collected and the residue (418.7 mg) was purified by prep. HPLC to give euscapholide glucoside (3) (128.4 mg).

Frs 18–23 were combined and concentrated *in vacuo* to give a residue (12.2 g), which was repeatedly chromatographed over silica gel (CHCl₃–MeOH, Et₂O and finally CHCl₃–MeOH) to give euscapholide (1) (1.21 g). *R_f* 0.27 (Et₂O) and 0.29 (CHCl₃–Me₂CO 4:1).

Euscapholide (1)

Colourless oil, [α]_D²⁰ +115.5° (MeOH; *c* 1.52). IR *v*_{max} (CHCl₃) cm⁻¹: 3420, 1700, 1230, 1200, 1050, 1025; UV *λ*_{max} (MeOH) nm (log *ε*): 209 (3.89); ¹H NMR (CDCl₃): δ 1.26 (3H, *d*, *J* = 6.3 Hz, 9-H₃), 1.77 (1H, *ddd*, *J* = 14.6, 5.4 and 4.4 Hz, 7-H₁), 2.03 (1H, *dt*, *J* = 14.6 and 7.8 Hz, 7-H₁), 2.42 (2H, *m*, 5-H₂), 4.08 (1H, *m*, 8-H), 4.65 (1H, *m*, 6-H), 6.02 (1H, *dt*, *J* = 10.1 and 1.7 Hz, 3-H) and 6.92 (1H, *ddd*, *J* = 10.1, 5.4 and 3.4 Hz, 4-H); ¹³C NMR:

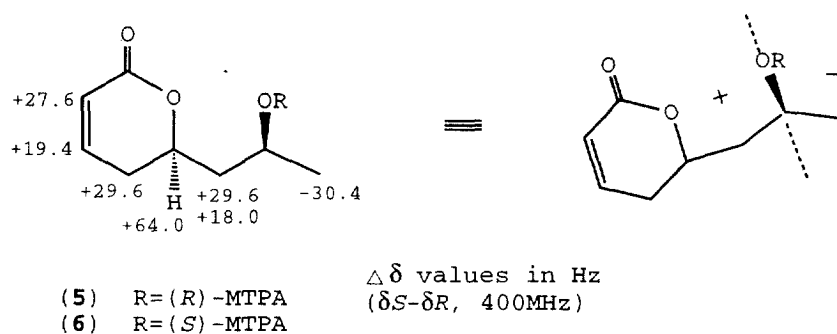


Fig. 1.

Table 1; CD (MeOH, c 1.41×10^{-4} M): $\Delta\epsilon$ (nm) + 3.0 (255); HR-EIMS m/z 156.0789 $[\text{M}]^+$ ($\text{C}_8\text{H}_{12}\text{O}_3$ requires 156.0786).

Euscapholide glucoside (3)

Amorphous powder, $[\alpha]_D + 36.4^\circ$ (MeOH, c 2.47). UV v_{\max} (MeOH) nm (log c): 209 (3.98); IR v_{\max} (dry film) cm^{-1} : 3387, 1703; ^1H NMR (CD_3OD): δ 1.33 (3H, d , $J = 5.9$ Hz, 9- H_3), 1.81 (1H, dt , $J = 14.2$ and 5.9 Hz, 7- H_1), 2.12 (1H, dt , $J = 14.2$ and 7.1 Hz, 7- H_1), 2.39 (1H, ddd , $J = 18.6$, 11.2 and 2.7 Hz, 5- H_1), 2.57 (1H, $br.d$, $J = 18.6$ and 4.9 Hz, 5- H_1), 3.14 (1H, dd , $J = 8.3$ and 7.8 Hz, H-2'), 3.66 (1H, dd , $J = 11.5$ and 4.9 Hz, 6'- H_1), 3.85 (1H, $br.d$, $J = 11.5$ Hz, 6'- H_1), 4.03 (1H, m , 8-H), 4.32 (1H, d , $J = 7.8$ Hz, 1'-H), 4.74 (1H, m , 6-H), 5.97 (1H, dd , $J = 9.8$ and 1.5 Hz, 3-H), 7.04 (1H, ddd , $J = 9.8$, 5.9 and 2.4 Hz, 4-H); ^{13}C NMR: Table 1; HR-FABMS (negative) m/z : 317.1229 $[\text{M}-\text{H}]^-$ ($\text{C}_{14}\text{H}_{21}\text{O}_8$ requires 317.1237).

Euscapholide monoacetate (2)

Euscapholide (1) (51.5 mg) was dissolved in a mixture of Ac_2O (0.5 ml) and pyridine (0.5 ml) and the mixture was left for 24 hr at room temp. Excess MeOH was added and the solution was concentrated *in vacuo*. The residue was purified by silica gel CC (CHCl_3 - Me_2CO) to give the monoacetate 3 (55.9 mg) as a colourless oil. IR v_{\max} (CHCl_3) cm^{-1} : 1710, 1630, 1370, 1220, 1040; ^1H NMR (CDCl_3): δ 1.31 (3H, d , $J = 6.4$ Hz, 9- H_3), 1.85 (1H, ddd , $J = 14.6$, 6.3 and 4.9 Hz, 7- H_1), 2.05 (3H, s , OAc), 2.19 (1H, ddd , $J = 14.6$, 8.1 and 6.6 Hz, 7- H_1), 2.33 (1H, ddd , $J = 18.8$, 11.7 and 2.4 Hz, 5- H_1), 2.47 (1H, dt , $J = 18.8$ and 4.7 Hz, 5- H_1), 4.53 (1H, m , 6-H), 5.13 (1H, m , 8-H), 6.02 (1H, dd , $J = 9.8$ and 1.5 Hz, 3-H), 6.90 (1H, ddd , $J = 9.8$, 5.9 and 2.4 Hz, 4-H); HR-EIMS m/z 198.0866 $[\text{M}]^-$ (Calculated for $\text{C}_{10}\text{H}_{14}\text{O}_4$: 198.0892).

(R)-MTPA ester (5) from euscapholide (1)

Euscapholide (1) (22.3 mg) was dissolved in CH_2Cl_2 (1 ml). (*R*)- α -methoxy- α -trifluoromethylphenyl acetic acid (MTPA) (146.7 mg), dimethylaminopyridine (42.3 mg) and dicyclohexylcarbodi-

imide (DCC) (126.0 mg) were added successively to the solution and the mixture was stirred for 5 min under ice cooling. The mixture was diluted with EtOAc (15 ml) and the solution was washed with 5% HCl, satd NaHCO_3 and satd NaCl successively. The dried EtOAc extract was concentrated *in vacuo* to give a residue which was purified by prep. TLC (CHCl_3 - Me_2CO 9:1) to give the (*R*)-MTPA ester 5 (40.3 mg). ^1H NMR (CDCl_3): δ 1.44 (3H, d , $J = 6.4$ Hz, 9- H_3), 1.88 (1H, ddd , $J = 14.2$, 6.4 and 4.9 Hz, 7- H_1), 2.19 (3H, m , 4- H_2 , 7- H_1), 3.56 (3H, d , $J = 1.0$ Hz, OMe), 4.30 (1H, m , 6-H), 5.34 (1H, m , 8-H), 5.97 (1H, $br.d$, $J = 9.8$ Hz, 3-H), 6.77 (1H, m , 4-H), 7.38 (3H, m), 7.52 (2H, m); HR-FABMS (negative): m/z 371.1087 (Calculated for $\text{C}_{18}\text{H}_{18}\text{O}_5\text{F}_3$: 371.1107).

(S)-MTPA ester (6) from euscapholide (1)

Euscapholide (1) (21.8 mg) was esterified with (*S*)-MTPA as described above to give the (*S*)-MTPA ester 6 (37.8 mg). ^1H NMR (CDCl_3): δ 1.36 (3H, d , $J = 6.3$ Hz, 9- H_3), 1.93 (1H, dt , $J = 14.7$ and 5.9 Hz, 7- H_1), 2.26 (1H, dt , $J = 14.7$ and 7.1 Hz, 7- H_1), 2.32 (2H, m , 5- H_2), 3.51 (3H, d , $J = 1.0$ Hz, OMe), 4.46 (1H, m , 6-H), 5.36 (1H, m , 8-H), 6.02 (1H, $br.d$, $J = 9.8$ Hz, 3-H), 6.84 (1H, m , 4-H), 7.41 (3H, m), 7.51 (2H, m); HR-FABMS (negative): m/z 371.1136 $[\text{M}-\text{H}]^-$ (Calculated for $\text{C}_{18}\text{H}_{18}\text{O}_5\text{F}_3$: 371.1107).

Euscapholide glucoside tetraacetate (4)

Compound 3 (5.7 mg) was acetylated as above to give the tetraacetate 4 (7.7 mg) as an amorphous powder. IR v_{\max} (CHCl_3) cm^{-1} : 1740, 1370, 1220, 1040; ^1H NMR (CDCl_3): δ 1.32 (3H, d , $J = 6.4$ Hz, 9- H_3), 1.73 (1H, ddd , $J = 14.2$, 6.8 and 5.4 Hz, 7- H_1), 2.00, 2.02, 2.03, 2.09 (each 3H, s , 4 \times OAc), 2.37 (2H, m , 5- H_2), 3.71 (1H, m , 5'-H), 4.05 (1H, m , 8-H), 4.14 (1H, dd , $J = 12.2$ and 2.5 Hz, 6'- H_1), 4.25 (1H, dd , $J = 12.2$ and 5.4 Hz, 6'- H_1), 4.49 (1H, m , 6-H), 4.58 (1H, d , $J = 7.8$ Hz, 1'-H), 4.95 (1H, dd , $J = 9.8$ and 7.8 Hz, 2'-H), 5.05 (1H, t , $J = 9.8$ Hz, 4'-H), 5.19 (1H, t , $J = 9.8$ Hz, 3'-H), 6.04 (1H, dd , $J = 9.8$ and 2.0 Hz, 3-H), 6.89 (1H,

m, 4-H); HR-FABMS (negative): m/z 485.1657 [$M-H$]⁻ (Calculated for C₂₂H₂₉O₁₂: 485.1659).

Enzymatic hydrolysis of euscapholide glucoside (3)

Compound **3** (20.1 mg) was dissolved in EtOH-H₂O (1:9, 10 ml) and β -glucosidase (ex almond) (purchased from Toyobo Co. Ltd., Osaka, Japan) (17.6 mg) was added. The mixture was incubated at 37° for 24 hr. The solution was extracted with EtOAc (10 ml \times 3). The EtOAc extracts were washed with sat. NaCl aq. solution, dried and evaporated *in vacuo* to give the aglucone (7.2 mg) as a colourless oil, $[\alpha]_D^{21} +116.4^\circ$ (MeOH, *c* 0.36), HR-EIMS: m/z 156.0736 [M]⁻ (Calculated for C₈H₁₂O₃: 156.0786). The compound was identified with the authentic sample of euscapholide (**1**) by comparisons of IR and ¹H and ¹³C NMR spectra.

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