



An amide of L-threo- γ -hydroxyglutamic acid from *Justicia ghiesbreghtiana*

Peter Lorenz^a, Frank R. Stermitz^{a,*}, Lotfy D. Ismail^b

^aDepartment of Chemistry, Colorado State University, Fort Collins, CO 80523, USA

^bDepartment of Pharmacognosy, Al-Azhar University, Nasr City, Cairo, Egypt

Received 2 December 1998; received in revised form 3 March 1999; accepted 3 March 1999

Abstract

A new amide of threo- γ -hydroxyglutamic acid, justiciamide, was isolated from *Justicia ghiesbreghtiana* and shown to be (–)-N-(2-hydroxy-4,5-dimethoxyphenyl)-(2 S,4 S)- γ -hydroxyglutamic acid. Justiciamide is the first amide of an uncommon amino acid found in this genus. © 1999 Elsevier Science Ltd. All rights reserved.

Keywords: *Justicia ghiesbreghtiana* Lem; Acanthaceae; γ -Hydroxyglutamic acid amide; Isolation

1. Introduction

The genus *Justicia* comprises about 600 species of perennial herbaceous plants and bushes (Mabberley, 1997). Lignans have been reported from this genus, some of which show biological activity (Chen, Hsin, & Huang, 1995; Rajasekhar, Subbaraju, Ravikumar, & Chandramohan, 1998; Ghosal, Banerjee, & Srivastava, 1979; Ghosal, Banerjee, & Jaiswal, 1980; Trujillo, Jorge, Navarro, & Boada, 1990; Asano, Chiba, Tada, & Yoshii, 1996; Olaniyi & Powell, 1980). The only aniline derivatives which had been found were 2-aminobenzyl alcohol and 2-(2-aminobenzylamino)benzyl alcohol from *J. gendarussa* (Chakravarty, Dastidar, & Pakrashi, 1982). Recently, however, we reported the isolation and synthesis of the α -malamic acid derivative 1 from the Mexican species *Justicia ghiesbreghtiana* Lem. (Ismail, Lorenz, & Stermitz, 1998). This plant, which seems to be known in the botanical garden literature under the synonym *J. spicigera* Schlechtend., has been used in traditional folk medicine as a medicinal herb and for dye purposes (Graham, 1998; Dominguez, Achenbach, González, & Ferré D'Amore,

1990; Euler & Alam, 1982). In the course of attempting to isolate additional 1 for further study, a new plant collection was made which yielded little or no 1, but a different, also novel amide.

2. Results and discussion

From 150 g of *J. ghiesbreghtiana*, 134 mg of a pure compound was isolated, which was shown by spectroscopic and chemical methods to be N-(2-hydroxy-4,5-dimethoxyphenyl)-(2 S,4 S)- γ -hydroxyglutamic acid (2), herein named justiciamide. The molecular formula for justiciamide (2) was established as C₁₃H₁₈N₂O₇ by HR-FAB mass spectral analysis. This formula, requiring six units of unsaturation, was fully supported by spectral data (Table 1). The noncrystalline solid had [α]_D –2.7° (c 0.07, H₂O) and showed on TLC a positive reaction with ninhydrin (NH₂-group). This observation, along with two carbonyl signals at δ 170.80 and 170.88 in the ¹³C NMR spectrum as well as strong IR absorptions at 1554, 1630 and 1671 cm^{–1}, suggested that the unknown 2 was an amino acid amide. The presence of two OCH₃ groups was shown by the observation of singlets at δ 3.69 and 3.76 in the ¹H NMR spectrum. Furthermore, a bathochromic

* Corresponding author. Fax: +1-970-491-5610.

E-mail address: frslab@lamar.colostate.edu (F.R. Stermitz)

Table 1
NMR spectral data for compound 2 (DMSO- d_6)

No.	^{13}C , δ , CH_n	^1H , δ (mult, J , no. of H)	HMBC
1	—	9.24 (bs, 1H)	C2, C7, C8, C12
2	170.80 (C=O)	—	—
3	70.73 (CH)	4.14 (dd, 2.4, 11.2, 1H)	C2, C4, C5
4	35.34 (CH_2)	2.22 (ddd, 2.4, 14.0, 11.2, 1H) 1.74 (ddd, 3.2, 10.4, 14.0, 1H)	C2, C3, C5, C6
5	52.82 (CH)	3.52 (dd, 2.8, 10.4, 1H)	C3, C6
6	170.88 (C=O)	—	—
7	118.17 (C)	—	—
8	142.87 (C)	—	—
9	100.62 (CH)	6.57 (s, 1H)	C7, C10, C11
10	142.92 (C)	—	—
11	140.42 (C)	—	—
12	106.01 (CH)	7.96 (s, 1H)	C7, C8, C9, C10, C11
13	56.64 (OCH_3)	3.69 (s, 3H)	C10
14	56.09 (OCH_3)	3.76 (s, 3H)	C11

baseshift (UV_{max} 296 nm to 304 nm) and a broad IR absorption at 3126 cm^{-1} showed the presence of a phenolic OH-group. The ^1H NMR spectrum (Table 1) also revealed three spin systems: two methine units at δ 3.52 and 4.14 and one methylene unit at δ 1.74 and 2.22. Singlets at δ 6.57 and 7.96 showed the presence of a 1,2,4,5-tetrasubstituted benzene ring. Spectral similarities with the α -malamidic acid derivative 1, suggested the same aromatic substitution pattern for the unknown. Further NMR experiments involving HMQC, DEPT as well as ^1H decoupling led to the determination of the structure as 2 (stereochemistry unknown). Elemental analysis data suggested that justiciamide crystallized as a half-hydrate.

To determine the stereochemistry of 2, it was hydrolyzed with 6 N HCl to yield 2-hydroxy-3,4-dimethoxyaniline hydrochloride and 2 *S*,4 *S*- γ -hydroxyglutamic acid. The structure of the latter was shown by comparison to an authentic sample synthesized as follows. A mixture of *threo*- and *erythro*- γ -hydroxyglutamic

acids (3) was prepared according to Kaneko, Lee, & Hanafusa (1962) (Fig. 1). The mixture in aqueous HCl was treated with HCl gas, from which solution lactone 4 crystallized. The lactone was converted back into *threo*- γ -hydroxyglutamic acid (3a) by stirring with 6 N aq. HCl. The ^1H - and ^{13}C -NMR spectra for 3a were identical with those of the amino acid isolated from the hydrolysis of 2. The *erythro*-acid 3b does not form a lactone and its hydrochloride was obtained from the mother liquor of the lactone-forming reaction. The 3b NMR spectra were different from those of 3a and the isolated hydrolysis product from 2. Thus, the final structure of justiciamide was shown to be 2.

2 *S*, 4 *S*- γ -Hydroxyglutamic acid (3a) has previously been found as a constituent of *Phlox* species (Polemoniaceae) (Bell, Meier, & Sørensen, 1981; Virtanen & Hietala, 1955), *Hemerocallis fulva* (Liliaceae) (Fowden & Steward, 1957) and *Linaria vulgaris* (Scrophulariaceae) (Hatanaka, 1962). Two 3a amides of aliphatic amines were reported from

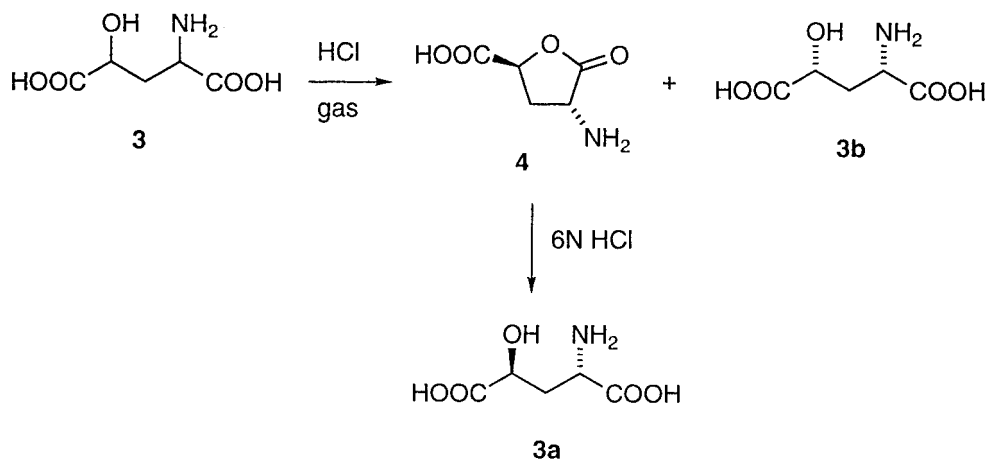


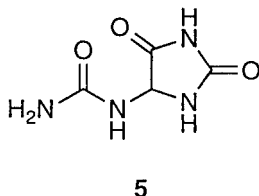
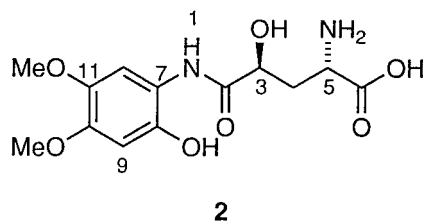
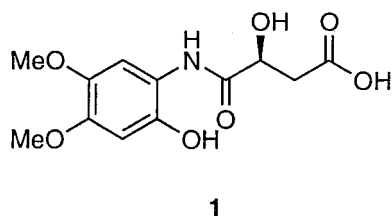
Fig. 1. Synthesis of *threo*- and *erythro*- γ -hydroxyglutamic acids.

Staphylea pinnata L. (Staphyleaceae) (Grove, Weisleder, & Daxenbichler, 1973). Our finding of 2, in contrast to 1 in the previous work, is as yet inexplicable since collection was from the same plant source at approximately the same season, although five years later.

A more polar chromatographic fraction yielded 17.4 mg of *rac*-allantoin (5), characterized by comparison of ^1H and ^{13}C NMR spectra with literature data and melting point (mp 229–230°C; lit. 230°C) (The Aldrich Library, 1997; Coxon, Fatiadi, Sniegowski, Hertz, & Schaffer, 1977). Compound 5 was previously reported from *Justicia spicigera* (Dominguez et al., 1990).

Justiciamide was inactive against gram-positive and gram-negative bacteria as well as against yeasts (*Candida albicans*, *Saccharomyces cerevisiae*). Compounds 1 and 2 were not cytotoxic in the brine shrimp test (*Artemia salina*) ($\text{ID}_{50} \approx 700$ ppm, after 24 h).

methanol was removed by rotaevaporation and the residual H_2O layer extracted with ether (3×300 ml). Evaporation of the H_2O from the water layer yielded an olive green foam (29.7 g), which was chromatographed on reverse phase silica gel (C_{18}), with water–MeOH (gradient: MeOH). Ten fractions were collected. Fractions 3 and 4 (water–MeOH 90:10 and 80:20) were unified and gave 1.70 g of a brown residue. The residue was rechromatographed on silica gel 60 (normal phase), solvent: CHCl_3 –MeOH (gradient: MeOH). Seven fractions were collected. Fractions 6 and 7 (100% MeOH) yielded 0.42 g of almost pure 2. Final purification was achieved by preparative HPLC on an ODS RP (C_{18}) HPLC column (25 cm), solvent: 40% MeOH– H_2O (UV detector) to yield 87 mg of pure 2. Another fraction of the first RP silica gel chromatography yielded further pure 2 (47 mg). **Compound 2.** ^1H (400 MHz) and ^{13}C (300 MHz) NMR: Table 1. Mp. 185°C, $[\alpha]_{\text{D}}^{25} -2.7^\circ$ (c 0.07, H_2O). UV (H_2O) λ_{max}



3. Experimental

3.1. Plant material

Aerial plants of *J. ghiesbreghtiana* were collected in December 1997 in the Giza zoological garden, Cairo, Egypt. The plant was identified by Professor Dr Nabeil El Hadedi, Department of Botany. A voucher specimen was deposited in the Department of Pharmacognosy herbarium, Al-Azhar University.

3.2. Extraction and isolation

150 g of air dried and powdered aerial parts were extracted with 2:3 EtOAc–*n*-hexane. The marc was dried and extracted twice with 2 l MeOH (containing 10% H_2O). From the unified MeOH extracts the

(log ϵ) 296 (5775), 244 nm (6220). IR (KBr) ν_{max} 3393, 3126, 1671, 1630, 1554, 1508, 1339, 1205, 1035, 907 cm^{-1} . HR FAB MS (MH^+) obsd. 315.1189 (calcd for $\text{C}_{13}\text{H}_{19}\text{N}_2\text{O}_7$, 315.1192). Anal. Calcd for $\text{C}_{13}\text{H}_{18}\text{N}_2\text{O}_7 \cdot 0.5 \text{H}_2\text{O}$: C, 48.30%, H, 5.92%, N, 8.66%. Found: C, 48.34%, H, 5.98%, N, 8.65%.

3.3. Hydrolysis of 2

A solution of 400 mg of 2 in 5 ml 6N HCl was stirred with a magnetic stirrer bar in a capped scintillation vial at 90°C for 12 h. The crude brown hydrolysate was filtered over Celite (rinsed with H_2O) and the HCl– H_2O removed by rotaevaporation. The residue was partially purified by vacuum liquid chromatography (VLC) on C_{18} -RP silica (solvent MeOH–water; gradient MeOH). A second VLC of the first fraction (352 mg) on silica gel (solvent: CHCl_3 –MeOH; gradi-

ent MeOH), yielded 49 mg of 4-hydroxyglutamic acid hydrochloride, $[\alpha]_D^{25} + 12$ (c 0.98, 5N HCl), lit. +4.5 (Lee & Kaneko, 1973). The ^1H and ^{13}C NMR data were essentially identical with those of a synthetic sample of *threo*- γ -hydroxyglutamic acid hydrochloride (3a), prepared as follows.

3.4. Synthesis of *threo*- and *erythro*- γ -hydroxyglutamic acid hydrochloride (3a, 3b)

Tetraethyl-1-acetamido-3-chloropropane-1,1',3,3'-tetracarboxylate (mp 90–92°C; mp_{lit.} = 91–92°C), prep'd according to Kaneko et al. (1962) was hydrolyzed with conc HCl (37%) to give an *erythro*/*threo* mixture, 3. Treatment of the mixture in acid with HCl gas yielded a crystalline *threo*-lactone hydrochloride (4) while the *erythro*-acid (3b), which does not form a lactone, stayed in solution (Fig. 1) (Kaneko et al., 1962; Lee & Kaneko, 1973). The *threo*- γ -hydroxyglutamic acid lactone hydrochloride (4) was transformed to 3a hydrochloride by stirring with 6 N HCl (90°C, 6 h). Evaporation of the mother liquor from the lactone formation and recrystallization of the residue yielded 3b HCl.

3.5. *Threo*- γ -hydroxyglutamic acid lactone HCl (4)

(mp 239–240°C, dec; mp_{lit.} 230–232°C); ^1H NMR (DMSO- d_6): δ 2.30 (*dd*, 1H, J = 11.7 Hz, CH₂), 2.87 (*ddd*, 1H, J = 6.6, 9.0, 12.3 Hz, CH₂), 4.45 (*dd*, 1H, J = 9.0, 11.7 Hz, CH), 5.04 (*dd*, 1H, J = 6.6, 10.8 Hz, CH), 9.04 (*bs*, 1H, COOH). ^{13}C NMR (DMSO- d_6): δ 30.19 (CH₂), 48.26 (CH), 73.49 (CH), 169.29 (C=O), 171.93 (C=O); anal. C, 33.19%, H, 4.51%, N, 7.46%, calcd for C₅H₈NO₄Cl, C 33.07%, H, 4.44%, N, 7.71%.

3.6. *Threo*- γ -hydroxyglutamic acid HCl (3a)

^1H -NMR (DMSO- d_6): δ 2.03 (*dd*, 1H, J = 3.6, 10.5 Hz, CH₂), 2.15 (*dd*, 1H, J = 3.6, 9.3 Hz, CH₂), 3.92 (*m*, 1H, CH), 4.45 (*m*, 1H, CH); ^{13}C NMR (DMSO- d_6): δ 34.21 (CH₂), 49.47 (CH), 66.00 (CH), 170.73 (C=O), 174.48 (C=O).

3.7. *Erythro*- γ -hydroxyglutamic acid HCl (3b)

^1H NMR (DMSO- d_6): δ 1.97 (*ddd*, 1H, J = 3.6, 5.1, 10.5 Hz, CH₂), 2.20 (*ddd*, 1H, J = 3.3, 7.5, 10.8 Hz,

CH₂), 3.95 (*m*, 1H, CH), 4.24 (*dd*, 1H, J = 3.3, 10.5 Hz, CH); ^{13}C NMR (DMSO- d_6): δ 34.21 (CH₂), 49.73 (CH), 66.72 (CH), 170.55 (C=O), 174.63 (C=O).

Acknowledgements

This work was supported by National Science Foundation grant CHE 9619213. Mass spectra were obtained on instruments supported by National Institutes of Health shared instrumentation grant GM49631.

References

- Asano, J., Chiba, K., Tada, M., & Yoshii, T. (1996). *Phytochemistry*, 42, 713.
- Bell, E. A., Meier, L. K., & Sørensen, H. (1981). *Phytochemistry*, 20, 2213.
- Chakravarty, A. K., Dastidar, P. P. G., & Pakrashi, S. C. (1982). *Tetrahedron*, 38, 1797.
- Chen, C. C., Hsin, W. C., & Huang, Y. L. (1995). *J. Nat. Prod.*, 61, 227.
- Coxon, B., Fatiadi, A. J., Sniegowski, L. T., Hertz, H. S., & Schaffer, R. (1977). *J. Org. Chem.*, 42, 3132.
- Dominguez, X. A., Achenbach, H., González, ChC, & Ferré D'Amore, A. R. (1990). *Rev. Latinoamer. Quím.*, 21, 142.
- Euler, K. L., & Alam, M. (1982). *J. Nat. Prod.*, 45, 220.
- Fowden, L., & Steward, F. C. (1957). *Ann. Botany N.S.*, 21, 53.
- Ghosal, S., Banerjee, S., & Jaiswal, D. K. (1980). *Phytochemistry*, 19, 332.
- Ghosal, S., Banerjee, S., & Srivastava, R. S. (1979). *Phytochemistry*, 18, 503.
- Graham, V. A. (1988). *Kew Bull.*, 43, 612.
- Grove, M. D., Weisleder, D., & Daxenbichler, M. E. (1973). *Tetrahedron*, 29, 2715.
- Hatanaka, S. (1962). *Acta Chem. Scand.*, 16, 513.
- Ismail, L. D., Lorenz, P., & Stermitz, F. R. (1998). *J. Nat. Prod.*, 61, 1174.
- Kaneko, T., Lee, Y. K., & Hanafusa, T. (1962). *Bull. Chem. Soc. Jpn*, 35, 875.
- Lee, Y. K., & Kaneko, T. (1973). *Bull. Chem. Soc. Jpn*, 46, 3494.
- Mabberley, D. J. (1997). *The Plant Book*. Melbourne: Cambridge University Press.
- Olaniyi, A. A., & Powell, J. W. (1980). *J. Nat. Prod.*, 43, 482.
- Rajasekhar, D., Subbaraju, G. V., Ravikumar, K., & Chandramohan, K. (1998). *Tetrahedron*, 54, 13,227.
- The Aldrich Library of ^{13}C and ^1H FT NMR Spectra (1997) Aldrich Chemical Company, Inc., Ed. 1, Vol. 1.
- Trujillo, J. M., Jorge, R., Navarro, E., & Boada, J. (1990). *Phytochemistry*, 29, 2991.
- Virtanen, A. I., & Hietala, P. K. (1955). *Acta. Chem. Scand.*, 9, 175.