



Tirucallane triterpenoids from *Dysoxylum hainanense*

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Abstract

Four tirucallane derivatives, 3 β ,22*S*-dihydroxy-tirucalla-7,24-dien-23-one, 22,23-epoxy-tirucalla-7-ene-3 β ,24,25-triol, 3 β ,25-dihydroxy-tirucalla-7,23-diene, 23,26-dihydroxy-tirucalla-7,24-dien-3-one, together with two known triterpenoids, 24,25-epoxy-3 β ,23-dihydroxy-7-tirucallene, tirucalla-7,24-diene-3 β ,23-diol, were isolated from *Dysoxylum hainanense*. Their structures were elucidated on the basis of spectroscopic evidence. © 2000 Elsevier Science Ltd. All rights reserved.

Keywords: *Dysoxylum hainanense*; Meliaceae; Tirucallanes; 3 β ,22*S*-Dihydroxy-tirucalla-7,24-dien-23-one; 22,23-Epoxy-tirucalla-7-ene-3 β ,24,25-triol; 3 β ,25-Dihydroxy-tirucalla-7,23-diene; 23,26-Dihydroxy-tirucalla-7,24-dien-3-one

1. Introduction

The genus *Dysoxylum* Bl., found in India, Southeast Asia, Australia and New Zealand, comprises about 200 species, 14 of which are distributed in China. About 10 species of this genus have been found in Yunnan province. *D. hainanense* Merr. is distributed in Guangxi Zhuang Autonomous, Hainan province, and Xishuangbanna, Yunnan province (Yunnan Institute of Botany, 1977). In previous reports (Aalbersberg and Singh, 1991; Singh and Aalbersberg, 1992; Govindachari et al., 1994), many dammarane triterpenoids were isolated from plants of this genus. However, no chemical work has been done on *D. hainanense*. As part of a program seeking bioactive compounds from Meliaceae plants (Luo et al., 1999), we examined the extracts of the bark from *D. hainanense*, and isolated four new triterpenoids (**1**)–(**4**), together with two known triterpenoids (**5**) and (**6**). In this paper, we describe the isolation and structural elucidation of these compounds.

2. Results and discussion

The ethanolic extract of the bark from *D. hainanense* was partitioned between H₂O and EtOAc, and the EtOAc-soluble fraction was subjected to repeated silica gel CC to yield six triterpenoids: 3 β ,22*S*-dihydroxy-tirucalla-7,24-dien-23-one (**1**), 22,23-epoxy-tirucalla-7-ene-3 β ,24,25-triol (**2**), 3 β ,25-dihydroxy-tirucalla-7,23-diene (**3**), 23,26-dihydroxy-tirucalla-7,24-dien-3-one (**4**), 24,25-epoxy-3 β ,23-dihydroxy-7-tirucallene (**5**), tirucalla-7,24-diene-3 β ,23-diol (**6**). The known compounds, (**5**) and (**6**), were identified by comparison of their spectroscopic data with those reported previously (Kumar et al., 1991; Mulholland and Taylor, 1988). The structures of the new compounds **1**–**4** were established as follows.

The molecular formula of **1** was assigned as C₃₀H₄₈O₃ by negative ion HR-FABMS. The IR spectrum exhibited absorptions at 3341 cm⁻¹ (OH), 1711 cm⁻¹ (C=O) and 1622 cm⁻¹ (C=C). The ¹H and ¹³C NMR spectra of **1** showed signals for seven tertiary methyls, a secondary methyl, and resonances for four methine carbons [δ_C 50.7 (C-5), 49.1 (C-9), 49.0 (C-17), 39.6 (C-20)], four quaternary carbons [δ_C 51.4 (C-14), 43.4 (C-13), 39.0 (C-4), 35.0 (C-10)], two olefinic carbons [δ_C 118.0 (*d*), 145.7 (*s*)], and one oxymethine

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carbon (δ_C 79.2, δ_H 3.22, *dd*, $J = 10.7$ and 5.3 Hz). These data are consistent with a tirucallane–euphane system having a double bond between C-7 and C-8, and a 3β -hydroxy group (Sherman et al., 1980; Chan et al., 1970; Jolad et al., 1981). In addition, the 1H and ^{13}C NMR spectra of **1** indicated the presence of α,β -unsaturated ketone signals [δ_C 201.4 (*s*), 159.3 (*s*), 119.3 (*d*), δ_H 6.06 (*s*)], and two downfield methyl protons [δ_H 2.19 (*s*), 1.95 (*s*)] attributed to a side chain, suggesting that a double bond was formed between C-24 and C-25, and a ketonic carbonyl group was present at C-23. These structural features were confirmed by cross signals from δ_H 6.06 (H-24) to δ_C 201.4 (C-23), H-24 to δ_C 26.7 (*q*, C-26), and H-24 to δ_C 21.3 (*q*, C-27) in the HMBC spectrum of **1**. In the HMBC spectrum, the resonance at δ_H 4.15 (*s*, H-22) showed cross peaks to δ_C 201.4 (C-23), 39.6 (*d*, C-20) and 12.0 (*q*, C-21), respectively, which revealed a hydroxyl substituent at C-22. The spectral data of the side chain were in good agreement with those of 22*S*-dihydroxy-tirucalla-7,24-dien-3,23-dione (Liang et al., 1988, 1989), the structure of which was confirmed by X-ray diffraction. Thus **1** was determined to be $3\beta,22S$ -dihydroxy-tirucalla-7,24-dien-23-one.

HR-FABMS of compound **2** suggested the molecular formula $C_{30}H_{50}O_4$. The ^{13}C and 1H NMR spectra of **2** were similar to those of **1**, except for the side chain data. A singlet at δ_H 1.22 (6H) was attributed to protons of two methyl groups, suggesting attachment to an oxygenated carbon (C-25). Furthermore, signals for an oxymethine (δ_C 74.1) and an oxirane group (δ_C 60.4, 58.9) were also evident, and were attributed to the side chain. The observation of HMBC cross peaks between δ_H 1.02 (*d*, 6.6, H-21) and δ_C 60.4 (*d*, C-22), δ_H 3.46 (*d*, 8.4, H-24) and δ_C 25.5 (*q*, C-26, C-27), and between the former and 72.8 (*s*, C-25), indicated the side chain of **2** to be 22,23-epoxy-24,25-dihydroxyl substituted. It was presumed that the C-20 configuration belongs to the tirucallane rather than the euphane series, since tirucallane derivatives occur widely in the Meliaceae while euphanes are restricted to *Melia* species (Purushothaman et al., 1985), and in light of the co-occurrence of **2** with compounds **1**, **5** and **6**. Thus **2** was deduced to be 22,23-epoxy-tirucalla-7-ene- $3\beta,24,25$ -triol.

Compound **3** possessed the molecular formula $C_{30}H_{50}O_2$ as determined by HR-FABMS. The 1H and ^{13}C NMR spectra of **3** were similar to those of **2**, with the exception of the side chain resonances. The 1H NMR spectrum showed a singlet for two methyl groups which resonated at δ_H 1.30 (6H), suggesting attachment to an oxygenated carbon (C-25). This was supported by the presence of cross peaks between δ_H 1.30 (6H, H-26, H-27) and δ_C 70.7 (*s*, C-25) in the HMBC spectrum. The HMBC spectrum also exhibited cross signals between an olefinic proton δ_H 5.57 (*d*,

$J = 8.0$ Hz, H-24) and δ_C 70.7 (*s*, C-25), which indicated a double bond between C-23 and C-24. The small coupling constant ($J = 8.0$ Hz) between H-23 and H-24 suggested a *Z*-type olefinic bond. Taking into account the co-occurrence of compounds **1**, **5** and **6** which also possess a tirucallane skeleton, compound **3** was determined to be $3\beta,25$ -dihydroxytirucalla-7,23-diene.

Compound **4** was assigned the molecular formula $C_{30}H_{48}O_3$ by HR-FABMS. The IR spectrum revealed absorptions for a hydroxyl group stretch at 3341 cm^{-1} , a carbonyl group stretch at 1713 cm^{-1} and an olefinic stretch at 1632 cm^{-1} . The 1H and ^{13}C NMR spectra of **4** displayed the presence of six tertiary methyls, one secondary methyl, nine methylenes, characteristic methines at δ_C 53.7, 52.3, 48.4, 33.0, quaternary carbons at δ_C 51.3, 47.8, 43.5, 35.0, four olefinic carbons including two at δ_C 145.8 (*s*) and 117.9 (*d*), and a ketonic carbonyl (δ_C 216.7). These signals were the characteristic of the tirucallane-7-ene system with a 3-ketone (Mulholland and Taylor, 1988; Mondon et al., 1981).

An olefinic linkage was placed between C-24 and C-25 based on the observation that one methyl proton resonance was shifted downfield to δ_H 1.66 and the oxymethylene protons [δ_H 3.95 (2H, H-26)] appeared as a singlet peak in the 1H NMR spectrum of **4**. The assumption was confirmed by the presence of cross peaks between δ_H 5.31 (H-24) and δ_C 67.9 (*t*, C-26), and H-24 and δ_C 14.1 (*q*, C-27) in the HMBC spectrum, in which a correlation between H-24 and δ_C 66.7 (*d*, C-23) also confirmed a hydroxyl group at C-23. The NOE interaction between H-24 (δ_H 5.31) and H-26 (δ_H 3.95) in the NOESY spectrum indicated that the olefinic bond was *E*-type. Compound **4** was deduced to be a tirucallane on the basis of co-occurrence, and was determined to be 23,26-dihydroxy-tirucalla-7,24-dien-3-one.

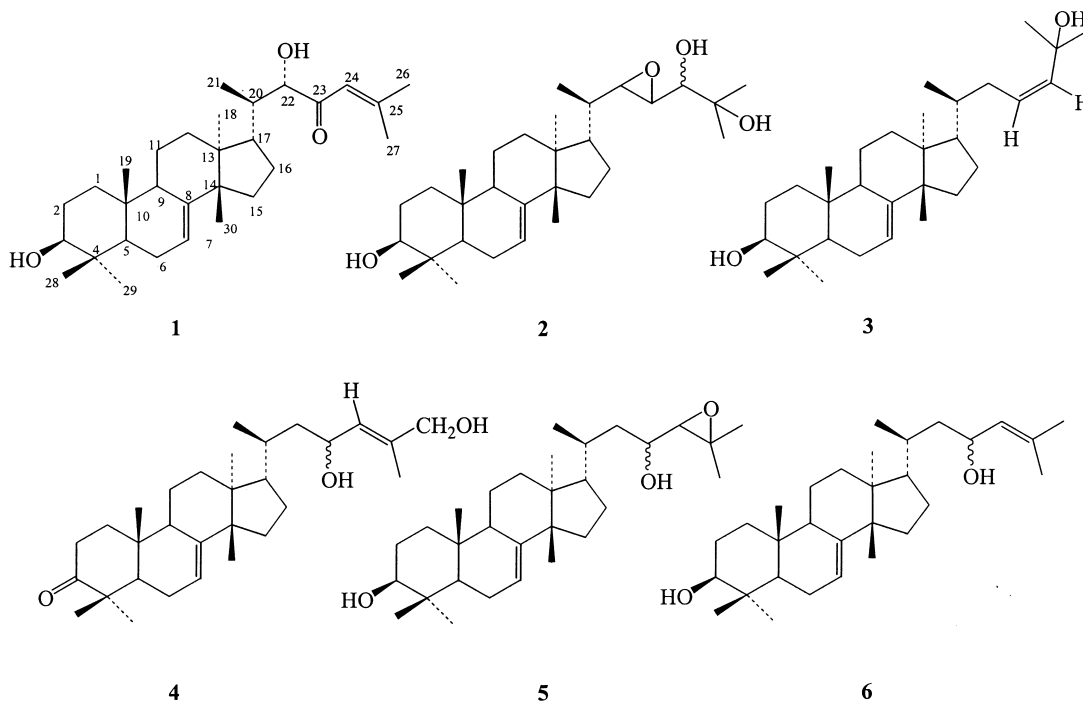
3. Experimental

3.1. General

Mps: uncorrected; UV: MeOH; IR: KBr; 1H NMR, ^{13}C NMR and 2D-NMR spectra were recorded on Bruker AM-400 MHz and a DRX-500 spectrometers with TMS as internal standard and $CDCl_3$ as solvent; chemical shift values (δ) were reported in ppm and coupling constants (J) in Hz; FABMS data were recorded on a VG Autospec-3000 spectrometer; EIMS: 70 eV.

3.2. Plant material

The bark of *D. hainanense* Merr. was collected from



Xishuangbanna, Yunnan province, P. R. China, in December 1996. The plant was identified by Prof. G.-D. Tao of the Xishuangbanna Botany Garden, Academia sinica. A voucher specimen was deposited in the herbarium of the Department of Taxonomy, Kunming Institute of Botany, Academia sinica, Kunming, P. R. China.

3.3. Extraction and isolation

The dried and powdered bark (4.2 kg) of *D. hainanense* was extracted with EtOH under reflux, and the solvent was evaporated in vacuo. The residue was partitioned in H₂O and extracted with EtOAc. The EtOAc extracts were concentrated in vacuo to afford 72 g of residue, which was subjected to silica gel column chromatography, using CHCl₃–Me₂CO (from CHCl₃ to CHCl₃–Me₂CO, 1:1) as eluent. The fractions were monitored by TLC and combined. Fractions 2–5 were further purified on silica gel CC with petrol–Me₂CO (9:1–2:1) as eluent followed by recrystallization to yield **1** (40 mg), **2** (25 mg), **3** (38 mg), **4** (36 mg), **5** (16 mg), **6** (10 mg).

3.4. 3β,22S-Dihydroxy-tirucalla-7,24-dien-23-one (**1**)

C₃₀H₄₈O₃, white powder; mp 80–82°C; [α]_D²⁶ +33.3° (c 0.45, CH₃OH); UV (MeOH) $\lambda_{\max}$ (log ϵ): 240 (3.71) nm; IR (KBr) $\nu_{\max}$: 3448, 2935, 2875, 1711, 1622, 1460, 1384, 1248, 1183, 1107, 1034, 993 and 822 cm⁻¹;

Table 1
¹³C NMR spectral data of compounds 1–6 (CDCl₃, 100 MHz)

C	1	2	3	4	5	6
1	37.2 t	37.2 t	37.2 t	38.4 t	37.2 t	37.3 t
2	27.6 t	27.7 t	27.7 t	34.8 t	27.7 t	27.7 t
3	79.3 d	79.2 d	79.3 d	216.7 s	79.2 d	79.3 d
4	39.0 s	39.0 s	39.0 s	47.8 s	39.0 s	39.0 s
5	50.7 d	50.7 d	50.7 d	52.3 d	50.7 d	50.7 d
6	24.0 t	24.0 t	24.0 t	24.3 t	23.9 t	24.0 t
7	118.0 d	118.3 d	117.9 d	117.9 d	118.1 d	117.9 d
8	145.7 s	145.3 s	145.8 s	145.8 s	145.6 s	145.8 s
9	49.1 d	49.0 d	49.0 d	48.4 d	49.0 d	49.0 d
10	35.0 s	35.0 s	35.0 s	35.0 s	35.0 s	35.0 s
11	18.1 t	18.0 t	18.1 t	18.3 t	18.1 t	18.1 t
12	33.6 t	33.5 t	33.7 t	33.8 t	33.8 t	33.9 t
13	43.4 s	44.0 s	43.6 s	43.5 s	43.6 s	43.6 s
14	51.4 s	50.8 s	51.2 s	51.3 s	51.2 s	51.2 s
15	34.0 t	34.2 t	34.1 t	34.1 t	34.0 t	34.0 t
16	28.1 t	27.3 t	28.1 t	28.2 t	28.7 t	28.5 t
17	49.0 d	50.5 d	52.6 d	53.7 d	53.3 d	53.6 d
18	21.9 q	21.9 q	21.9 q	22.2 q	21.7 q	21.8 q
19	13.1 q	13.1 q	13.1 q	12.7 q	13.1 q	13.1 q
20	39.6 d	38.8 d	36.5 d	33.0 d	33.6 d	33.7 d
21	12.0 q	16.4 q	18.4 q	19.2 q	19.9 q	19.4 q
22	80.7 d	60.4 d	38.9 t	43.2 t	40.8 t	44.4 t
23	201.4 s	58.9 d	125.6 d	66.7 d	69.3 d	67.3 d
24	119.3 d	74.1 d	139.4 d	127.9 d	68.5 d	128.5 d
25	159.3 s	72.8 s	70.7 s	138.2 s	60.3 s	135.6 s
26	26.7 q	25.5 q	29.9 q	67.9 t	24.8 q	25.8 q
27	21.3 q	25.5 q	29.9 q	21.5 q	19.8 q	22.4 q
28	27.2 q	27.5 q	27.6 q	24.5 q	27.6 q	27.6 q
29	14.7 q	14.7 q	14.7 q	14.1 q	14.7 q	14.7 q
30	27.4 q	27.6 q	27.2 q	27.4 q	27.2 q	27.2 q

EIMS m/z (rel. int.): 456 $[M]^-$ (25), 441 (23), 421 (13), 386 (10), 373 (10), 355 (13), 343 (15), 327 (37), 309 (10), 299 (10), 213 (8), 185 (13), 171 (22), 161 (25), 145 (27), 133 (31), 109 (46), 95 (57) and 83 (100); HR-FABMS m/z 455.3531 $[M - H]^-$ (calculated for $C_{30}H_{47}O_3$, 455.3525); 1H NMR spectral data ($CDCl_3$, 400 MHz): δ 6.06 (1H, *s*, H-24), 5.24 (1H, *br s*, H-7), 4.15 (1H, *s*, H-22), 3.22 (1H, *dd*, $J = 9.0$ and 4.1 Hz, H-3), 2.19, 1.95 (each 3H, *s*, H-26, H-27), 1.01 (3H, *s*, H-30), 0.94 (3H, *s*, H-28), 0.83 (6H, *s*, H-29, H-18), 0.72 (3H, *s*, H-19), 0.61 (3H, *d*, $J = 6.6$ Hz, H-21); ^{13}C NMR spectral data: see Table 1.

3.5. 22,23-Epoxy-7-tirucalla-7-ene-3 β ,24,25-triol (2)

$C_{30}H_{50}O_4$, colorless needles (Me_2CO); mp 118–120°C; $[\alpha]_D^{26} -4.7^\circ$ (c 0.95, CH_3OH); IR (KBr) ν_{max} : 3395, 2965, 2933, 2887, 1642, 1463, 1384, 1245, 1170, 1157, 1117, 1066, 1041, 990, 913, 886 and 731 cm^{-1} ; EIMS m/z (rel. int.): 474 $[M]^+$ (40), 459 (10), 442 (15), 413 (10), 395 (12), 383 (10), 342 (27), 327 (25), 299 (17), 255 (17), 227 (15), 213 (16), 187 (30), 173 (36), 161 (84), 147 (49), 133 (57), 119 (60), 105 (67), 81 (63) and 59 (100); HR-FABMS m/z : 473.3541 $[M - H]^-$ (calculated for $C_{30}H_{49}O_4$, 473.3630); 1H NMR ($CDCl_3$, 400 MHz): δ 5.25 (1H, *d*, $J = 3.1$ Hz, H-7), 3.46 (1H, *d*, $J = 8.4$ Hz, H-24), 3.22 (1H, *dd*, $J = 10.7$ and 5.3 Hz, H-3), 3.08 (1H, *d*, $J = 2.2$ Hz, H-22), 2.74 (1H, *dd*, $J = 8.0$ and 2.2 Hz, H-23), 1.22 (6H, *s*, H-26, H-27), 1.02 (3H, *d*, $J = 6.6$ Hz, H-21), 0.97 (3H, *s*, H-30), 0.95 (3H, *s*, H-28), 0.84 (3H, *s*, H-29), 0.79 (3H, *s*, H-18), 0.73 (3H, *s*, H-19); ^{13}C NMR spectral data: see Table 1.

3.6. 3 β ,25-Dihydroxy-tirucalla-7,23-diene (3)

$C_{30}H_{50}O_2$, colorless needles (Me_2CO); mp 168–170°C; $[\alpha]_D^{27} -31.0^\circ$ (c 0.45, $CHCl_3$); IR (KBr) ν_{max} : 3381, 2966, 2931, 2885, 1719, 1461, 1443, 1383, 1245, 1160, 1102, 1035, 971, 918 and 824 cm^{-1} ; EIMS m/z (rel. int.): 442 $[M]^+$ (57), 427 (20), 409 (100), 391 (15), 327 (46), 309 (10), 255 (8), 215 (11), 203 (12), 187 (20), 161 (21), 109 (67), 95 (46), 81 (49), 69 (57); HR-FABMS m/z : 441.3758 $[M - H]^-$ (calculated for $C_{30}H_{49}O_2$, 441.3733) 1H NMR spectral data ($CDCl_3$, 400 MHz): δ 5.57 (1H, *d*, $J = 8.0$ Hz, H-24), 5.55 (1H, *dd*, $J = 8.0$ and 5.5 Hz, H-23), 5.23 (1H, *d*, $J = 3.2$ Hz, H-7), 3.21 (1H, *dd*, $J = 11.2$ and 4.2 Hz, H-3), 1.30 (6H, *s*, H-26, H-27), 0.94 (6H, *s*, H-28, H-30), 0.85 (3H, *s*, H-29), 0.84 (3H, *d*, $J = 5.9$ Hz, H-21), 0.79 (3H, *s*, H-18), 0.72 (3H, *s*, H-19), ^{13}C NMR spectral data: see Table 1.

3.7. 23,26-Dihydroxy-tirucalla-7,24-dien-3-one (4)

$C_{30}H_{48}O_3$, colorless needles (Me_2CO); mp 138–

140°C $[\alpha]_D^{19} -72.5^\circ$ (c 0.29, CH_3OH); IR (KBr) ν_{max} : 3341, 2952, 2861, 1713, 1632, 1462, 1452, 1388, 1371, 1310, 1263, 1013, 1008, 970 and 838 cm^{-1} ; EIMS m/z (rel. int.): 456 $[M]^+$ (33), 438 (55), 423 (90), 405 (15), 380 (12), 340 (28), 325 (100), 313 (45), 271 (47), 245 (35), 203 (28), 189 (34), 173 (48), 161 (56), 147 (60), 135 (65), 119 (72), 107 (75) and 95 (81); HR-FABMS m/z : 455.3600 $[M - H]^-$ (calculated for $C_{30}H_{47}O_3$, 455.3525); 1H NMR spectral data ($CDCl_3$, 400 MHz): δ 5.31 (1H, *d*, $J = 9.1$ Hz, H-24), 5.24 (1H, *d*, $J = 2.7$ Hz, H-7), 4.44 (1H, *dt*, $J = 3.9$ and 9.1 Hz, H-23), 3.95 (2H, *s*, H-26), 2.70 (1H, *dt*, $J = 4.4$ and 14.5 Hz, H-2a), 2.18 (*br d*, $J = 14.1$ Hz, H-2b), 1.66 (3H, *s*, H-27), 1.05 (3H, *s*, H-29), 0.99 (3H, *s*, H-28), 0.94 (3H, *s*, H-30), 0.95 (3H, *s*, H-19), 0.80 (3H, *d*, $J = 5.6$ Hz, H-21), 0.72 (3H, *s*, H-18); ^{13}C NMR spectral data: see Table 1.

3.8. 24,25-Epoxy-3 β ,23-dihydroxy-7-tirucallene (5)

Prisms (Me_2CO); mp 155–157°C; IR (KBr) ν_{max} : 3480, 2953, 2880, 1684, 1460, 1381, 1335, 1302, 1278, 1252, 1163, 1147, 1035 and 990 cm^{-1} ; EIMS m/z (rel. int.): 458 $[M]^+$ (35), 443 (12), 425 (13), 386 (10), 371 (100), 353 (45), 335 (10), 327 (28), 309 (15), 187 (30), 147 (33), 135 (45), 121 (52), 107 (55), 95 (52), 81 (48) and 69 (51); 1H NMR spectral data correspond to those of Mulholland and Taylor (1988); ^{13}C NMR spectral data: see Table 1.

3.9. 3 β ,23-Dihydroxy-tirucalla-7,24-diene (6)

White powder: mp 80–83°C; IR (KBr) ν_{max} : 3445, 2935, 2873, 1720, 1461, 1380, 1252, 1180, 1036, 990, 891 and 549 cm^{-1} ; EIMS m/z (rel. int.): 442 $[M]^+$ (20), 424 (12), 409 (30), 371 (12), 327 (35), 302 (10), 255 (14), 203 (17), 187 (25), 159 (30), 147 (35), 121 (65), 109 (100), 91 (78), 81 (82) and 69 (68); 1H NMR spectral data are in agreement with those of Kumar et al. (1991); ^{13}C NMR spectral data: see Table 1.

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