



Chemotaxonomic value of tocopherols in Brassicaceae

Fernando D. Goffman*, Werner Thies, Leonardo Velasco

Institut für Pflanzenbau und Pflanzenzüchtung, Georg-August-Universität Göttingen, Von-Siebold-Str. 8, D-37075 Göttingen, Germany

Received 8 July 1998

Abstract

The significance of tocopherols as chemotaxonomic markers and their relationship with oil content and fatty acid profile was investigated in a collection of 91 species of the family Brassicaceae. Total tocopherols content ranged from 68 mg kg⁻¹ oil in *Diplotaxis viminea* to 2479 mg kg⁻¹ oil in *Schivereckia doerfleri*. The collection also showed wide variability for tocopherol composition. The average tocopherol profile consisted of 65.4% γ -, 28.7% α -, 5.1% δ - and 0.8% β -tocopherol. Individual tocopherols were found to have great taxonomic value in the Brassicaceae. © 1999 Elsevier Science Ltd. All rights reserved.

Keywords: Brassicaceae; Chemotaxonomy; Tocopherols; Vitamin E

1. Introduction

Tocopherols are a group of closely related derivatives of the phenolic tocol marked by an extensive ring alkylation (Larson, 1988). They occur in four forms of derivatives (α -, β -, γ - and δ -tocopherol; the α -derivative is also known as vitamin E), differing in the methylation of the tocol head group (1). The relative content of individual tocopherols is known to be characteristic of the seed oil of different cultivated plants. The germ oil of wheat (*Triticum aestivum*), for example, contains 60% of α -tocopherol and 23% of β -tocopherol and the oil of linseed (*Linum usitatissimum*) oil contains 76% of γ -tocopherol and 24% of plastochromanol-8, an unusual tocol derivative in plant oils (Balz, Schulte, & Thier, 1992). In the oil of soybean (*Glycine max* L.), d -tocopherol represents ca. 30% of the total tocopherols (Van Niekerk, & Burger, 1985; Yoshida, Hirooka, & Kajimoto, 1990). Conversely, α -tocopherol is the main fraction in the seed oil of sunflower (*Helianthus annuus*) (>95%) (Gustafsson, Haglund, & Johansson, 1993). *Brassica napus* contains ca. 65% of γ -tocopherol and 35% of α -tocopherol (Appelqvist, 1972; Goffman, & Becker, 1998).

Individual glucosinolates and fatty acids are widely recognized as constituents of systematic interest in Brassicaceae (Seigler, 1981). No information about the significance of tocopherols as chemotaxonomic markers is available. The objective of this study was to determine the chemotaxonomic value of tocopherols in Brassicaceae.

2. Results and discussion

Oil content, fatty acid composition and tocopherol contents in the seeds of the 91 Brassicaceae accessions are listed in Table 1. Variation for total tocopherol content was extremely wide, ranging from 68 mg kg⁻¹ oil in *Diplotaxis viminea* to 2479 mg kg⁻¹ oil in *Schivereckia doerfleri*. The average content was 995 mg kg⁻¹ oil. The collection also showed large variation for the relative presence of individual tocopherols, γ -tocopherol being the most abundant. The average tocopherol profile con-

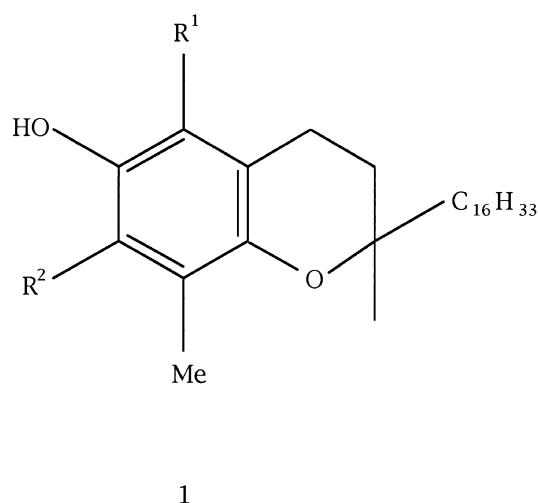


Fig. 1. Chemical formulae of tocopherols: α -tocopherol ($R^1 = \text{Me}$, $R^2 = \text{Me}$); β -tocopherol ($R^1 = \text{Me}$, $R^2 = \text{H}$); γ -tocopherol ($R^1 = \text{H}$, $R^2 = \text{Me}$); δ -tocopherol ($R^1 = \text{H}$, $R^2 = \text{H}$).

* Corresponding author. Fax: +49-551-39-4601; e-mail: fgoffma@gwdg.de.

Table 1

Oil content^a, fatty acid composition^b, total tocopherols content^c, and concentration of individual tocopherols^d in 91 accessions of Brassicaceae

| | Oil ^a | Fatty acids ^b | | | | | | | | Tocopherols | | | | |
|--|------------------|--------------------------|------|------|------|------|------|------|------|------------------|--------------------------|-------------------------|--------------------------|--------------------------|
| | | 16:0 | 18:0 | 18:1 | 18:2 | 18:3 | 20:1 | 22:1 | 24:1 | TTC ^c | α -T ^d | β -T ^d | δ -T ^d | γ -T ^d |
| Tribe Brassicaceae: subtribe Brassicinae | | | | | | | | | | | | | | |
| <i>Brassica elongata</i> | 17.5 | 3.8 | 1.1 | 8.9 | 21.3 | 33.3 | 3.7 | 20.2 | 1.3 | 1017 | 4.5 | 0.0 | 1.7 | 93.8 |
| <i>B. juncea</i> | 27.7 | 4.2 | 1.4 | 13.8 | 23.5 | 21.5 | 9.5 | 19.8 | 1.4 | 508 | 22.7 | 0.0 | 0.0 | 77.3 |
| <i>B. napus</i> | 40.6 | 3.7 | 1.0 | 45.6 | 18.5 | 10.3 | 3.5 | 14.7 | 0.4 | 581 | 42.0 | 0.0 | 0.9 | 57.2 |
| <i>B. nigra</i> | 26.3 | 4.0 | 1.1 | 10.4 | 17.9 | 19.2 | 5.0 | 32.6 | 1.7 | 430 | 7.1 | 0.0 | 2.7 | 90.3 |
| <i>B. rapa</i> | 31.4 | 2.6 | 1.4 | 18.4 | 13.6 | 10.7 | 10.1 | 37.6 | 1.1 | 606 | 46.3 | 0.0 | 2.1 | 51.6 |
| <i>Diplotaxis muralis</i> | 22.6 | 8.1 | 2.1 | 11.3 | 25.2 | 26.4 | 3.6 | 17.2 | 0.0 | 770 | 14.4 | 0.0 | 10.9 | 74.7 |
| <i>D. tenuifolia</i> | 25.4 | 6.0 | 1.7 | 11.3 | 18.7 | 33.8 | 4.0 | 18.8 | 0.5 | 746 | 5.8 | 0.0 | 3.2 | 91.0 |
| <i>D. viminea</i> | 22.1 | 8.3 | 2.4 | 8.9 | 20.6 | 27.7 | 3.3 | 20.8 | 0.6 | 68 | 0.0 | 0.0 | 0.0 | 99.9 |
| <i>Eruca sativa</i> | 19.0 | 5.7 | 0.8 | 10.8 | 15.4 | 19.5 | 4.4 | 37.3 | 1.3 | 813 | 13.0 | 0.0 | 2.6 | 84.4 |
| <i>Erucastrum nasturtifolium</i> | 25.9 | 4.3 | 1.2 | 12.8 | 22.2 | 32.7 | 5.6 | 16.8 | 0.9 | 672 | 15.5 | 0.0 | 2.3 | 82.2 |
| <i>Hirschfeldia incana</i> | 25.2 | 6.6 | 1.3 | 7.9 | 12.6 | 35.4 | 2.1 | 26.4 | 1.4 | 739 | 3.8 | 0.0 | 4.8 | 91.4 |
| <i>Sinapis alba</i> | 19.4 | 4.3 | 0.8 | 18.2 | 15.9 | 13.8 | 8.9 | 33.0 | 2.1 | 774 | 16.0 | 0.0 | 3.3 | 80.7 |
| <i>S. arvensis</i> | 24.3 | 3.9 | 0.9 | 9.2 | 16.0 | 19.3 | 10.8 | 34.2 | 1.2 | 679 | 4.2 | 0.0 | 3.6 | 92.1 |
| Tribe Brassicaceae: subtribe Raphaninae | | | | | | | | | | | | | | |
| <i>Calepina irregularis</i> | 13.0 | 7.2 | 1.7 | 11.2 | 13.5 | 32.2 | 20.9 | 6.3 | 0.3 | 795 | 3.9 | 0.0 | 2.9 | 93.2 |
| <i>Crambe maritima</i> | 32.2 | 4.1 | 0.6 | 26.7 | 25.3 | 4.8 | 14.5 | 21.6 | 0.4 | 1331 | 11.4 | 0.0 | 0.9 | 87.7 |
| <i>Rapistrum perenne</i> | 11.5 | 4.6 | 2.6 | 14.2 | 25.6 | 32.6 | 5.2 | 11.4 | 0.5 | 486 | 28.6 | 0.0 | 0.0 | 71.4 |
| <i>R. rugosum</i> | 12.7 | 5.0 | 1.3 | 7.6 | 13.7 | 28.9 | 3.2 | 31.4 | 1.9 | 506 | 12.5 | 0.0 | 0.0 | 87.5 |
| Tribe Brassicaceae: subtribe Moricandiinae | | | | | | | | | | | | | | |
| <i>Conringia orientalis</i> | 26.7 | 2.6 | 0.3 | 6.7 | 29.2 | 2.8 | 22.0 | 24.5 | 4.4 | 718 | 98.4 | 0.0 | 0.0 | 1.6 |
| <i>Moricandia arvensis</i> | 24.5 | 10.4 | 1.7 | 11.5 | 19.6 | 32.6 | 5.3 | 14.8 | 0.5 | 1054 | 0.8 | 0.0 | 6.6 | 92.6 |
| Tribe Heliophileae | | | | | | | | | | | | | | |
| <i>Heliophila longifolia</i> | 21.6 | 4.4 | 0.5 | 30.4 | 7.4 | 8.7 | 0.8 | 27.1 | 15.2 | 1424 | 1.0 | 0.0 | 3.9 | 95.1 |
| Tribe Lepidieae: subtribe Lepidiinae | | | | | | | | | | | | | | |
| <i>Coronopus didymus</i> | 14.7 | 7.4 | 3.1 | 16.0 | 18.1 | 39.6 | 8.7 | 1.8 | 0.5 | 871 | 14.8 | 0.0 | 4.7 | 80.5 |
| <i>C. squamatus</i> | 15.0 | 11.9 | 3.0 | 23.1 | 9.9 | 39.2 | 9.0 | 0.5 | 0.3 | 848 | 1.6 | 0.0 | 3.2 | 95.3 |
| <i>Lepidium campestre</i> | 13.9 | 4.3 | 1.0 | 12.4 | 11.1 | 44.1 | 4.3 | 18.0 | 1.5 | 1289 | 6.7 | 0.0 | 0.0 | 93.3 |
| <i>L. graminifolium</i> | 27.4 | 8.3 | 2.3 | 15.0 | 11.5 | 51.0 | 6.3 | 0.6 | 0.0 | 736 | 4.0 | 0.0 | 1.0 | 95.1 |
| <i>L. heterophyllum</i> | 19.0 | 5.6 | 1.8 | 21.8 | 10.7 | 35.6 | 5.9 | 15.0 | 0.8 | 885 | 1.8 | 0.0 | 1.8 | 96.4 |
| <i>L. sativum</i> | 19.0 | 8.8 | 2.0 | 25.7 | 10.5 | 37.6 | 8.6 | 3.0 | 0.6 | 1492 | 0.0 | 0.0 | 3.2 | 96.8 |
| <i>Biscutella auriculata</i> | 33.9 | 6.4 | 1.0 | 31.9 | 8.1 | 13.3 | 36.1 | 1.6 | 0.0 | 602 | 5.9 | 0.0 | 1.5 | 92.7 |
| Tribe Lepidieae: subtribe Iberidinae | | | | | | | | | | | | | | |
| <i>Iberis amara</i> | 19.4 | 4.4 | 0.3 | 19.8 | 17.6 | 17.9 | 4.6 | 29.1 | 3.9 | 618 | 88.3 | 0.0 | 0.0 | 11.7 |
| <i>I. umbellata</i> | 22.6 | 3.9 | 0.2 | 9.2 | 25.7 | 10.1 | 2.8 | 39.5 | 4.4 | 527 | 92.4 | 1.7 | 2.5 | 3.4 |
| Tribe Lepidieae: subtribe Thlaspidinae | | | | | | | | | | | | | | |
| <i>Aethionema grandiflora</i> | 13.7 | 7.5 | 1.4 | 10.9 | 11.8 | 63.6 | 0.7 | 0.8 | 0.0 | 2649 | 0.6 | 0.0 | 4.4 | 95.1 |
| <i>A. saxatile</i> | 21.5 | 8.2 | 3.4 | 14.9 | 8.7 | 56.4 | 2.6 | 0.2 | 0.0 | 825 | 0.0 | 0.0 | 2.3 | 97.7 |
| <i>Thlaspi alliaceum</i> | 26.6 | 3.6 | 0.8 | 15.4 | 21.2 | 5.3 | 5.9 | 42.6 | 2.8 | 568 | 72.2 | 0.0 | 0.0 | 27.8 |
| <i>T. alpestre</i> | 31.7 | 2.9 | 0.4 | 8.2 | 26.3 | 12.0 | 9.4 | 33.9 | 2.5 | 621 | 77.2 | 14.2 | 3.6 | 5.1 |
| <i>T. alpinum</i> | 21.6 | 2.9 | 0.3 | 10.1 | 21.1 | 14.3 | 12.7 | 29.5 | 3.8 | 626 | 70.4 | 2.2 | 2.2 | 25.2 |
| <i>T. arvense</i> | 31.1 | 2.9 | 0.4 | 13.5 | 24.3 | 18.2 | 10.5 | 23.6 | 2.6 | 557 | 79.8 | 0.0 | 0.0 | 20.2 |
| Tribe Lepidieae: subtribe Capsellinae | | | | | | | | | | | | | | |
| <i>Hornungia petraea</i> | 24.7 | 6.8 | 1.7 | 7.3 | 18.7 | 35.8 | 12.9 | 11.4 | 0.4 | 576 | 86.6 | 0.0 | 3.5 | 9.9 |
| Tribe Lepidieae: subtribe Cochleariinae | | | | | | | | | | | | | | |
| <i>Cochlearia officinalis</i> | 21.1 | 3.9 | 1.7 | 14.0 | 37.7 | 22.5 | 4.9 | 9.9 | 1.6 | 2009 | 9.7 | 4.3 | 44.1 | 42.0 |
| Tribe Euclidieae | | | | | | | | | | | | | | |
| <i>Bunias erucago</i> | 12.8 | 10.1 | 2.1 | 27.0 | 11.3 | 45.1 | 0.8 | 2.3 | 0.0 | 870 | 15.3 | 0.0 | 5.4 | 79.3 |
| <i>B. orientalis</i> | 11.1 | 4.7 | 1.6 | 12.7 | 21.5 | 57.3 | 0.4 | 0.9 | 0.2 | 696 | 2.6 | 0.0 | 7.8 | 89.6 |
| <i>Myagrum perfoliatum</i> | 10.9 | 6.1 | 1.3 | 10.7 | 16.2 | 29.8 | 5.6 | 23.0 | 1.9 | 470 | 0.0 | 0.0 | 5.9 | 94.1 |
| <i>Neslia paniculata</i> | 17.1 | 5.1 | 1.4 | 14.6 | 17.3 | 37.9 | 13.1 | 6.1 | 0.3 | 987 | 0.0 | 0.0 | 1.8 | 98.2 |

Table 1 (continued)

Oil content^a, fatty acid composition^b, total tocopherols content^c, and concentration of individual tocopherols^d in 91 accessions of Brassicaceae

| | Oil ^a | Fatty acids ^b | | | | | | | | Tocopherols | | | | |
|---|------------------|--------------------------|------|------|------|------|------|------|------|------------------|------------------|------------------|------------------|------------------|
| | | 16:0 | 18:0 | 18:1 | 18:2 | 18:3 | 20:1 | 22:1 | 24:1 | TTC ^c | α-T ^d | β-T ^d | δ-T ^d | γ-T ^d |
| Tribe Lunarieae | | | | | | | | | | | | | | |
| <i>Lunaria annua</i> | 30.1 | 2.1 | 0.3 | 29.4 | 6.7 | 1.5 | 1.9 | 37.5 | 20.0 | 821 | 30.4 | 6.5 | 2.4 | 60.7 |
| <i>L. rediviva</i> | 27.0 | 2.2 | 0.0 | 20.1 | 14.9 | 4.0 | 12.3 | 37.7 | 6.8 | 946 | 14.5 | 2.0 | 8.2 | 75.3 |
| <i>Peltaria turkmena</i> | 37.7 | 4.3 | 0.6 | 21.4 | 21.1 | 12.4 | 9.2 | 28.2 | 2.0 | 764 | 33.7 | 0.0 | 1.4 | 64.9 |
| Tribe Alysseae | | | | | | | | | | | | | | |
| <i>Alyssoides utriculata</i> | 13.0 | 6.9 | 1.4 | 13.5 | 16.7 | 55.5 | 0.6 | 2.7 | 0.4 | 734 | 77.9 | 0.0 | 1.1 | 21.1 |
| <i>Alyssum alyssoides</i> | 18.2 | 7.7 | 1.3 | 9.2 | 10.1 | 69.8 | 0.0 | 0.6 | 0.0 | 341 | 95.2 | 0.0 | 0.0 | 4.8 |
| <i>A. corymbosum</i> | 17.3 | 6.6 | 1.9 | 14.1 | 15.9 | 58.7 | 0.2 | 0.2 | 0.0 | 1123 | 20.6 | 0.0 | 1.0 | 78.4 |
| <i>A. granatense</i> | 12.2 | 5.9 | 1.3 | 15.9 | 14.6 | 59.4 | 0.4 | 0.9 | 0.0 | 703 | 97.7 | 0.0 | 0.0 | 2.3 |
| <i>A. murale</i> | 19.1 | 6.0 | 1.2 | 15.9 | 19.6 | 54.7 | 0.0 | 0.0 | 0.0 | 640 | 96.7 | 0.0 | 0.0 | 3.3 |
| <i>A. repens</i> | 16.4 | 6.8 | 0.9 | 10.1 | 14.4 | 65.3 | 0.0 | 0.0 | 0.0 | 1309 | 31.6 | 0.0 | 0.9 | 67.4 |
| <i>A. saxatile</i> | 14.2 | 5.7 | 1.1 | 12.0 | 20.3 | 56.5 | 0.0 | 0.0 | 0.0 | 981 | 85.6 | 0.0 | 0.0 | 14.4 |
| <i>Berteroa incana</i> | 21.6 | 4.7 | 1.6 | 15.1 | 19.4 | 55.4 | 0.6 | 1.5 | 0.2 | 906 | 82.1 | 1.0 | 1.5 | 15.3 |
| <i>Clypeola jouthlaspi</i> | 12.1 | 10.7 | 2.3 | 21.6 | 13.4 | 48.5 | 0.5 | 0.9 | 0.2 | 537 | 69.2 | 3.1 | 9.2 | 18.5 |
| <i>Fibigia clypeata</i> | 18.8 | 7.2 | 1.9 | 16.5 | 22.0 | 47.7 | 0.4 | 1.6 | 0.3 | 696 | 94.7 | 0.0 | 0.0 | 5.3 |
| <i>Lobularia maritima</i> | 28.9 | 4.1 | 4.3 | 31.8 | 9.2 | 11.6 | 35.6 | 0.4 | 0.2 | 582 | 20.8 | 0.0 | 3.6 | 75.6 |
| Tribe Drabeae | | | | | | | | | | | | | | |
| <i>Draba aurea</i> | 24.7 | 5.6 | 1.9 | 12.2 | 27.4 | 51.2 | 0.2 | 0.4 | 0.0 | 1533 | 2.1 | 0.0 | 4.8 | 93.1 |
| <i>D. incana</i> | 18.0 | 4.2 | 1.1 | 8.1 | 32.3 | 52.5 | 0.0 | 0.2 | 0.0 | 1763 | 3.8 | 0.0 | 4.7 | 91.5 |
| <i>D. magellanica</i> | 22.6 | 4.3 | 1.0 | 10.2 | 30.5 | 52.6 | 0.2 | 0.0 | 0.0 | 1963 | 3.4 | 0.0 | 2.7 | 93.9 |
| <i>Kernera saxatilis</i> | 28.3 | 4.0 | 0.7 | 10.4 | 41.9 | 15.5 | 5.7 | 13.6 | 1.0 | 802 | 98.7 | 1.3 | 0.0 | 0.0 |
| <i>Schivereckia doerfleri</i> | 16.4 | 6.6 | 1.2 | 7.5 | 27.9 | 51.6 | 0.3 | 0.8 | 0.0 | 2749 | 15.6 | 0.0 | 1.8 | 82.7 |
| Tribe Arabideae | | | | | | | | | | | | | | |
| <i>Arabis alpina</i> | 23.5 | 6.2 | 1.8 | 14.3 | 28.3 | 46.8 | 0.0 | 0.0 | 0.0 | 1091 | 10.6 | 0.4 | 24.2 | 64.8 |
| <i>A. caucasica</i> | 20.8 | 6.2 | 1.7 | 13.0 | 28.3 | 48.5 | 0.3 | 0.0 | 0.0 | 721 | 7.3 | 0.0 | 14.7 | 78.0 |
| <i>A. corymbiflora</i> | 20.9 | 9.4 | 2.7 | 14.2 | 24.1 | 46.9 | 0.0 | 0.0 | 0.0 | 703 | 27.9 | 0.0 | 2.7 | 69.4 |
| <i>A. glabra</i> | 30.3 | 5.3 | 1.0 | 6.8 | 30.6 | 31.4 | 9.3 | 8.9 | 0.3 | 1223 | 5.7 | 0.0 | 1.4 | 93.0 |
| <i>A. procurrens</i> | 25.0 | 5.7 | 2.3 | 16.4 | 33.3 | 40.1 | 0.3 | 0.0 | 0.0 | 1550 | 28.6 | 0.0 | 3.1 | 68.3 |
| <i>A. rosea</i> | 25.2 | 9.6 | 2.5 | 10.8 | 32.9 | 42.7 | 0.0 | 0.0 | 0.0 | 955 | 3.3 | 0.0 | 3.7 | 93.0 |
| <i>A. soyeri</i> | 20.1 | 8.1 | 1.9 | 11.9 | 35.0 | 41.4 | 0.0 | 0.0 | 0.0 | 1681 | 6.8 | 0.0 | 12.7 | 80.5 |
| <i>A. virginica</i> | 19.1 | 9.1 | 1.5 | 6.5 | 36.1 | 45.3 | 0.0 | 0.0 | 0.0 | 1804 | 20.4 | 0.0 | 16.0 | 63.7 |
| <i>Barbarea stricta</i> | 30.5 | 2.0 | 0.3 | 20.0 | 26.8 | 14.4 | 10.5 | 21.1 | 1.3 | 414 | 98.4 | 0.8 | 0.0 | 0.8 |
| <i>B. verna</i> | 31.0 | 3.4 | 0.4 | 10.7 | 20.2 | 9.0 | 4.7 | 44.7 | 1.6 | 394 | 93.4 | 2.5 | 1.6 | 2.5 |
| <i>B. vulgaris</i> | 27.5 | 2.6 | 0.6 | 25.1 | 23.1 | 8.4 | 10.4 | 24.7 | 2.1 | 382 | 98.1 | 0.0 | 0.0 | 1.9 |
| <i>Cardaminopsis neglecta</i> | 32.4 | 3.8 | 1.6 | 13.2 | 31.4 | 29.5 | 5.8 | 10.0 | 0.6 | 1059 | 12.2 | 0.0 | 4.1 | 83.7 |
| <i>Dentaria bulbifera</i> | 11.5 | 15.3 | 1.1 | 15.4 | 31.6 | 23.5 | 4.3 | 5.8 | 0.3 | 462 | 77.4 | 17.0 | 0.0 | 5.7 |
| Tribe Matthioleae | | | | | | | | | | | | | | |
| <i>Aubrieta gracilis</i> | 15.9 | 9.7 | 1.8 | 13.4 | 24.4 | 49.1 | 0.0 | 0.0 | 0.0 | 1823 | 4.2 | 0.0 | 2.4 | 93.4 |
| <i>Matthiola incana</i> | 20.8 | 8.0 | 2.5 | 13.6 | 9.0 | 65.3 | 0.5 | 0.8 | 0.0 | 2078 | 0.9 | 0.0 | 73.4 | 25.6 |
| Tribe Hesperideae | | | | | | | | | | | | | | |
| <i>Cheiranthus cheiri</i> | 20.7 | 4.2 | 0.6 | 10.7 | 21.4 | 25.2 | 5.9 | 23.6 | 1.4 | 914 | 1.6 | 0.0 | 8.5 | 90.0 |
| <i>Erysimum hieraciifolium</i> | 36.7 | 3.3 | 0.8 | 10.8 | 26.3 | 23.9 | 4.1 | 22.3 | 2.5 | 1147 | 20.7 | 0.0 | 3.6 | 75.8 |
| <i>E. marschallianum</i> | 33.1 | 4.1 | 1.2 | 6.1 | 32.2 | 26.1 | 2.5 | 17.7 | 2.1 | 1327 | 2.5 | 0.0 | 3.2 | 94.3 |
| <i>E. odoratum</i> | 29.4 | 4.2 | 1.5 | 9.7 | 28.3 | 30.5 | 3.9 | 14.3 | 1.1 | 1050 | 23.6 | 0.0 | 1.0 | 75.4 |
| <i>E. repandum</i> | 22.2 | 7.2 | 2.0 | 6.8 | 19.8 | 38.7 | 4.3 | 13.9 | 0.8 | 2012 | 1.4 | 0.0 | 3.8 | 94.8 |
| <i>E. wittmannii</i> | 32.0 | 5.0 | 0.9 | 7.0 | 28.9 | 27.1 | 2.1 | 18.6 | 1.4 | 1199 | 26.9 | 0.0 | 1.3 | 71.8 |
| <i>Hesperis laciniata</i> | 19.3 | 6.8 | 2.0 | 11.6 | 12.6 | 64.1 | 0.0 | 0.2 | 0.0 | 2031 | 9.2 | 0.0 | 27.6 | 63.3 |
| <i>H. matronalis</i> | 25.7 | 7.2 | 2.1 | 14.3 | 24.2 | 50.5 | 0.0 | 0.0 | 0.0 | 1865 | 4.2 | 0.0 | 34.8 | 61.0 |
| Tribe Sisymbrieae: subtribe Alliariinae | | | | | | | | | | | | | | |
| <i>Alliaria petiolata</i> | 26.2 | 2.5 | 0.2 | 10.8 | 19.1 | 6.0 | 6.2 | 45.8 | 6.4 | 880 | 15.2 | 0.0 | 1.3 | 83.5 |

(Continued on next page)

Table 1 (continued)

Oil content^a, fatty acid composition^b, total tocopherols content^c, and concentration of individual tocopherols^d in 91 accessions of Brassicaceae

| | Oil ^a | Fatty acids ^b | | | | | | | | Tocopherols | | | | |
|--|------------------|--------------------------|------|------|------|------|------|------|------|------------------|--------------------------|-------------------------|--------------------------|--------------------------|
| | | 16:0 | 18:0 | 18:1 | 18:2 | 18:3 | 20:1 | 22:1 | 24:1 | TTC ^c | α -T ^d | β -T ^d | δ -T ^d | γ -T ^d |
| Tribe Sisymbrieae: subtribe Sisymbriinae | | | | | | | | | | | | | | |
| <i>Sisymbrium austriacum</i> | 30.8 | 6.1 | 1.3 | 10.0 | 14.6 | 37.2 | 8.0 | 13.9 | 0.6 | 1200 | 2.4 | 0.0 | 5.1 | 92.4 |
| <i>S. loeselii</i> | 29.7 | 7.3 | 1.1 | 11.1 | 16.2 | 36.3 | 8.4 | 13.8 | 0.3 | 1244 | 2.7 | 0.0 | 3.8 | 93.5 |
| <i>S. orientale</i> | 22.2 | 9.4 | 0.9 | 6.4 | 13.3 | 41.4 | 3.6 | 17.3 | 0.6 | 913 | 4.4 | 0.0 | 2.0 | 93.6 |
| <i>S. strictissimum</i> | 35.6 | 3.9 | 0.8 | 8.1 | 19.8 | 36.2 | 6.4 | 15.6 | 0.8 | 960 | 42.1 | 12.9 | 9.4 | 35.7 |
| Tribe Sisymbrieae: subtribe Cameliniinae | | | | | | | | | | | | | | |
| <i>Camelina pilosa</i> | 25.1 | 7.5 | 2.3 | 12.7 | 22.7 | 36.0 | 11.5 | 1.9 | 0.5 | 846 | 0.9 | 0.0 | 0.9 | 98.1 |
| Tribe Sisymbrieae: subtribe Descuriinae | | | | | | | | | | | | | | |
| <i>Descurainia sophia</i> | 30.3 | 5.9 | 1.4 | 9.5 | 17.3 | 44.9 | 8.2 | 7.1 | 0.6 | 1585 | 1.3 | 0.0 | 2.9 | 95.8 |
| <i>Hugueninia tanacetifolia</i> | 24.9 | 6.8 | 1.2 | 9.1 | 20.1 | 30.6 | 11.6 | 15.2 | 0.9 | 1076 | 1.9 | 0.0 | 1.5 | 96.6 |

^a Expressed as % wt/wt.^b Expressed as % of total fatty acids.^c Tocopherols: TTC = total tocopherols content, expressed as mg kg⁻¹ oil.^d α -T = α -tocopherol, β -T = β -tocopherol, δ -T = δ -tocopherol, γ -T = γ -tocopherol, all individual tocopherols expressed as % of total tocopherols.

sisted of 65.4% γ -, 28.7% α -, 5.1% δ - and 0.8% β -tocopherol. α -Tocopherol was not detected (<1%) in ten accessions (*Diplotaxis viminea*, *Moricandia arvensis*, *Heliphila longifolia*, *Lepidium sativum*, *Aethionema grandiflora*, *A. saxatile*, *Myagrurn perfoliatum*, *Neslia paniculata*, *Matthiola incana* and *Camelina pilosa*), but it was the predominant tocopherol (>50%) in 21 accessions. γ -Tocopherol was absent in only two accessions (*Kernera saxatilis* and *Barbarea stricta*), being predominant in 67 accessions. The highest levels of δ -tocopherol were found in *Matthiola incana* (73.4%), *Cochlearia officinalis* (44.1%), *Hesperis matronalis* (34.8%), *H. laciniata* (27.6), *Arabis alpina* (24.2%), *A. virginica* (16.0%) and *A. soyeri* (12.7); in 66 of the 91 accessions, this tocopherol derivative was present as a secondary constituent (>1%) only. β -Tocopherol was found in no more than 11 of the accessions, the highest values occurring in seed oils of *Dentaria bulbifera* (17.0%), *Thlaspi alpestre* (14.2%) and *Sisymbrium strictissimum* (12.9%).

The average oil content was 23.0%, ranging from 10.9% in *Myagrurn perfoliatum* to 40.6% in *Brassica napus*. Kumar and Tsunoda (1980) reported a variation for oil content in Brassicaceae between 6 and 49% after reviewing published results from different sources. Since our results were obtained from plants grown under the same environment, they provide reliable information on the relative oil content values of the different species. The variability for the fatty acid composition of the seed oil altogether was found within the limits determined in previous studies and reviewed by Kumar and Tsunoda (1978).

2.1. Study of correlations

Correlation coefficients between oil content, fatty acids and tocopherols are shown in Table 2. Total tocopherols content was positively correlated with linolenic and negatively correlated with eicosenoic and erucic acid. A positive correlation between tocopherols content and linolenic acid also had been revealed in rapeseed mutants (Röbbelen, & Thies, 1980). Moreover, Kamal-Eldin and Andersson (1997) analysed data of oil samples from 14 botanical species and reported a positive correlation between tocopherols content and linoleic and linolenic acid. Oil content in the present study was negatively correlated with the main saturated fatty acids (palmitic and stearic) and with linolenic acid and positively correlated with eicosenoic and erucic acid. After grouping all fatty acids by their level of unsaturation (data not shown), the oil content was negatively correlated with the sum of saturated ($r = -0.43$) and with the sum of polyunsaturated fatty acids ($r = -0.43$), but it was positively correlated with the sum of monounsaturated fatty acids ($r = 0.44$). These results suggest that the species accumulating the highest amounts of oil in their seeds show preference for synthesizing monounsaturated fatty acids. None of these correlations were found in the above mentioned study of Kumar and Tsunoda (1978) on 159 species of Brassicaceae. The present correlations involving fatty acids were basically similar to those reported by Kumar and Tsunoda (1978). A strong negative correlation ($r = -0.80$) was found between linolenic and erucic acid, which is easily explained taking into account that the accessions exhibited basically two main patterns,

Table 2

Correlation coefficients between oil content (%), individual fatty acids (% of the total fatty acids), total tocopherols content (mg kg⁻¹ oil) and concentration of individual tocopherols (% of the total tocopherols) in a set of 91 accessions of Brassicaceae. Significance limits: $r = \pm 0.21$ (5%), $r = \pm 0.27$ (1%)

| | C16:0 | C18:0 | C18:1 | C18:2 | C18:3 | C20:1 | C22:1 | C24:1 | TTC ^a | α -T | β -T | δ -T | γ -T |
|-------------|-------|-------|-------|-------|-------|-------|-------|-------|------------------|-------------|------------|-------------|-------------|
| Oil | -0.47 | -0.30 | 0.16 | 0.20 | -0.53 | 0.33 | 0.37 | 0.18 | -0.10 | 0.03 | 0.08 | -0.06 | -0.02 |
| C16:0 | | 0.56 | -0.05 | -0.12 | 0.54 | -0.24 | -0.59 | -0.41 | 0.15 | -0.22 | 0.08 | 0.13 | 0.17 |
| C18:0 | | | 0.10 | -0.14 | 0.49 | -0.02 | -0.65 | -0.45 | 0.11 | -0.36 | -0.21 | 0.24 | 0.30 |
| C18:1 | | | | -0.37 | -0.30 | 0.29 | -0.03 | 0.27 | -0.19 | 0.06 | -0.02 | -0.05 | -0.04 |
| C18:2 | | | | | -0.10 | -0.20 | -0.09 | -0.22 | 0.17 | 0.15 | 0.13 | 0.03 | -0.17 |
| C18:3 | | | | | | -0.54 | -0.80 | -0.52 | 0.42 | -0.17 | -0.20 | 0.25 | 0.11 |
| C20:1 | | | | | | | 0.19 | 0.02 | -0.30 | 0.00 | 0.01 | -0.19 | 0.05 |
| C22:1 | | | | | | | | 0.55 | -0.40 | 0.17 | 0.14 | -0.22 | -0.11 |
| C24:1 | | | | | | | | | -0.11 | 0.07 | 0.18 | -0.09 | -0.06 |
| TTC | | | | | | | | | | -0.45 | -0.13 | 0.44 | 0.33 |
| α -T | | | | | | | | | | | 0.27 | -0.25 | -0.95 |
| β -T | | | | | | | | | | | | 0.03 | -0.36 |
| δ -T | | | | | | | | | | | | | -0.05 |

^a Abbreviations of tocopherols as for Table 1.

characterized by high linolenic and high erucic acid content, respectively. Palmitic and stearic acids were positively correlated with linolenic acid and negatively correlated with erucic acid.

2.2. Taxonomic potential of tocopherols

The investigated collection of Brassicaceae displayed great variability for total tocopherols content and tocopherol profile. Based on the classification of Schulz (1936), it is possible to conclude that the tocopherols have a significant taxonomic meaning, at least as important as that of the fatty acids have. Some clear lines can be observed: for example, the tribe Brassiceae was characterized by a high concentration of γ -tocopherol. The only exception was *Conringia orientalis*, but this accession was marked by a fatty acid profile (low linolenic acid and high nervonic acid, Table 1) atypical for this taxonomic group. Likewise, a high number of the accessions with the highest α -tocopherol percentage belonged to the tribe Alysseae. In some of the species an interesting similarity occurred from fatty acids and tocopherols for the informations. For example, two genera of the subtribe Thlaspidinae (tribe Lepidieae), were analysed. The accessions of *Aethionema* had a high linolenic acid and practically no erucic acid contents, while the accessions of *Thlaspi* showed considerably lower linolenic and higher erucic acid contents. Such differences were paralleled by a completely different profile of tocopherols, with γ -tocopherol being the predominant in *Aethionema* and of α -tocopherol in *Thlaspi*. A similar case was observed in the subtribe Iberidinae (tribe Lepidieae) and in the tribe Drabaeae. The accessions of the tribe Arabideae are another excellent example of the taxonomic potential of tocopherols and of the unsatisfactory taxonomic assign-

ments within this family (Warwick, & Black, 1993). According to the tocopherols information, the genera *Arabis* and *Cardaminopsis*, with high γ -tocopherol content, are clearly different from the genera *Barbarea* and *Dentaria*, characterized by high α -tocopherol content. Schulz (1936) assigned all these genera to the same tribe, while Janchen (1942) divided the tribe into the subtribe Cardamininae (including *Barbarea* and *Dentaria*) and the subtribe Arabidinae (including *Cardaminopsis* and *Arabis*). Tocopherols composition clearly supports the latter subtribe division. In conclusion, this study demonstrates that tocopherols possess an important chemotaxonomic value in Brassicaceae and may play a major role in taxonomic studies on this family.

3. Experimental

3.1. Plant materials

This study included 91 germplasm accessions from 91 species (48 genera) of the family Brassicaceae. The samples belonged to the Brassicaceae collection were kindly provided by the Botanical Garden of the University of Göttingen, Germany. The accessions had been multiplied in 1996/97 in the Botanical Garden. The complete list of the germplasm collection can be found in the Index Seminum (1997). Taxonomic classification was based on Schulz (1936).

3.2. Analysis of seed oil content

Oil content was determined nondestructively by near-infrared reflectance spectroscopy (NIRS). Ca. 300 mg of intact-seed samples were analysed on a NIRS instrument.

Oil content was calculated from NIRS spectral information by using a calibration equation developed from a wide range of genera of Brassicaceae (Velasco, Goffman, & Becker, 1998).

3.3. Analysis of fatty acid composition

Fatty acid composition of seed oil was determined by the GLC method described by Thies (1971), using a FFAP capillary column, 25 m × 0.25 mm and a FID detector. The temperatures of oven, detector and injector were 200°C, 250°C and 250°C, respectively. The carrier gas was hydrogen, at a pressure of 100 kPa. Two ml of sample were injected, at a split rate of 1:70.

3.4. Analysis of tocopherols

Tocopherols content was determined by HPLC as described by Thies (1997), with a fluorescence detector ($I_{\text{ex}} = 295$ nm and $I_{\text{em}} = 330$ nm), a C-18 diol column (250 × 3 mm), and isooctane/tert-butyl-methylether (94/6) as eluent at a flow rate of 0.7 ml/min. β -Tocopherol was used as internal standard. The samples were first analysed without the internal standard to check the presence of β -tocopherol. A calibration curve of β -tocopherol was used as external standard for the quantification of samples containing this tocopherol derivative.

Acknowledgements

Financial support for this investigation was provided by Niedersächsisches Ministerium für Wissenschaft und Kultur, Forschungsstelle für Biologische Rohstoffe, Göttingen. The authors thank Professor G. Röbbelen for useful suggestions on the manuscript and the Botanical Garden of the University of Göttingen, for kindly providing the germplasm material. Michaela Grote, Chri-

stine Reuter and Uwe Ammermann provided technical assistance. LV received a grant from the Dirección General de Enseñanza Superior (Ministerio de Educación y Cultura, Spain). FDG was funded by Deutscher Akademischer Austauschdienst (DAAD).

References

- Appelqvist, L. Å. (1972). In *Rapeseed, Cultivation, Composition, Processing and Utilization* (pp. 145–147). Elsevier Publishing Co., Amsterdam.
- Balz, M., Schulte, E., & Thier, H. P. (1992). *Fat Science Technology*, 94, 209.
- Goffman, F. D., & Becker, H. C. (1998). *Vorträge Pflanzenzüchtung*, 42, 105.
- Gustafsson, I. B., Haglund, A., & Johansson, L. (1993). *Journal of the Science of Food and Agriculture*, 62, 273.
- Index Seminarum (1997) (pp. 12–14). Botanischer Garten der Georg-August-Universität Göttingen, Germany.
- Janchen, E. (1942). *Österreichische Botanische Zeitschrift*, 91, 1.
- Kamal-Eldin, A., & Andersson, R. (1997). *Journal of the American Oil Chemists Society*, 74, 375.
- Kumar, P. R., & Tsunoda, S. (1978). *Journal of the American Oil Chemists Society*, 55, 320.
- Kumar, P. R., & Tsunoda, S. (1980). In *Brassica Crops and Wild Allies* (pp. 235–252). Japan Scientific Societies Press, Tokyo.
- Larson, R. A. (1988). *Phytochemistry*, 27, 969.
- Röbbelen, G., and Thies, W. (1980). In *Brassica Crops and Wild Allies* (pp. 253–283). Japan Scientific Societies Press, Tokyo.
- Schulz, O.E. (1936). In *Die natürlichen Pflanzenfamilien* (pp. 227–658). Wilhelm Engelmann Verlag, Leipzig.
- Seigler, D.S. (1981). In *The Biochemistry of Plants: a Comprehensive Treatise* (Vol. 7, pp. 139–176). Academic Press, New York.
- Thies, W. (1971). *Zeitschrift Pflanzenzüchtung*, 65, 181.
- Thies, W. (1997). *Angewandte Botanik*, 71, 62.
- Van Niekerk, P. J., & Burger, A. E. C. (1985). *Journal of the American Oil Chemists Society*, 62, 531.
- Velasco, L., Goffman, F.D., & Becker, H.C. (1998). *Journal of the American Oil Chemists Society* (submitted).
- Warwick, S. I., & Black, L. D. (1993). *Canadian Journal of Botany*, 71, 906.
- Yoshida, H., Hirooka, N., & Kajimoto, G. (1990). *Journal of Food Science*, 55, 1412.